The Mangrove Strengthen and Ecological Risk Assessment of Mercury Contamination in the Segara Anakan Cilacap Lagoon, Indonesia

by Endang Hilmi

Submission date: 09-Jan-2023 05:59PM (UTC+0700)

Submission ID: 1990158235

File name: 7455-Article Text-59433-67194-18-20230104 submit.docx (4.08M)

Word count: 7539 Character count: 42760 DOI: https://dx.doi.org/10.21123/bsj.2023.7455

The Mangrove Strengthen and Ecological Risk Assessment of Mercury Contamination in the Segara Anakan Cilacap Lagoon, Indonesia

Endang Hilmi^{1*}

Teuku Junaidi²

Arif Mahdiana²

Rose Dewi³

P-ISSN: 2078-8665

E-ISSN: 2411-7986

¹Program of Aquatic Resources Management and Magister SDA program, Faculty of Fisheries and Marine Sciences,

E-mail addresses: teuku.junaidi@unsoed.ac.id , arifmahdiana@gmail.com , rose.83unsoed@gmail.com.

ORCID:0000-0002-2390-5596 (Endang Hilmi)

Received 26/5/2022, Accepted 22/8/2022, Published Online First 20/1/2023



This work is licensed under a Creative Commons Attribution 4.0 International License.

Abstract:

Ecological risk assessment of mercury contaminant has a means to analyze the ecological risk aspect of ecosystem using the potential impact of mercury pollution in soil, water and organism. The ecological risk assessment in a coastal area can be shown by mangrove zonation, clustering and interpolation of mercury accumulation. This research aims to analyze ecological risk assessment of potential mercury (including bioaccumulation and translocation) using indicators of species distribution, clustering, zonation and interpolation of mercury accumulation. The results showed that the Segara Anakan had a high risk of mercury pollution, using indicators like as the potential of mercury contaminant in water body was 0137±0.0137 ppm, substrate and sediment were 0.0134±0.0212 ppm. To reduce the impact of mercury pollution could be conducted by mangrove planting, following the ability of mercury accumulation in stem and bark between 0.011 and 0.064 ppm, in mangrove roots between 10.0260 and 0.0690 ppm and in mangrove leaves between 0.0020 and 0.0120 ppm,. The second indicator of mangrove ability to reduce the impact of mercury contaminant used the indicator of bioaccumulation factors, which had a range between 0.021 and 0.4751, and the translocation factors were between 0.0459 and 1.0547. The results also showed that: Avicennia marina, Sonneratia alba, Rhizophora apiculate, Rhizophora mucronata and Nypa frutican had a good ability to accumulate and reduce the impact of mercury contamination.

Keywords: Bioaccumulation factor, C and mangrove zoning, Ecological risk assessment, Mercury contamination, Translocation factor.

Introduction:

The ecological risk assessment of mercury contaminants is described through the measurement of biotic responses, including mangrove ecosystems that have bioavailability to reduce the impact of metals contaminant and their influence on the aquatic and terrestrial ecosystem 1,2. Basically, the mangrove vegetation has the ability to reduce mercury pollution with absorption, filtering, binding and trapping activities ^{3,4}. The potential mercury contaminants comes from the oil and cement industry, garbage, and household 3,5. The mangrove stand has a specific metabolism system, specific nutrient absorption and specific root activity 3,5,6. In Eastern Segara Anakan, the mangrove stand has a

specific freshwater supply from Sapuregel, Donan and Kembang Kuning Rivers 4, 7-10 and seawater from Samundra, Indian Ocean.

The mangrove ecosystem can be used as a suitable area to support the activity of mercury disposal from industry, transportation and anthropogenic activities 3,4,11. These activities support mercury contaminant in coastal ecosystem and estuary ecosystem 3, 12- 14. The mercury contaminants including (CH₃)-Hg (methyl mercury) waste disposal from the oil refinery petroleum industry, cement industry and laboratories that are characterized as a liquid substance at room temperature 25°C, boils at 365, 68°C and a freezing point of -39°C 15,16,17, which has

Universitas Jenderal Soedirman, Jl. Dr Soeparno, Purwokerto Utara, Banyumas 53122, Central Java, Indonesia. ²Aquatic Resources Management Program, Faculty of Fisheries and Marine Sciences, Universitas Jenderal Soedirman.

Jl. Dr Soeparno, Purwokerto Utara, Banyumas 53122, Central Java, Indonesia.

³Marine Science Program, Faculty of Fisheries and Marine Sciences, Universitas Jenderal Soedirman. Jl. Dr Soeparno, Purwokerto Utara, Banyumas 53122, Central Java, Indonesia.

^{*}Correspondence author: dr.endanghilmi@gmail.com

degradable properties and easily accumulation in water and sediments. Mercury also has high toxicity level ¹⁸, very hazardous properties, and very strong binding properties ^{16, 17, 19-21}. Mercury also has a negative impact on aquatic organisms, causing the organism to be genetically altered, have stunted growth, organ damage and cause death 17,20,22,23 Mercury contaminants also have high risks in fishponds, due to human community activity and coastal stabilization 24.

The ecological risk assessment of mercury contaminant is developed by the distribution, clustering, and interpolation of accumulation activity and translocation activity and are used as an index of ecological risk assessment in the mangrove ecosystem ^{3,17,18,25}. The ecological risk assessment of mercury contaminant can be analyzed by mercury potency in mangrove stem, mangrove roots, and mangrove leaves bioaccumulation of mercury contaminant is an indicator of ecological risk assessment can be analyzed by absorption process, accumulation process, and utilization activity of mercury in a mangrove root and surface area of vegetation 17,28,29. This activity aims to reduce the impact of mercury toxic effect with dilution activity and mercury translocation to dead organs ^{26,30} and organic absorption ^{31,32}. The second indicator is a translocation of mercury contaminants as an activity to transfer contaminants to other organs stem, branches and leaves through cells and the vascular tissue. The translocation process is a passive transport system following the activity of distribution and nutrient absorption ^{26,32,33}.

The ecological risk assessment of the mangrove ecosystem using bioaccumulation and translocation of mercury contaminant give information and data on the adaptation of mangrove vegetation in mercury pollution area. Mangrove vegetations must have the ability to reduce the effect of mercury contaminant 4,19,20. The ecological risk assessment of mercury contaminant also describe the relationship and adaptation of mangrove vegetation in pollution area using the mangrove landscaping, zonation, clustering and association 4,5,9,34. This research aims to analyze the ecological risk assessment of mercury contaminants (including bioaccumulation and translocation) using indicators of distribution, clustering, zonation and interpolation

Materials and Methods: Research area

The research of ecological risk assessment of mercury contaminants was conducted in a waste disposal area in Eastern Segara Anakan (E-SAL) on June -July 2021 and January-March 2022 8,35. The research area could be shown in Fig.1 and Table. 1. The area of waste disposal in the mangrove ecosystem was dominated by Rhizophora apiculata, Rhizophora mucronata, Rhizophora styllosa, Bruguiera gymnorrhiza, Sonneratia caseolaris and Avicennia marina ^{8,10,34,36}. The sampling of mercury contaminants in the mangrove ecosystem can be conducted in Kalipanas River (Station 1), the Sleko Port (Station 2), Pertamina /oil refinery Area (Station 3), the Cement Plant (Station 4), and East Pelawangan/estuary. (Station 5).

Table 1. Research area and stations

	Table 1. Research and stations							
No	Research stations	The coordinates						
		Latitude (South)	Longitude (East)					
1.	Kalipanas River	07°42'36,60"	108°59'43,91"					
2.	The Sleko Port	07°43'17,11"	108°59'31,00"					
3.	Pertamina Area/oil refinery	07°41'48,64"	108°59'34,98"					
4.	Cement Plant	07°40′59,81"	109°00'40,35"					
5.	East Pelawangan/estuary	07°43'40.87"	108°59'03,31"					

The number of sampling plots to analyze mercury contaminant in sediment and water was 15 sampling plots (3 sampling plots/stations). Whereas the number of sampling for mangroves (collecting roots, barks, stems, and leaves) from 15 sampling

plots were 75 individual samples (5 samples of vegetation/mangrove species) 37,38. The samples total from part of mangrove tree to analysis heavy metal accumulation were 225 samples

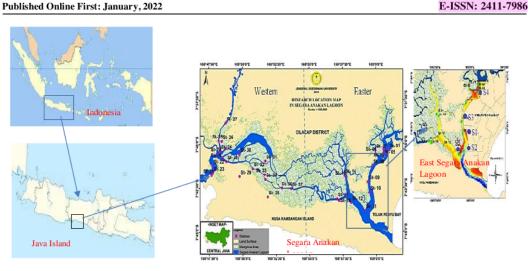


Figure 1. Research area

The sampling of mercury contaminant

a. The sampling in the water body and substrate's The sampling in water bodies and substrates could be conducted by the collection of 600 mL of water samples and be placed and labeled into a bottle. The water samples be added by 0.75 mL concentrated HNO₃ until the pH until two ^{3,21,39}. Substrate samples were collected 250 g using Eckman grab until 50-100 cm form the bottom. The substrate samples were placed and labeled into the plastic bag ^{3,11,21}.

The sampling of mangrove vegetation

Mangrove vegetation was collected by sampling 150 – 350 g. The samples (bark, stem, leaves and roots) were collected by destructive methods and then materials were extracted. Specifically the mangrove roots were collected from actual roots beneath the sediment including the respiratory roots. The mangrove samples were collected, labeled and placed into plastic bags and the plastic bags were put into an icebox ^{3.26}.

Mercury analysis

The mercury accumulation from mangrove leaves, stems and roots were analyzed by a spectrophotometric method using Shimatsu® the accuracy level is 2). x10⁻⁴ pmBefore the analysis of accumulation mercury using spectrophotometric, the mangrove samples were extracted by a filtrate system using the mixed system of 10 ml H₂SO₄, 2 ml KMnO₄ 2%, 1 ml K₂S 2O₈, and 1 ml stannous chloride SnCl₂, 10% were extracted system using tetra dithizone liquid. Hg was measured by mercury analyzer (SP-3D) method with a wavelength of 480 nm. This method uses: Hg $^{2+}$ + SnCl₂ \rightarrow HgO and then uses the Hg Detector analyzer 40.

P-ISSN: 2078-8665

The bioaccumulation factor (BAF) of mercury contaminant

The Bioaccumulation factor (BAF) of mercury contaminant was analyzed by the equation of 3,4,41 .

$${\rm BAF} = \frac{{\rm mercury}\ {\rm accumulation\ of\ mangrove\ leaves, roots\ and\ stem\ (mg\ kg^{-1})}}{{\rm mercury\ accumulation\ of\ mangrove\ substrates\ (mg\ kg^{-1})}}$$

Bioaccumulation Factor (BAF) had categories were

BAF ≤1 describe low or unable activity to accumulate mercury pollution

BAF >1 describe high ability to accumulate mercury pollution.

The Translocation factor (TF) of mercury contaminant

The translocation factor (TF) was analyzed by the equation of 3,4,41 .

mercury accumulation of mangrove leaves, roots and stem (mg kg^{-1})

mercury accumulation of mangrove roots (mg kg^{-1})

Translocation Factor (TF) had categories were 3,24,42 TF \leq 1 describe low or unable activity of translocate mercury pollution to other organs

TF > 1 describe to good activity to translocate mercury pollution to other organs

The clustering of mangrove vegetation using indicator mercury contaminant accumulation

The clustering of mangrove vegetation using indicator mercury contaminant accumulation used Euclidian distance analysis based on dissimilarity accumulation 8,43,44.

Euidian distance
$$_{jk} = \sqrt{\sum_{i=1}^{s} (xij - xik)^2}$$

P-ISSN: 2078-8665

E-ISSN: 2411-7986

Stage 2.

$D(j,k)h = \alpha_1 D(j,h) + \alpha_2 D(k,h) + \beta D(j,k)$									
Stations	2	3	4		22				
1	EuDi ₁₂	EuDi13	EuDi14						
2		EuDi23	EuDi24						
3			EuDi34						
22			EuDi22						
	1								

Notes 44,8:

β

EuDijk : Euclidean Distance of mercury accumulation

: species i

: mercury accumulation of species- j Xij: mercury accumulation of species - k Xik: Distance between potency of

mercury accumulation : 0.625 α1 $\alpha 2$: 0.625 : - 0.25 44,8

The interpolation analysis mercury accumulation

The interpolation analysis of mercury contaminant accumulation was conducted by mapping analysis. The mapping analysis used the combined approach among sampling data, Landsat data, NDVI and NDWI method, and interpolation tool in ArcGIS software 45,46.

The landscaping of mangrove vegetation

The landscaping of mangrove vegetation using the data of mercury contaminant accumulation based on BAF and TF scores. The landscape of mangrove vegetation showed the zonation of mangrove species following the score of mercury accumulation 13,19.

Results and Discussion

The ecological risk assessment of the mangrove ecosystem based on the potential for mercury

The ecological risk assessment of Segara Anakan Lagoon is influenced by mercury contaminants in water and sediments coming from sea water treatment of oil refinery industry, aquaculture pesticides, domestic pollution, charcoal industry and cement industry 31. The potential for mercury contaminations gives a negative impact on the environment, organisms and the local human community ^{12,18,47} (Fig.2).

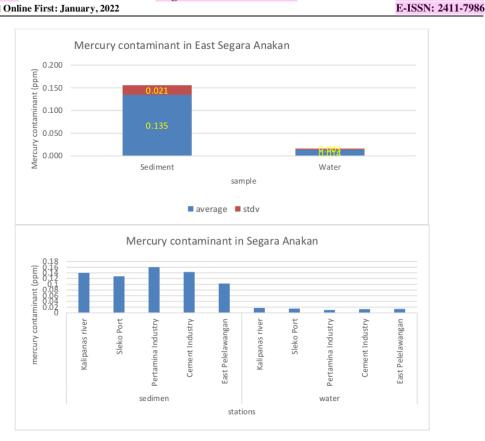


Figure 2. The potential of mercury contaminant on sediment and water body in Segara Anakan Lagoon.

Fig. 2 shows the potential of mercury contaminant in sediment 0.135±0.0021 ppm > potential in water 0.014±0.003 ppm. Based on the Government Regulation of the Indonesia Republic, Number 101/2014 and Number 82/2001 and the data potential mercury in sediment and water noted that Segara Anakan lagoon was polluted. 48 indicated that mercury concentration in the coastal sediment in Buyat Bay had potential up to 7 mg kg $^{-1}$, and 33 also indicated that mercury contaminant in sediments of the mangroves ecosystem have ranges SJM 414.50 ng g $^{-1}$ > XXM 272.30 ng g $^{-1}$, FTM 216.47 ng g $^{-1}$ > BGM 80.91 ng g $^{-1}$; SJM 356.25 ng g $^{-1}$ > XXM 234.57 ng g $^{-1}$, FTM 197.23 ng g $^{-1}$ > BGM 65.35 ng g $^{-1}$.

Basically¹⁴, also indicated 90% of mercury contaminants are deposited in sediments because the heavy metal contaminant was easy to bond and deposited in sediments ^{49,50}. Whereas based on the distribution in sediments from every station showed that the mercury contaminant in oil refinery station (pertamina stations) > cement industry > Kalipanas

River > Sleko port > east of Pelawangan (estuary station), then the mercury contaminant of water body in Kalipana Rivers > Sleko Port > oil refinery Industry, cement industry and East of Pelawangan. The pollution category of Government Regulation of the Indonesia Republic, Number 82 (2001) explains that the Segara Anakan Lagoon was polluted with mercury contaminant. Otherwise, 31 only found that Zn, Cd and Pb contaminated the Red Sea coast of Egypt are 14.94 - 134.22 □g/g (Zn), $3.17-40.25 \square g/g$ (Pb) and $0.12-1.25 \square g/g$ (Cd)²⁶. also reported the potential contamination in sediments show that potential Cd between 0.15-1.62 mg/g < Pb 1.36-6.28 mg/g < Ni 17.9-24.3 mg/g < Cu 9.27-36.47 mg/g < Cr 27.68 -84.62 mg/g. According to 51 reports that Busan city has the potential for contaminant $Zn \le Pb < Cu < Cr \le As$ < Ni ≤ Cd < Hg. This data is not different from 52 in China's Hainan and Zhoushan coastal areas. 18 using the PCA analysis show that potential for contamination by Ni, Cr, Cu, As, Hg and Zn from

P-ISSN: 2078-8665

natural sources and Cd and Pb from anthropogenic source.

The accumulation of mercury contaminants in water and sediments also is influenced by the following environmental factors: dissolved oxygen (DO); chemical oxygen demand (COD); biological oxygen demand (BOD); total suspended solids (TSS); pH; conductivity; ammonium (NH4⁺-N);

nitrate (NO3⁻-N); Kjeldahl nitrogen; and total phosphorus ^{13,53}. The data also showed that salinity was 16 PSU – 25.7 PSU, pH 5.7 – 7.1, COD 22.9 ppm – 41.5 ppm, sediment salinity was 19.7 – 23.7 PSU and sediment pH was 5.3 – 5.8 (Table. 2). Based on data COD showed that Pertamina industry is designated as a polluted area (COD > 25 ppm)

Table 2. Environmental factors affecting mercury contamination in the area of Segara Anakan

Cilacap								
Stations	Tools	Water			Sediments			
		Salinity (PSU)	pН	COD (ppm)	Salinity (PSU)	pН		
Kalipanas	Average	25.0	6.6	22.9	23.7	5.8		
rivers	Standard deviation	0.1	0.1	1.6	0.6	0.1		
Sleko port	Average	24.3	7.1	26.5	20.3	5.6		
	Standard deviation	0.6	0.2	0.7	0.6	0.1		
Pertamina	Average	21.3	5.7	41.5	22.0	5.8		
area	Standard deviation	0.6	0.1	7.2	1.0	0.1		
Cement	Average	16.3	6.6	32.9	22.7	5.6		
industry	Standard deviation	0.6	0.2	4.4	0.6	0.1		
East	Average	25.7	6.8	27.1	19.7	5.3		
Pelalawang	Standard deviation	0.6	0.1	1.6	0.6	0.2		

Based on COD, salinity and pH mangrove have sensitive characteristics since they can be influenced by the potential for mercury contamination and other pollutants ^{19,26}. To reduce the impact of contamination, salinity, pH and potential COD, mangroves must have highly adaptative using activities of the excretion gland, exclusion gland and accumulation gland Waste disposal from the cement industry and oil refinery are the major source of mercury contamination and mercury easily accumulates through a binding and deposition process of organic matter 6,38. However, the mercury accumulation within the East Segara Anakan Lagoon sediments is still lower than the US EPA standard (< 0.2 mg/Kg). But based the Government Decree No. 82 (2001) and the Decision of the State Minister of the Environment No. 51 (2004) showed that mercury contamination in this lagoon was polluted since the potential for mercury contamination > the mercury standards for aquatic organisms mercury > 0.001 mg/L. The mercury accumulation in this lagoon also is distributed by tidal currents and water inundation 13,19,56. In rivers, mangrove stands and lagoon ecosystems in Segara Anakan as semiclsoed

estuary give a specific distribution of mercury accumulation.

The ecological risk assessment of mangrove stands base on potential for mercury contaminantion

Potential of mercury accumulation in mangrove stands

The ecological risk assessment of mangrove stands using the distribution of mercury accumulation in Segara Anakan Lagoon was shown in Table. 3 and Fig.3. Table. 3, describes that potential accumulation of mercury contamination in the mangrove stem had a range of 0.0110 - 00640 ppm, mangrove leaves ranged 0.0020-0.0120 ppm, and mangrove roots ranged 0.0260-0.0690 ppm. Based on the species distribution Avicennia marina, Sonneratia alba, Rhizophora apiculate, Rhizophora mucronata and Nypa frutican, had a high ability to accumulate mercury contaminants. According to 1 Aviccenia marina had a good ability to accumulate Fe (2892.83 -2902,83 ppm), Mn (2.53-127.3 ppm), Cu (27.84 -60.81 ppm), and Ni (15.55-78.85 ppm). The potential mercury accumulation of mangrove stands in Segara Anakan is relatively different than ¹⁸ and ¹⁹, which reported that the potential mercury contaminant in Lumnitzera racemose

P-ISSN: 2078-8665 E-ISSN: 2411-7986

approximately 0.52 μ g g-1, and ²⁶ also indicated that Avicennia marina had the ability to accumulate Cr > Cu > Ni > Pb > Cd ^{19,57}.

The accumulation of mercury contaminants in mangrove roots, stems, and leaves had higher potency than in water but was still smaller than the mercury accumulation in sediments. The potential of mercury accumulation has a correlation with the ability to absorption, accumulation and extract of mercury from water and sediments. These activities are following the activity of nutrient absorption and metabolic process to support mangrove growth ^{19,29}. The absorption, transferring and translocating activity of mangrove roots to other parts of the tree influence the rate of mangrove growth ^{26, 32}. The highest potential of mercury accumulation was

influenced by root activity as direct contact and nutrient absorption from water column and sediment ^{19,31}, which are translocated to other parts ^{3,26,33}. Similarly ⁵⁸, reported that potential concentration ion of roots still is higher than stem, branches and leaves. Mangrove roots have to metabolize to avoid excessive mercury input and have the ability to reduce mercury contamination to support mangrove growth. The mercury absorption by the roots is influenced by the mangrove roots system and potential of lentic 1 size ^{21,28}, because the mangrove roots have the function as a direct contact and nutrient absorber, which is followed by mercury absorption from sediment and water column ^{19,31} and then translocated to other parts ^{3,26,33}

Table 3. The mercury accumulation distribution of mangrove species

mangrove species	Hg accumaltion (ppm)							
	Mangrove stem	Mangrove leaves	Mangrove roots					
Aegiceras corniculatum	0.0200-0.0260	0.0040-0.0060	0.0468-0.0500					
Aegiceras floridum	0.0201-0.0210	0.0060-0.0080	0.0468-0.0501					
Avicennia marina	0.0220-0.0520	0.0090-0.0160	0.0270-0.0670					
Bruguiera gymnorrhiza	0.0300-0.0370	0.0030-0.0040	0.0436-0.0502					
Bruguiera sexanggula.	0.0180-0.0187	0.0020-0.0030	0.0436-0.0500					
Ceriops tagal	0.0130-0.0150	0.0021-0.0030	0.0438-0.0505					
Excoecaria agallocha	0.0118-0.0120	0.0040-0.0050	0.0451-0.0507					
Hibistus tiliaceus	0.0110-0.0118	0.0030-0.0050	0.0436-0.0501					
Melaluca leucadendron	0.0170-0.0172	0.0070-0.0090	0.0436-0.0505					
Nypa frutican.	0.0405-0.0450	0.0070-0.0090	0.0427-0.0440					
Rhizophora apiculata	0.0120-0.0240	0.0080-0-0090	0.0260-0.0590					
Rhizophora mucronata	0.0150-0.2300	0.0030-0.0040	0.0460-0.0690					
Rhizophora stylosa	0.0150-0.0180	0.0020-0.0040	0.0436-0.0500					
Sonneratia alba	0.0250-0.0640	0.0090-0.0120	0.0427-0.0440					
Xylocarpus granatum	0.0200-0.0260	0.0030-0.0040	0.0418-0.0422					

In other conditions, mangrove species still must have the ability to reduce the impact of mercury pollution, mangroves must have a toxic mechanism for mercury alleviation, mercury dilution and mercury translocation mechanism and

must have the ability to increase absorption of organic matter ^{31,32}. Mercury contamination will have an increasing proline and malonaldehyde contents, glutathione, non-protein thiols, inhibit the photosynthetic pigment and phytochelatins ^{20,54}.

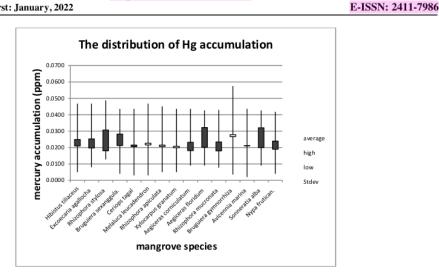


Figure 3. Distribution of mangrove species to accumulate mercury contaminant

The distribution of mangrove species to accumulate mercury contaminant in Fig. 4 explained that the average mercury accumulation > Stdev. The data showed that mercury accumulation of mangrove species had ranges 0.020-0.032 mg/L with an average accumulation 0.025 mg/L and stuff accumulation 0.045 mg/L. The ability of Avicennia marina, Sonneratia alba, Nypa frutican and Rhizophora apiculata, to accumulate mercury contaminants without harm, support these species as the best to rehabilitate in Segara Anakan Lagoon $^{3.8,9.59}$, due to their good respiratory system and spreading root systems 34 to grow in mercury contamination area $^{60.61}$.

The Bioaccumalation factor (BAF) and the Translocation Factor (TF) of mercury contaminant in a mangrove stand

The bioaccumulation factor and the translocation factor of mercury accumulation were shown in Table. 4 and Fig. 4. The data shows that

the BAF of mercury concentrations in the mangrove stem was Sonneratia alba > Nypa frutican > Bruguiera gymnorrhiza > Melaluca leucadendron > Avicennia marina > other mangrove species. BAF of mercury concentrations in the mangrove leaves shows that Avicennia marina > Sonneratia alba > Nypa frutican > Aegiceras floridum > other mangrove species. And BAF of mercury concentrations in the mangrove roots shows that Ceriops tagal > Rhizophora mucronate > Hibiscus tiliaceus > other mangrove species. The potential BAF of mercury concentrations in mangrove stem had ranged between 0.1259 and 0.3262 BAF of mercury concentrations leaves between 0.0156-0.0904 and BAF of mercury concentrations in roots ranges between 0.2984 and 0.4338. This data is different from 26 that reported a BAF of mercury concentrations in mangrove leaves for Cr (0.43), Cu (0.88), Ni (0.47), Pb (1.57), and Cd (0.39). And BAF of areal roots were Cr (0.47), Cu (0.59), Ni (0,49), Pb (1.60) and Cd (0.23)

P-ISSN: 2078-8665

Table 4. The Bioaccumulation factor (BAF) and The Translocation Factors (TF) of mercury

contaminant in mangrove stands										
mangrove species	BAF						TF	F		
	stem	stdev	leaves	stdev	root	stdev	stem	stdev	leaves	stdev
Aegiceras corniculatum	0.1715	0.0447	0.0366	0.0076	0.3463	0.0271	0.8240	0.2163	0.5993	0.2124
Aegiceras floridum	0.1641	0.0171	0.0625	0.0145	0.3654	0.0271	0.7856	0.0810	0.5826	0.0944
Avicennia marina	0.2199	0.1243	0.0904	0.0253	0.3486	0.1471	0.7804	0.0496	0.5211	0.1378
Bruguiera gymnoriza	0.2643	0.0979	0.0286	0.0054	0.3114	0.0046	0.7511	0.0876	0.4645	0.1215
Bruguiera sexanggula.	0.1259	0.0779	0.0210	0.0146	0.3049	0.0154	0.8750	0.0856	0.6363	0.1195

Ceriops tagal

Baghdad Science Journal

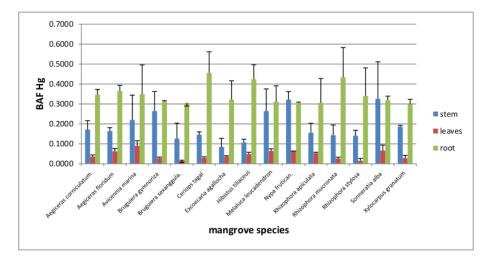
E-ISSN: 2411-7986 0.1460 0.0142 0.0292 0.0058 0.4552 0.1063 0.9445 0.0491 0.6313 0.0035

P-ISSN: 2078-8665

- I - G										
Excoecaria	0.0843	0.0436	0.0357	0.0046	0.3221	0.0941	0.7090	0.1665	0.4519	0.1269
agallocha										
Hibistus tiliaceus	0.1071	0.0161	0.0487	0.0092	0.4243	0.0723	0.9685	0.1835	0.4942	0.0299
Melaluca	0.2643	0.1112	0.0643	0.0110	0.3114	0.0798	1.2339	0.1877	1.0627	0.4019
leucadendron										
Nypa frutican.	0.3214	0.0404	0.0643	0.0000	0.3048	0.0047	0.3419	0.6308	0.2358	0.5847
Rhizophora	0.1559	0.0473	0.0530	0.0048	0.3057	0.1224	0.7525	0.3165	0.5520	0.2514
apiculata										
Rhizophora	0.1434	0.0513	0.0263	0.0069	0.4338	0.1488	0.8415	0.0314	0.5954	0.0486
mucronata										
Rhizophora	0.1406	0.0276	0.0156	0.0110	0.3406	0.1403	1.0043	0.1309	0.7836	0.1087
stylosa										
Sonneratia alba	0.3262	0.1851	0.0663	0.0275	0.3190	0.0202	0.5728	0.0481	0.4141	0.0077
Xylocarpus	0.1857	0.0068	0.0286	0.0129	0.2984	0.0247	1.3057	0.4942	1.0716	0.4610
granatum										
							1			

The translocation factor (TF) of mercury contaminant in mangrove vegetation between 0.578-1.3057 (mangrove stem) and 0.2358-1.0716 (mangrove leaves). The species distribution of translocation factor showed that Xylocarpus granatum > Melaluca Leucadendron > Rhizophora stylosa > Hibiscus tiliaceus > ceriops tagal > other mangrove species (Mangrove stem) and Xylocarpus granatum > Melaluca leucadendron > Rhizophora stylosa > other mangrove species (Mangrove leaves) According ²⁶ reports that the translocation factor in aerial roots are Cd (2.72) > Cu (1.74) > Ni (1.42) > Pb (1.29) > Cr (0.90).

The accumulation process of mercury phytoextraction contaminant influenced by the absorption ability of mercury process as contaminant from waterbody or substrate through mangrove roots stored in leaves plant 39,62 Phytovolatilization Process as the absorption of mercury contaminant using evaporative process and transpired by mangrove leaves phytodegradation or phytotransformation process as they absorb and destroy the activity of mercury contaminant enzymes metabolism or compounds, phytostabilization process as transforming process of mercury contaminant become compounds 63,65,66 and rhizofiltration process as the pollutant absorbing process by mangrove root 63,67,68. Whereas the Translocation Factor (TF) shows the mercury transfer and translocation process from root to leaf and another organ 3, 17, 19, ³³. TF also show transport process and increase in mercury accumulation ^{19, 26, 3, 46} The data also showed that mangrove had a good ability to accumulate mercury contaminant from substrate or sediment, but must have high adaptation to grow and live in mercury pollution 17, 19,68



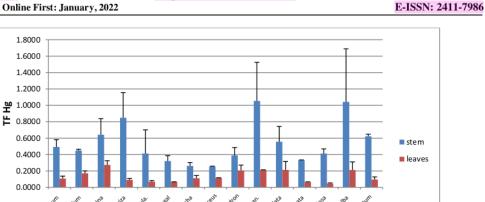


Figure 4. Bioaccumulation factor (BAF) and Translocation Factors (TF) of mangrove vegetation

mangrove species

The mapping interpolation of ecological risk assessment of mercury contaminant in the mangrove ecosystem

The interpolation mapping of mercury contaminants as a model of ecological risk

assessment in mangrove ecosystems was developed by the potential mercury accumulation in mangrove stands, sediments and water. The interpolation mapping of ecological risk of mercury contamination could be shown in Fig.5.

P-ISSN: 2078-8665

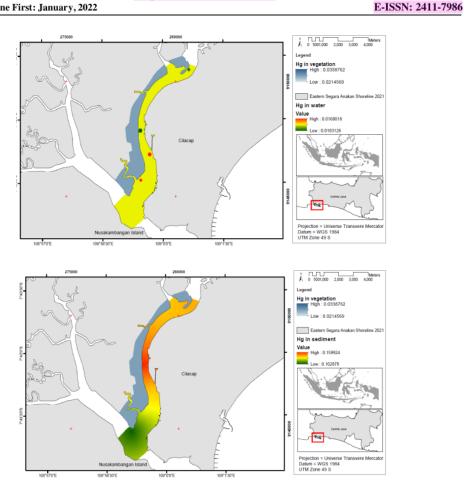


Figure 5. Interpolation of heavy metal contaminant distribution

The interpolation mapping in Fig.5 shows mercury accumulation in stands and water < mercury accumulation in stands and sediment. The potential mercury accumulation can be categorized as moderate to high potential. The interpolation mapping of mercury contamination also shows the critical and toxicity of mercury in vegetation, sediment and water. The water water water water are sponse of phenolic metabolism to reduce the impact of heavy metals in mangroves, including mercury. The mercury contaminant both of a single element or mercury in a compound has high toxicity for many organisms 16.69. According to 21, the concentration of mercury in the environment must be lower than 0.2 mg/Kg, because if more than the standard accumulation, the mercury will have a high toxicity impact. The mercury toxicity symptoms of trees, in general, are reducing membranes of root

cells, growth limitation, chlorophyll damage leading to low photosynthesis, limitation of respiration, can interference with uptake of metabolic of water, disturbance nutrients absorption, and disturbance chlorophyll synthesis ^{26,29}.

P-ISSN: 2078-8665

The mangrove landscaping to reduce the ecological risk of mercury contaminant

The mangrove landscape was developed to reduce the potential for mercury pollution by zonation of mercury accumulation ability (Fig.6). The ecological risk assessment with the mangrove landscape describes the pattern of mangrove zoning based on the accumulation and reduced ability of mercury contamination and can be used as an adaption pattern and model of mangrove species to grow to live in mercury polluted areas.

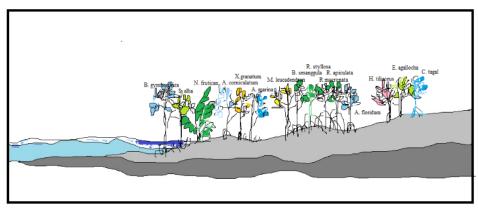


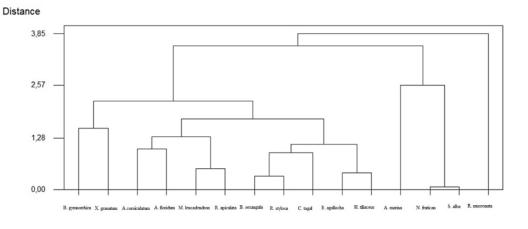
Figure 6. The mangrove landscaping uses the indicator of the mercury accumulation

The mangrove landscape is the model and pattern of ecological risk assessment to reduce mercury contamination showed that the first zone was dominated by Sonneratia alba, Bruguiera gymnorrhiza, Nypa frutican, the second zone was dominated by Aegiceras corniculatum, Xylocarpus granatum and Avicennia marina the third zone was dominated by Melaleuca Leucadendron, Bruguiera sexanggula, Aegiceras floridum, Rhizophora mucronata, Rhizophora stylosa and Rhizophora apiculata, the last zone was dominated by Hibiscus tilaceus, Excoecaria agallocha and Ceriops tagal. The mangrove landscape to reduce mercury contaminantion is influenced by the ability to reduce mercury contaminantion with activities of phytostabilization, phytoextraction, phytodegradation or phytotransformation, phytovolatilization, and rhizofiltration 4, 22. The mangrove landscaping also protects the marine and coastal ecosystems and reduces the impact toxic of mercury with the dilution process and translocation process 3, 19, 69

The ecological risk assessment uses the clustering of mangrove species to accumulate mercury contamination

The clustering of mangrove species in the contamination area was used to describe the ecological risk of mercury contamination as shown in Fig.7. The mangrove species clustering refers to a grouping of mangrove species following the absorption and accumulation ability of mercury contamination 8, 9 using the Hierarchical and Nonhierarchical Clustering Methods ^{8, 36, 43, 70}. The clustering of mercury accumulation in mangrove species shows that Sonneratia alba, Nypa frutican and Avicennia marina (Group 1); Bruguiera sexangula, Rhizophora stylosa, Ceriops tagal, Excoecaria agallocha, Hibiscus tiliaceus (Group 2); Aegiceras floridum, Aegiceras corniculatum, Melalauca Leucadendron, Rhizophora apiculata (Group 3); Bruguiera gymnorrhiza and xylocarpus granatum (Group 4) and Rhizophora mucronata as single species (Group 5)





Mangrove species

Figure 7. Ecological Risk Assessment using indicator of species mangrove clustering in mercury contaminant area

According to 12, 18, 50, 71 clustering of heavy metals including mercury is influenced by water, inundation, environmental condition. pollution sources and substrate. The results show that clustering of mangrove stands to reduce mercury contaminantion is relatively different from mangrove zonantion, except Group 1, which is dominated by Sonneratia alba and Sonneratia alba.

Conclusion

The ecological risk assessment of mercury contaminant in the Segara Anakan Lagoon had characteristics are potential contamination in sediments (0.135±0.0021 ppm) and in water (0.014±0.003 ppm). The second indicator is the potential accumulation of mercury contamination are 0.0110 - 00640 ppm (mangrove stem), $0.\overline{0020}$ -0.0120 ppm (mangrove leaves), and 0.0260-0.0690 mm (mangrove roots). The third indicator is Avicennia marina, Sonneratia alba, Rhizophora apiculate, Rhizophora mucronata and Nypa frutican, which had a good ability to accumulate mercury contaminants. The forth indicator is the mangrove landscape that reduces mercury contaminantion with the first zone dominated by Bruguiera gymnorrhiza, Sonneratia alba, Nypa frutican, the second zone dominated by Aegiceras corniculatum, Xylocarpus granatum and Avicennia marina the third zone dominated by Melaleuca Leucadendron, Bruguiera sexanggula, Aegiceras floridum, Rhizophora mucronata, Rhizophora stylos and Rhizophora apiculata, and the last zone was dominated by Hibiscus tilaceus, Excoecaria

agallocha and Ceriops tagal. The last conclusion, the author would like thanks to LPPM Unsoed for the Institutional Research Grant 2022 (RISIN 2022) and also thanks for constructive reviews, journal editors for their cooperation in supporting the publication of this journal

Author's declaration:

- -Conflicts of Interest: None.
- -I hereby confirm that all the Figures and Tables in the manuscript are ours. Besides, the Figures and images, which are not ours, have been given permission for re-publication attached with the manuscript.
- -Ethical Clearance: The project was approved by the local ethical committee at Jenderal Soedirman University.

Authors' contributions statement

Endang Hilmi made the protocol of the study. Teuku Junaidi, Arif Mahdiana and Rose Dewi supported the study. All authors (Endang Hilmi, Teuku Junaidi. Arif Mahdiana. Dewi) interpreted the data, and read the manuscript carefully and approved the final version of their manuscript.

References

1. Aljahdali MO, Alhassan AB. Ecological risk assessment of heavy metal contamination in mangrove habitats, using biochemical markers and pollution indices: A case study of Avicennia marina L. in the Rabigh lagoon, Red Sea. Saudi J Biol Sci. 2020; 27(4): 1174–1184.

P-ISSN: 2078-8665 E-ISSN: 2411-7986

- Al-Akeel KA, Al-Fredan MA, Desoky ESM. Impact of wastewater discharge on the plant diversity, community structure and heavy metal pollution of range plants in eastern Saudi Arabia. Saudi J Biol Sci. 2021; 28(12): 7367–7372.
- Hilmi E, Siregar AS, Syakti AD. Lead (Pb) distribution on soil, water and mangrove vegetation matrices in Eastern Part of Segara Anakan Lagoon, Cilacap. Omni-Akuatika. 2017; 13(2): 25–38.
- Syakti AD, Ahmed MM, Hidayati N V, Hilmi E, Sulystyo I, Piram A, et al. Screening of Emerging Pollutants in the Mangrove of Segara Anakan Nature Reserve, Indonesia. IERI Procedia. 2013; 5: 216–222.
- Hilmi E, Pareng R, Vikaliana R, Kusmana C, Iskandar I, Sari LK, et al. The carbon conservation of mangrove ecosystem applied REDD program. Reg Stud Mar Sci. 2017; 16: 152–161.
- Wolswijk G, Satyanarayana B, Dung LQ, Siau YF, Ali AN Bin, Saliu IS, et al. Distribution of mercury in sediments, plant and animal tissues in Matang Mangrove Forest Reserve, Malaysia. J Hazard Mater. 2020; 387(June): 121665 (10p)
- Hilmi E. Mangrove landscaping using the modulus of elasticity and rupture properties to reduce coastal disaster risk. Ocean Coast Manag. 2018; 165(July): 71–79
- Hilmi E, Sari LK, Cahyo TN, Muslih M, Mahdiana A, Samudra SR. The affinity of mangrove species using association and cluster index in north coast of jakarta and segara anakan of cilacap, indonesia. Biodiversitas . 2021; 22(7): 2907–2918. https://doi.org/10.13057/biodiv/d220743
- Hilmi E, Amron A, Sari LK, Cahyo TN, Siregar AS.
 The Mangrove Landscape and Zonation following Soil Properties and Water Inundation Distribution in Segara Anakan Cilacap. J Man Hut Trop. 2021; 27(3): 152–164.
- Hilmi E, Sari LK, Siregar AS, Sulistyo I, Mahdiana A, Junaidi T, et al. Tannins in mangrove plants in segara anakan lagoon, central java, indonesia. Biodiversitas. 2021; 22(8): 3508–3516.
- Prastyo Y, Batu DT. L, Sulistiono S. Heavy Metal Contain Cu and Cd on the Mullet in the estuary of Donan River, Cilacap, Central Java. Jumal Pengolahan Hasil Perikanan Indonesia. 2017; 20(1): 18-28
- Cao Z, Wang L, Yang L, Yu J, Lv J, Meng M, et al. Heavy metal pollution and the risk from tidal flat reclamation in coastal areas of Jiangsu, China. Mar Pollut Bull. 2020; 158: 111427 (11p).
- Kibria G, Hossain MM, Mallick D, Lau TC, Wu R. Trace/heavy metal pollution monitoring in estuary and coastal area of Bay of Bengal, Bangladesh and implicated impacts. Mar Pollut Bull. 2016; 105(1): 303, 402
- 14. Xin K, Huang X, Hu J, Li C, Yang X, Arndt SK. Land use Change Impacts on Heavy Metal Sedimentation in Mangrove Land use Change ILand use Change Impacts on HeavyMetal Sedimentation in Mangrove Wetlands—A Case Study in Dongzhai Harbor of Hainan, China. Wetlands. 2014; 34: 1–8.

- Lei P, Zhong H, Duan D, Pan K. A review on mercury biogeochemistry in mangrove sediments: Hotspots of methylmercury production? Sci. Total Environ. 2019; 680: 140–150.
- Li R, Chai M, Guo M, Qiu GY. Sediment accumulation and mercury (Hg) flux in Avicennia marina forest of Deep Bay, China. Estuar. Coast Shelf Sci. 2016; 177: 41–46.
- Shi C, Yu L, Chai M, Niu Z, Li R. The distribution and risk of mercury in Shenzhen mangroves, representative urban mangroves affected by human activities in China. Mar Pollut Bull. 2020; 151: 110866 (11p)
- Jeong H, Choi JY, Choi DH, Noh JH, Ra K. Heavy metal pollution assessment in coastal sediments and bioaccumulation on seagrass (Enhalus acoroides) of Palau. Mar Pollut Bull. 2021; 163: 1–7.
- Analuddin K, Sharma S, Jamili, Septiana A, Sahidin I, Rianse U, et al. Heavy metal bioaccumulation in mangrove ecosystem at the coral triangle ecoregion, Southeast Sulawesi, Indonesia. Mar Pollut Bull. 2017; 125 (1–2): 472–480.
- 20. de Almeida Duarte LF, de Souza CA, Pereira CDS, Pinheiro MAA. Metal toxicity assessment by sentinel species of mangroves: In situ case study integrating chemical and biomarkers analyses. Ecotoxicol. Environ Saf. 2017; 145: 367–376.
- Mapenzi LL, Shimba MJ, Moto EA, Maghembe RS, Mmochi AJ. Heavy metals bio-accumulation in tilapia and catfish species in Lake Rukwa ecosystem Tanzania. J Geochem Explor. 2020; 208: 106413 (12p)
- 22. Costa-Böddeker S, Thuyên LX, Hoelzmann P, de Stigter HC, van Gaever P, Huy HD, et al. Heavy metal pollution in a reforested mangrove ecosystem (Can Gio Biosphere Reserve, Southern Vietnam): Effects of natural and anthropogenic stressors over a thirty-year history. Sci Total Environ. 2020; 716; 137035 (15p)
- Melville F, Andersen LE, Jolley DF. The Gladstone (Australia) oil spill - Impacts on intertidal areas: Baseline and six months post-spill. Mar Pollut Bull. 2009; 58(2): 263–271.
- 24. Marambio AY, Saavedra JV, Enciso LÑ, Marras AL, Serrano AE, Peláez RM, et al. Data on metal accumulation in the tails of the lizard Microlophus atacamensis in a coastal zone of the Atacama Desert, northern Chile: A non-destructive biomonitoring tool for heavy metal pollution. Data in Brief . 2020; 32: 106032 (12p).
- Nadgórska–Socha A, Kandziora-Ciupa M, Trzęsicki M, Barczyk G. Air pollution tolerance index and heavy metal bioaccumulation in selected plant species from urban biotopes. Chemosphere. 2017; 183: 471– 482.
- Alzahrani DA, Selim E-MM, El-Sherbiny MM. Ecological assessment of heavy metals in the grey mangrove (Avicennia marina) and associated sediments along the Red Sea coast of Saudi Arabia. Oceanologia. 2018; 60(4): 513–526.
- 27. Oo CW, Kassim MJ, Pizzi A. Characterization and

performance of Rhizophora apiculata mangrove

polyflavonoid tannins in the adsorption of copper (II) and lead (II). Ind Crops Prod. 2009; 30(1): 152–161.

28. Ma W, Li X, Wang Q, Ren Z, Crabbe MJC, Wang L. Tandem oligomeric expression of metallothionein enhance heavy metal tolerance and bioaccumulation

in Escherichia coli. Ecotoxicol. Environ Saf. 2019;

- 181: 301–307.
 29. Zhang Z, Fang Z, Li J, Sui T, Lin L, Xu X. Copper, zinc, manganese, cadmium and chromium in crabs from the mangrove wetlands in Qi'ao Island, South China: Levels, bioaccumulation and dietary exposure. Watershed Ecol. Environ 2019; 1: 26–32.
- Li R, Wu S, Chai M, Xie S. Denitrifier communities differ in mangrove wetlands across China. Mar Pollut Bull. 2020; 155: 111160 (11p)
- Nour HE, El-Sorogy AS, Abd El-Wahab M, Nouh ES, Mohamaden M, Al-Kahtany K. Contamination and ecological risk assessment of heavy metals pollution from the Shalateen coastal sediments, Red Sea, Egypt. Mar Pollut Bull. 2019; 144: 167–172.
- 32. Xiao R, Bai J, Lu Q, Zhao Q, Gao Z, Wen X, et al. Fractionation, transfer, and ecological risks of heavy metals in riparian and ditch wetlands across a 100year chronosequence of reclamation in an estuary of China. Sci Total Environ. 2015; 517: 66–75.
- Chai M, Li R, Qiu Z, Niu Z, Shen X. Mercury distribution and transfer in sediment-mangrove system in urban mangroves of fast-developing coastal region, Southern China. Estuar. Coast Shelf Sci. 2020; 240: 106770 (11p).
- Hilmi E, Siregar AS, Febryanni L, Novaliani R, Amir SA, Syakti AD. Struktur Komunitas, Zonasi Dan Keanekaragaman Hayati Vegetasi Mangrove Di Segara Anakan Cilacap. Omni-Akuatika . 2015; 11(2): 20–32.
- 35. Hilmi E, Amron A, Sari LK, Cahyo TN, Siregar AS. The Mangrove Landscape and Zonation following Soil Properties and Water Inundation Distribution in Segara Anakan Cilacap. J Man Hut Trop. 2021; 27(3): 152–164.
- 36. Hilmi E, Sari LK, Cahyo TN, Mahdiana A, Soedibya PHT, Sudiana E. Survival and growth rates of mangroves planted in vertical and horizontal aquaponic systems in North Jakarta, Indonesia. Biodiversitas. 2022; 23(2): 686–693.
- 37. Monteiro JM, de Souza JSN, Lins Neto EMF, Scopel K, Trindade EF. Does total tannin content explain the use value of spontaneous medicinal plants from the Brazilian semi-arid region? Rev Bras Farmacogn. 2014; 24(2): 116–123.
- Xiong Y, Liao B, Proffitt E, Guan W, Sun Y, Wang F, et al. Soil carbon storage in mangroves is primarily controlled by soil properties: A study at Dongzhai Bay, China. Sci Total Environ. 2018; 619–620: 1226– 1235.
- Win S, Towprayoon S, Chidthaisong A. Adaptation of mangrove trees to different salinity areas in the Ayeyarwaddy Delta Coastal Zone, Myanmar. Estuar Coast Shelf Sci. 2019; 228: 106389-106397
- 40. SNI. SNI 6989.7. Air dan Air Limbah Bagian 7:

Cara Uji Seng (Zn) secara Spektrofotometri Serapan Atom (SSA) - Nyala. Badan Standarisasi Nasional. Jakarta 2009.

P-ISSN: 2078-8665

E-ISSN: 2411-7986

- 41. Lin Y, Lu J, Wu J. Heavy metals pollution and health risk assessment in farmed scallops: Low level of Cd in coastal water could lead to high risk of seafood. Ecotoxicol. Environ Saf. 2021; 208: 111768 (10p).
- MacFarlane G., Pulkownik A, Burchett M. Accumulation and distribution of heavy metals in the grey mangrove, Avicennia marina (Forsk.)Vierh.: biological indication potential. Environ Pollut. 2003; 123(1): 139–151.
- 43. Hilmi E, Sari LK, Amron A, Cahyo TN, Siregar AS. Mangrove cluster as adaptation pattern of mangrove ecosystem in Segara Anakan Lagoon. IOP Conf Ser: Earth Environ Sci. 2021; 746(1). 012022 (11p)
- Ludwig J, Renold J. Statistical Ecology (A primer on Methods and computing). Toronto, Canada: John Wiley & Sons. In; 1988. 244 p.
- Sari SP, Rosalina D. Mapping and Monitoring of Mangrove Density Changes on tin Mining Area. Procedia Environ Sci Eng Manag. 2016; 33: 436– 442.
- Ismail, Sulistiono, Hariyadi S, Madduppa H. Condition and mangrove density in Segara Anakan, Cilacap Regency, Central Java Province, Indonesia. AACL Bioflux. 2018;11(4): 1055–1068.
- 47. Jiang S, Weng B, Liu T, Su Y, Liu J, Lu H, et al. Response of phenolic metabolism to cadmium and phenanthrene and its influence on pollutant translocations in the mangrove plant Aegiceras corniculatum (L.) Blanco (Ac). Ecotoxicol Environ Saf. 2017; 141: 290–297.
- 48. Adyasari D, Pratama MA, Teguh NA, Sabdaningsih A, Kusumaningtyas MA, Dimova N. Anthropogenic impact on Indonesian coastal water and ecosystems: Current status and future opportunities. Mar Pollut Bull. 2021; 171(March): 112689 (14p)
- Barreto MB, Lo Mónaco S, Díaz R, Barreto-Pittol E, López L, Peralba M do CR. Soil organic carbon of mangrove forests (Rhizophora and Avicennia) of the Venezuelan Caribbean coast. Org Geochem. 2016; 100: 51–61.
- 50. Chen S, Chen B, Chen G, Ji J, Yu W, Liao J, et al. Higher soil organic carbon sequestration potential at a rehabilitated mangrove comprised of Aegiceras corniculatum compared to Kandelia obovata. Sci Total Environ. 2021; 752: 142279 (9p).
- 51. Choi JY, Jeong H, Choi KY, Hong GH, Yang DB, Kim K, et al. Source identification and implications of heavy metals in urban roads for the coastal pollution in a beach town, Busan, Korea. Mar Pollut Bull. 2020; 161: 111724 (12p).
- 52. Hao Z, Chen L, Wang C, Zou X, Zheng F, Feng W, et al. Heavy metal distribution and bioaccumulation ability in marine organisms from coastal regions of Hainan and Zhoushan, China. Chemosphere. 2019; 226: 340–350.
- 53. Xiao K, Li H, Shananan M, Zhang X, Wang X, Zhang Y, et al. Coastal water quality assessment and groundwater transport in a subtropical mangrove

Published Online First: January, 2022

646: 1419-1432. 54. Dai M, Lu H, Liu W, Jia H, Hong H, Liu J, et al. Phosphorus mediation of cadmium stress in two mangrove seedlings Avicennia marina and Kandelia obovata differing in cadmium accumulation. Ecotoxicol. Environ Saf. 2017; 139: 272-279.

swamp in Daya Bay, China. Sci Total Environ. 2019;

- 55. Xie Q, Qian L, Liu S, Wang Y, Zhang Y, Wang D. Assessment of long-term effects from cage culture practices on heavy metal accumulation in sediment and fish. Ecotoxicol Environ Saf. 2020; 194: 110433 (7p).
- 56. Liu S, Liu Y, Yang D, Li C, Zhao Y, Ma H, et al. Trace elements in shellfish from Shenzhen, China: Implication of coastal water pollution and human exposure. Environ Pollut. 2020; 263: 114582 (8p)
- 57. Liu L, Wang H jun, Yue Q. China's coastal wetlands: Ecological challenges, restoration, and management suggestions. Reg Stud Mar Sci. 2020; 37: 101337
- 58. MacFarlane GR, Burchett MD. Zinc distribution and excretion in the leaves of the grey mangrove, Avicennia marina (Forsk.) Vierh. Environ Exp Bot. 1999; 41(2): 167-175.
- 59. Hilmi E, Sari LK, Setijanto. The mangrove landscaping based on Water Quality: (Case Study in Segara Anakan Lagoon and Meranti Island). IOP Conf. Ser: Earth Environ Sci . 2019; 255(1). https://doi.org/ 10.1088/1755-1315/255/1/012028
- 60. Penha-Lopes G, Kristensen E, Flindt M, Mangion P, Bouillon S, Paula J. The role of biogenic structures on the biogeochemical functioning of mangrove constructed wetlands sediments - A mesocosm approach. Mar Pollut Bull. 2010; 60(4): 560-572.
- 61. Sitoe AA, Mandlate LJC, Guedes BS. Biomass and carbon stocks of Sofala Bay mangrove forests. Forests. 2014; 5(8): 1967-1981.
- 62. Yu F, Tang S, Shi X, Liang X, Liu K, Huang Y, et al. Phytoextraction of metal(loid)s from contaminated soils by six plant species: A field study. Sci Total Environ. 2022; 804: 150282 (12 p)
- McCutcheon SC, Susarla S, Medina VF. Phytoremediation: An ecological solution to organic chemical contamination. Ecol Eng. 2002; 18(5): 647-
- 64. Kagalkar AN, Jadhav MU, Bapat VA, Govindwar SP. Phytodegradation of the triphenylmethane dye Malachite Green mediated by cell suspension cultures of Blumea malcolmii Hook. Bioresour Technol. 2011; 102(22): 10312-10318.
- 65. Radziemska M, Gusiatin ZM, Cydzik-Kwiatkowska A, Cerdà A, Pecina V, Bęś A, et al. Insight into metal

immobilization and microbial community structure in soil from a steel disposal dump phytostabilized with composted, pyrolyzed or gasified wastes. Chemosphere. 2021; 272: 129576 (16p).

P-ISSN: 2078-8665

E-ISSN: 2411-7986

- 66. Zhang X, Yu J, Huang Z, Li H, Liu X, Huang J, et al. Enhanced Cd phytostabilization and rhizosphere bacterial diversity of Robinia pseudoacacia L. by endophyte Enterobacter sp. YG-14 combined with sludge biochar. Sci Total Environ. 2021; 787: 147660 (12p).
- 67. de Oliveira DCM, Correia RRS, Marinho CC, Guimarães JRD. Mercury methylation in sediments of a Brazilian mangrove under different vegetation covers and salinities. Chemosphere. 2015; 127: 214-
- 68. Yin P, Yin M, Cai Z, Wu G, Lin G, Zhou J. Structural inflexibility of the rhizosphere microbiome in mangrove plant Kandelia obovata under elevated CO2. Mar Environ Res. 2018; 140: 422-432.
- St. Gelais AT, Costa-Pierce BA. Mercury concentrations in Northwest Atlantic winter-caught, male spiny dogfish (Squalus acanthias): A geographic mercury comparison and risk-reward framework for human consumption. Mar Pollut Bull. 2016; 102(1): 199-205.
- 70. Ariani F, Effendi H, Suprihatin. Water and sediment oil content spread in Dumai coastal waters, Riau Province, Indonesia. Egypt J Aquat Res. 2016; 42(4): 411-416.
- Rachmatin D. Aplikasi Metode-metode Agglomerative Dalam Analisis Klaster Pada Data Tingkat Polusi Udara. Jurnal Ilmiah Infinity. 2014; 3(2): 133-149.

Open Access Published Online First: January, 2022

تقييم المخاطر البيئية للتلوث بالزئيق في النظام البيئي لأشجار المانغروف في سيجارا أناكان سيلاكاب، التقييم المخاطر البيئية للتلوث بالزئيق في النظام البيئي المخاطر البيئية المخاطر المخ

 2 روز ديوي 2 اندانج حلمي 1* تيوكو جنايدي 2 عارف مهديانة

لبر نامج إدارة الموارد المانية وبرنامج Magister SDA ، كلية المصايد والعلوم البحرية ، جامعة .Jenderal Soedirman جي .دكتور سوبارنو ، بوروكيرتو أوتارا ، بانيوماس 53122 ، جاوة الوسطى ، إندونيسيا.

²برنامج إدارة الموارد المانية ، كلية المصايد وعلوم البحار ، جامعة جيندير ال سوديرمان جي .دكتور سوبارنو ، بوروكيرتو أوتارا ، باتيوماس 53122 ، جاوة الوسطى ، إندونيسيا.

3. برنامج العلوم البحرية ، كلية المصايد وعلوم البحار ، جامعة جينديرال سوديرمان .جي .دكتور سوبارنو ، بوروكيرتو أوتارا ، بانيوماس 53122 ، جاوة الوسطى ، إندونيسيا

الخلاصة:

P-ISSN: 2078-8665

E-ISSN: 2411-7986

في هذه الدراسة, يعتبر تقييم المخاطر البينية للتلوث بالزنبق وسيلة لتحليل جانب المخاطر البينية للنظام البيني باستخدام التأثير المحتمل للتلوث في التربة والمياه والكاننات الحية. يمكن إظهار تقييم المخاطر البينية في المنطقة الساحلية من خلال تقسيم مناطق المنغروف ، والتكتل واستيفاء تراكم الزنبق. تهدف هذه الدراسة إلى تحليل تقييم المخاطر البينية للزنبق المحتمل (بما في ذلك الانتقال و التراكم البيولوجي) باستخدام مؤشر توزيع الأنواع ، والتكتل ، والتقسيم إلى مناطق واستيفاء تراكم الزنبق. أظهرت النتائج أن Segara Anakan كانت معرضة لخطر التلوث بالزنبق ، باستخدام مؤشرات مثل احتمال تلوث الزنبق في الجسم المائي كان (0.013 ± 0.013 جزء في المليون ، الركيزة والرواسب كانت 0.013 ± 0.001 جزء في المليون. للحد من تأثير التلوث بالزنبق بمكن إجراؤه عن طريق زراعة المنغروف ، بعد قدرة تراكم الزنبق في الساق واللحاء بين 0.001 و 0.001 جزء في المليون . استخدم المؤشر الثاني لقدرة المنغروف في تقليل تأثير ملوثات الزنبق في أوراق المنغروف بين 0.002 و 0.002 جزء في المليون . استخدم المؤشر الثاني لقدرة المنغروف في تقليل تأثير ملوثات الزنبق في مؤشر عوامل النراكم الأحيائي ، والتي تراوحت بين 0.001 و 0.002 و مناس التراكم الأحيائي ، والتي تراوحت بين 0.002 و 0.002 والتربق بالزنبق و التراكم وتقليل تأثير التلوث بالزنبق و والم التراكم بالأربق و المراكم والتورث بالزنبق و والمراكم والتورث و المراكم والتورث و المراكم والتورث و المراكم والتورث والمراكم والتورث والتورث والمراكم والتورث والتورث والتورث والتورث والمراكم والتورث والتور

الكلمات المفتاحية: التلوث بالزئبق، التكتل، تقسيم مناطق المنغروف؛ عامل التراكم الأحيائي؛ عامل الانتقال تقييم المخاطر البيئية.

The Mangrove Strengthen and Ecological Risk Assessment of Mercury Contamination in the Segara Anakan Cilacap Lagoon, Indonesia

ORI			

SIMILARITY INDEX

INTERNET SOURCES

PUBLICATIONS

% STUDENT PAPERS

PRIMARY SOURCES

assets.researchsquare.com

Internet Source

csg.uobabylon.edu.iq

Internet Source

Exclude quotes

Off

Exclude matches

< 3%

Exclude bibliography