

30-DOMESTICATION_RED SEAWEED_ANDRE-TJAHJO

by Turnitin Checker

Submission date: 05-Apr-2023 06:03PM (UTC+1000)

Submission ID: 2056438720

File name: 30-DOMESTICATION_RED_SEAWEED_ANDRE-TJAHJO.pdf (297.55K)

Word count: 3430

Character count: 17649

1 Domestication of Red Seaweed (*Gelidium latifolium*) in Different Culture Media

Andri Wijayanto^{1*}, Ita Widowati² and Tjahjo Winanto³

7
4
¹Master of Coastal Resources Management, Department of Aquatic Resources Management, Fisheries and Marine Sciences Faculty, Diponegoro University

²Department of Marine Science, Faculty of Fisheries and Marine Science, Diponegoro University
Jl. Prof. H. Soedarto, S.H, Tembalang, Semarang 50275, Indonesia

³Faculty of Fisheries and Marine Science, Jenderal Soedirman University
Jl. Dr Soeparno, Komplek GOR Soesilo Soedarman, Purwokerto, Jawa Tengah 53122, Indonesia
Email: andriwijayanto03@yahoo.com

Abstract

Gelidium latifolium is one of red seaweed types potentially can be developed as an industrial raw material. Since *Gelidium* is currently taken from ocean, the availability of seaweed from aquaculture is necessary to overcome the small number of its availability in nature. In Indonesia, *G. latifolium* cultivation has not been carried out so that domestication is required. The use of macro and micro nutrients in growth media is essentially needed for the domestication process. Domestication requires fast media and place for growth. The purpose of this study is to determine the growth of biomass and the survival of *G. latifolium* in different culture media. The study was conducted in a semi-outdoor research laboratory. The method used in this research is laboratory experimental method and Completely Randomized Design (CRD) with the treatment applied using 3 types of culture media (Urea: Za: TSP) by comparison (A) 100: 50: 50% (2 g.L⁻¹), (B) 75: 75: 50% (2 g.L⁻¹) and (C) 75: 50: 75% (2 g.L⁻¹), with 3 replications. The seaweed was kept in 10 L of water in aeration equipped aquarium and filled with 10 g of *G. latifolium* on each treatment. The best growth rate of *G. latifolium* biomass is 5.67± 0.58 g and 100±0% are survived in C culture medium with a concentration of 75% Urea: 50% ZA: 75% TSP (2 g.L⁻¹).

Keywords: red seaweed, weight growth, survival, culture media, semi-outdoor

Introduction

Indonesia has high seaweed resource potential and it is spread almost in all sea waters of the country, including *Gelidium latifolium*. As one of red seaweed types, this species has high potential for industrial raw material. Red seaweed particularly can be used as a source of products in pharmaceuticals, food, and aquaculture (Widowati et al., 2014). The seaweed cultivation has prospect since its demand in global trade is relatively high and it is increasing (Sanjeeva et al., 2017).

Gelidium is usually used for jelly production (Kang et al., 2013), paper (Seo et al., 2009), natural antifungal against fungus *Candida albicans* (Lutfiyanti et al., 2012), antioxidant (Seo et al., 2012), raw material in bioethanol production (Meinita et al., 2013; Hong et al., 2014; Meinita et al., 2017), antimicrobial (Kang et al., 2016), and for reducing hepatic lipids and plasma in diabetics (Yang et al., 2017). To date *Gelidium* grows in fast-growing parts of the banks, farmers still depend on the season, and harvest from nature so that it is feared that sustainable use will cause *Gelidium* to

experience extinction so that it needs domestication (Sjafrie, 1999). To have a normal growth and high survival, the seaweed need an adaptation from conditions in nature to cultivation. The trial of the cultivation has been carried out on the *Gelidium amansii* (Aries and Jubaedah, 2011).

Domestication requires a sufficient nutrient for growth Growth stimulant is one of the media components demanded for growth and regeneration (Fadel et al., 2013). The purpose of this study is to determine the growth of biomass and the survival of *G. latifolium* in different culture media.

Materials and Methods

The study was conducted in a research laboratory (semi-outdoor). Samples of *G. latifolium* seedlings were taken from Kebumen Coastal Waters-Southern coast of Central Java. Seedlings were acclimatized in the laboratory for 1-2 weeks. The selection of seedlings was based on their quality, number of branches, healthy, natural-bright colored, clean, disease-free, have good *thallus* and

*) Corresponding author
© Ilmu Kelautan, UNDIP

holdfast and those that are young shoots (Yong et al., 2011). The seaweed domestication was kept by used 10 L of water in aeration equipped aquarium and filled with 10 g of *G. latifolium* seaweed in each treatment. The research used 3 types of culture media as treatments (Urea: Za: TSP) i.e. (A) 100: 50: 50% (2 g.L⁻¹), (B) 75: 75: 50% (2 g.L⁻¹) and (C) 75: 50: 75% (2 g.L⁻¹), with 3 replications for each treatment.

Observed parameters

The weight gain or biomass of seaweed (G) is the ratio between the differences in the final weight of the experiment (W₁) minus the initial weight of the experiment (W₀). This research was conducted by weighing the seeds using analytical scales. The weight gain can be calculated by the Effendi formula, (1979).

Survival rate (SR) is a comparison between the percentage of the total number of seaweed individuals that live at the end of the experiment (N₁) divided by the total number of seaweed individuals at the beginning of the experiment (N₀). Survival rates calculated using the formula of Goddard, (1996).

Data Analysis

The data obtained were analyzed descriptively in the form of tables and graphs. To find out the difference in growth rate and survival rate of treatment, ANOVA test was used. If there is distinction between treatments (P<0.05) then the analysis will be proceed with Tukey test. Data analysis was performed by using SPSS software version 24.

Results and Discussion

Absolute Growth Rate

The results of observations of absolute biomass growth rate on different culture media showed that the highest *G. latifolium* biomass was in C culture medium treatment of 5.67± 0.58 g, followed by B culture medium of 3.33± 1.15 g, and the lastly, the lowest in A culture medium of 2.67±0.058 g (Figure 1). ANOVA resulted that the combination of Urea, ZA, and TSP fertilizers between treatments showed a significant difference (P <0.05) to increase the of *G. latifolium* biomass. Meanwhile the results of further Tukey tests showed that the addition of *G. latifolium* biomass in the treatment of C culture medium was significantly different (P <0.05) from the treatment of A culture medium.

The growth rate of biomass in C culture medium was higher than in B culture medium, and the A culture medium was the lowest. From the initial phase of planting, it was observed a continuously gradual increase in biomass until the 28th day of the final phase. The C culture medium Showed the fastest growth and in biomass as well, whereas the B and A culture media showed a slower growth and their biomass were low. It was suggested that the combination of growth media (Urea: ZA: TSP) with the good concentration could increase the growth of *G. latifolium* offesting on well- fast-growing holdfast and many branches thallus. According to Widyawati et al. (2019), the nitrogen element in Urea can accelerate thallus growth. While the nitrogen element in ZA fertilizer can accelerate plant growth and increase protein content. Meanwhile according to Wahyurini (2014), element nitrogen in Urea fertilizer and element phosphate in TSP fertilizer will greatly affect the growth rate of seaweed.

Phosphate content in culture media is an important component of stimulating thallus growth, accelerating and strengthening the growth of young plants into mature plants. Phosphate causes the high growth rate so that the weight of the biomass becomes high (Lingga and Marsono, 2007). According to Prakoeswa et al. (2009), the use of appropriate growth regulators in the media helps thallus and organs of seaweed grow, while in the media without the addition of growth regulators, the growth could be very stunted and may not even grow at all.

Survival Rate

The results of survival observations showed that *G. latifolium* in C culture media treatment was the highest of 100±0%, followed by 99.67±0.58% in B culture medium and the lowest was in (A) culture medium of 98.67±2.31% (Figure 2). ANOVA results showed that the combination of Urea, ZA, and TSP fertilizers between treatments showed no significant difference (P> 0.05) to the increase of *G. latifolium* biomass.

Survival rate in this study showed, that in the culture media C was the highest, compared to culture media B, and the lowest was in culture media A. Based on the beginning of planting until the 14th day, the survival of *G. latifolium* was, stable; it was continued until days 21st to 28th when the growth decreased due to the death. This decrease of survival rate can be seen on the higher of the standard deviation, and the value of the data is far from the mean survival in *G. latifolium*. Seaweed with morphological changes in the color of pale white stems, on lead stems rather red color that

causes the stems to become soft and broken. The death of explants may due to the ability of explant adaptation when explaining hormones in different nutritional compositions when excess nutrient composition will divert interactions that can activate explants (Fadel *et al.*, 2013).

The survival rate in C culture medium is considered to be higher because the application of fertilizer concentration is fit with the need for the development of seaweed growth cells. Seaweed requires nutrients for growth and survival (Mukhlis *et al.*, 2016). Urea is a single type of fertilizer which nitrogen function accelerates *thallus* growth (Stiaji *et al.*, 2012). According to Muarif *et al.* (2017) to maintain the survival of seaweed, adequate nutrition is required for the formation of new tissue or of shoots to stay alive. Essentially a lot of nitrogen is

needed by seaweed as an energy supplier in the process of photosynthesis (Kushartono *et al.*, 2009). Meanwhile, the lowest survival rate on A culture medium was thought to be the high composition of culture media resulting in death. According to Mulyaningrum *et al.* (2014), media inhibition factors is, if main elements of micronutrients were in excessive amounts and become toxic. When the amounts of B was too high, it ends to high mortality. The utilization of micronutrients by explants is too high, resulting in high mortality. According to Wahyurini (2014), too many doses of fertilizer (Urea and TSP) given will cause growth to be stunted and seaweed undergoes have a morphological changes including the color of pale white stems, on lead stems rather red which causes the stems to become soft and broken.

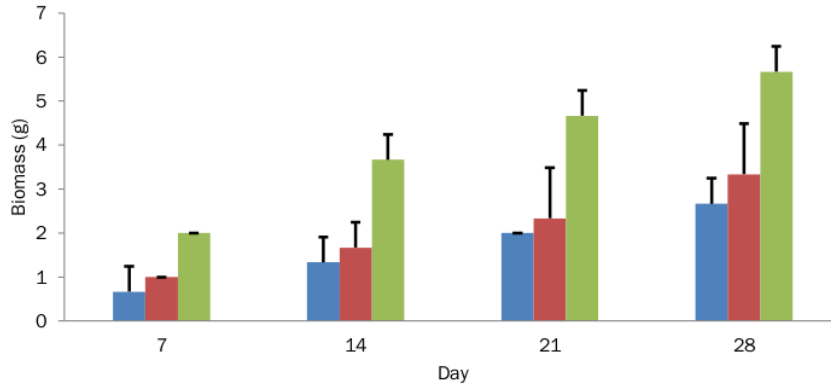


Figure 1. The average weight growth rate of *G. latifolium* cultured in different media
 Note. ■ = (A) urea, ZA, TSP (100:50:50) %; ■ = (B) urea, ZA, TSP (75:75:50) %; ■ = (C) urea, ZA, TSP (75:50:75) %

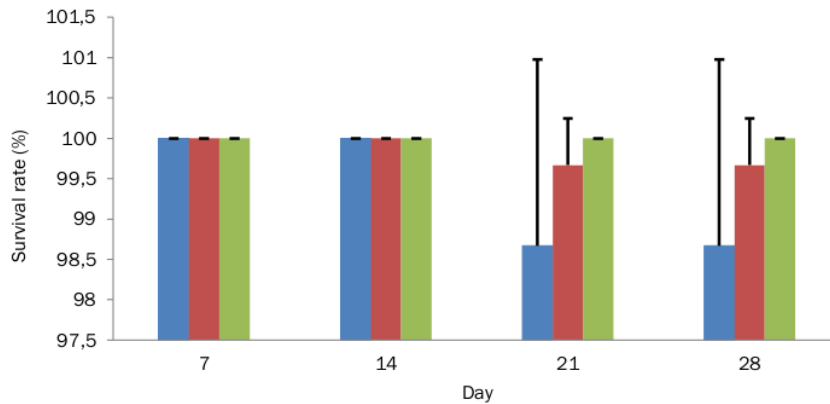


Figure 2. The average survival rate of *G. latifolium* seaweed cultured in different media
 Note. ■ = (A) urea, ZA, TSP (100:50:50) %; ■ = (B) urea, ZA, TSP (75:75:50) %; ■ = (C) urea, ZA, TSP (75:50:75) %

The results of temperature measurements demonstrated that from the beginning of planting until the harvest time there were no significant fluctuations, the values were ranging from 29-30 °C. Temperature measurements showed that the temperature in the study water media was suitable for the growth of *G. latifolium*. As stated by Aslan (1998), good temperatures for seaweed growth ranged from 26-33 °C. The requirements for cultivation, good water temperature for seaweed growth is 24-30 °C (Sulma and Manoppo, 2008). The result of pH measurements is pH 7, meaning, that the pH on the media is still within the normal range for the growth of *G. latifolium* cells. According to Kordi and Tancung (2007), cultivation will work well at pH 6.5 - 9.0. Meanwhile, according to Mudeng et al. (2015), the optimum pH required for seaweed cultivation is 6.5 - 8.5. All algae with a pH ranging from 6.8 to 9.6 can still live and grow (Ain et al., 2014; Burdames and Ngangi, 2014). The degree of acidity (pH) affects the level of water fertility. Acidic waters will be less productive. At low pH (high acidity) the dissolved oxygen content will decrease, so oxygen consumption decreases (Afandi et al., 2015). Salinity results on *G. latifolium* indicates conditions that support growth. Based on these results it can be seen that salinity of 30-33 ppt support growth of *G. latifolium*. According to Guo et al. (2014), the optimal range of salinity for seaweed growth is 25-33 ppt. Seaweed will growth slowly if the salinity is too low (<15 ppt) or too high (> 35 ppt), from the salinity range following its living conditions it can cause interference on osmoregulation process that occurs in cells and physiological seaweed (Choi et al., 2010).

Conclusion

The best growth rate in culture media C with a concentration of 75% Urea: 50% ZA: 75% TSP (2 g. L⁻¹). The biomass growth rate on different culture media showed that the highest *G. latifolium* biomass increased in C culture medium treatment 5.67± 0.58 g, followed by B culture medium 3.33± 1.15 g, and the lastly, the lowest A culture medium 2.67±0.058 g and The survival observations showed that *G. latifolium* seaweed in the highest C culture media treatment was 100±0%, followed by 99.67±0.58% in B culture medium and the lowest was in (A) culture medium of 98.67±2.31%.

References

- Afandi, A., Nirmala, K. & Budiardi, T. 2015. Production, Rendemen and Gel Strength of Three Seaweed Varieties *Kappaphycus alvarezii* Cultivated by Long Line Method. *J. Kel. Nasional*, 10(1): 43-53
- Ain, N., Ruswahyuni & Widyorini, N. 2014. Hubungan Kerapatan Rumput Laut dengan Substrat Dasar Berbeda di Perairan Pantai Bandengan, Jepara. *Diponegoro J. Maquares*. 3(1): 99-107.
- Aries, G. & Jubaedah, I. 2011. Ujicoba Pengembangan Budidaya Rumput Laut (*Gelidium amansii*) dengan Metode Vertikal Longline. *J. Penyuluhan Perikanan dan Kelautan*, 5(1):9-16
- Arikunto, S. 2003. Research Procedure: A Practical Approach. Revised Edition. Rineka Cipta, Jakarta. 378 pp.
- Aslan M.L, 1998. Seaweed Cultivation. Kanisius, Yogyakarta. 96 pp.
- Burdames, Y & Ngangi, E.L.A. 2014. Kondisi Lingkungan Perairan Budi Daya Rumput Laut di Desa Arakan, Kabupaten Minahasa Selatan. *Jurnal Budidaya Perairan*. 2(3):69-75.
- Choi, T.S., Kang, J.H. & Kim., K.Y. 2010. Effect of salinity on growth and nutrient uptake of *Ulva pertusa* (Chlorophyta) from an eelgrass bed. *Algae*, 25(1):17-25. doi: 10.4490/algae.2010.25.1.017
- Effendi, M.I., 1979. Fisheries Biology. Dewi Sri Foundation, Bogor. 112 pp.
- Fadel, A.H., Gerung, G.S., Suryati, E. & Rumengan, I.F., 2013. The Effects of Stimulant Growth Hormones on Tissue Culture of Seaweed *Kappaphycus alvarezii* in vitro. *Aquatic Science & Management*, Edisi Khusus 1:77-84.
- Goddard. S. 1996. Feed Management in Intensive Aquaculture. Chapman and Hall, New York.
- Guo, H., Yao, J., Sun, Z. & Duan, D. 2014. Effect of Temperature, Irradiance on the Growth of the Green Alga *Caulerpa lentillifera* (Bryopsidophyceae, Chlorophyta). *J. App. Phycol.*, 27(2):879-885.
- Hong, I.K., Jeon H. & Lee, S.B. 2014. Comparison of red, brown and green seaweeds on enzymatic saccharification process. *J. Ind. Eng. Chem.* 20: 2687-2691.
- Kang, M., Kim, S.W., Kim, J.W., Kim, T.H. & Kim, J.S. 2013. Optimization of levulinic acid production from *Gelidium amansii*. *Renewable Energy*. 54: 173-179.
- Kang, M.C., Kang N, Kim SY, Lima I.S., Ko S.C., Kim, Y.T., Kim, Y.B., Jeung, H.D., Choi, K.S. & Jeon, Y.J. 2016. Popular edible seaweed, *Gelidium*

- amansii* prevents against diet induced obesity. *Food Chem. Toxicol.* 90:181-7.
- Kordi, K.M.G.H. & Tancung A.B., 2007. Pengelolaan Kualitas Air dalam Budidaya Perairan. Rineka Cipta. Jakarta.
- Kushartono, E.W., Suryono. & Setyaningrum, M.R.E., 2009. Aplikasi perbedaan komposisi N, P dan K pada budidaya *Euचेuma cottonii* di Perairan Teluk Awur, Jepara. *Ilmu Kelautan : Ind. J. Mar. Sci.*, 14(3):164-169. doi : 10.14710/ik.ijms.14.3.164-169
- Lingga, P., & Marsono. 2007. Instructions for use of fertilizers. Self-help Spreaders (In Indonesia). Jakarta. 8 – 38.
- Lutfiyanti, R., Ma'ruf, W.F. & Dewi, E.N. 2012. Aktivitas Antijamur Senyawa Bioaktif Ekstrak *Gelidium latifolium* Terhadap *Candida albicans*. *J. Pengolahan dan Bioteknologi Hasil Perikanan*, 1(1):1-8.
- Meinita, M.D.N., Marhaeni, B., Hong, Y.K. & Jeong, G.T. 2017. Enzymatic saccharification of agar waste from *Gracilaria verrucosa* and *Gelidium latifolium* for bioethanol production. *J Appl. Phycol.*, 29(6):3201-3209.
- Meinita, M.D.N., Marhaeni, B., Winanto, T., Jeong, G.T., Khan, M.N.A. & Hong, Y.K. 2013. Comparison of Agarophytes (*Gelidium*, *Gracilaria*, and *Gracilariopsis*) as Potential Resources for Bioethanol Production. *J. App. Phycol.*, 25(6):1957-1961.
- Muarif, Ya'la., Z.R. & Rusaini. 2017. Growth of *Euचेuma cottonii* Seaweed Cultivated Culturally *In Vitro* With a Different Number of Thallus (In Indonesia). Prosiding Simposium Nasional Kelautan dan Perikanan IV Universitas Hasanuddin.,251-259:
- Mudeng, J.D., Kolopita, M.E.F. & Rahman, A. 2015. Kondisi Lingkungan Perairan Pada Lahan Budidaya Rumput Laut *Kappaphycus alvarezii* di Desa Jayakarsa Kabupaten Minahasa Utara. *J. Budidaya Perairan*, 3(1):172-186
- Mukhlis, Ya'la, Z.R. & Rusani. 2016. The Growth Seaweed *Kappaphycus alvarezii* Explants Immersed at Different Duration in Urea and TSP Solution. *Simposium Nasional Kelautan dan Perikanan III Universitas Hasanuddin*, 434-446: ISBN: 978-602-71759-2-1
- Mulyaningrum, S.R.H., Daud, R. & Badraeni. 2014. Vegetative Propagation of *Gracilaria* sp. Seaweed. Through Tissue Culture. *J. Ris. Aquaculture*, 9(2):203-214.
- Sanjewa, K.A., Lee, J.S., Kim, W.S. & Jeon, Y.J., 2017. The potential of brown-algae polysaccharides for the development of anticancer agents: an update on anticancer effects reported for fucoidan and laminaran. *Carbohydr Polym.* 177: 451-9. doi: 10.1016/j.carbpol.2017.09.005
- Seo, M.J., Lee, O.H., Choi, H.S. & Lee, B.Y., 2012. Extract from edible red seaweed (*Gelidium amansii*) inhibits lipid accumulation and ROS production during differentiation in 3T3-L1 cells. *Prev. Nutr. Food Sci.* 17:129-35.
- Seo, Y.B., Lee, Y.W., Lee, C.H. & You, H.C. 2009. Red algae and their use in papermaking. *Bioresource Technology*, 101(7):2549-2553.
- Setiaji, K., Santosa, G. W., Sunaryo. 2012. Pengaruh Penambahan NPK dan UREA pada Media Air Pemeliharaan Terhadap Pertumbuhan Rumput laut *Caulerpa racemosa* var. *uvifera*. *Journal of Marine Research*, 1 (2): 45-50
- Sjafrie, N.D.M. 1999. Some Notes About *Gelidium* (Rhodophyta). *Oceana*, 24 (3):1-10.
- Steel, R.G.D. & Torrie, J.H. 1993. Statistics Principles and Procedures of a Biometric Approach. P.T. Gramedia Pustaka Utama, Jakarta. 748 pp.
- Sulma, S., & Manoppo, A. 2008. Kesesuaian fisik perairan untuk budidaya rumput laut di perairan Bali menggunakan data penginderaan jauh. Bandung: Pusat Pengembangan Pemanfaatan dan Teknologi Penginderaan Jauh.LAPAN. PIT MAPIN XVII, 10 hlm.
- Wahyurini, E.T. 2014. Rasio Pupuk Urea dan Triple Superfosfat (TSP) yang Berbeda Terhadap Laju Pertumbuhan Rumput Laut (*Gracillaria gigas* Harvey) denga Skala Lab. *AGROSAINS*, 01 (01):1-9.
- Widowati, I., Lubac, D., Puspita, M. & Borgougnon, N. 2014. Antibacterial and Antioxidant Properties of The Red Alga *Gracilaria verrucosa* from the North Coast of Java, Semarang, Indonesia. *Int. J. Latest Res. Sc. Technol.*, 3(3):179-185.
- Widyawati, Patang, & Mustarin, A. 2019. The Influence of Various Nitrogen Sources in Increasing the Growth of *Euचेuma cottonii* Seaweed which is Maintained in Controlled Containers. *J. Pendidikan Teknologi Pertanian*, 5(2):93-99

Yang, T.H., Yao, H.T. & Chiang, M.T. 2017. Red algae (*Gelidium amansii*) hot-water extracts ameliorates lipid metabolism in hamsters fed a high-fat diet. *J. Food Drug. Anal.* 25:931-8.

Yong, W.T.L., Ting, S.H., Chan, W. L., Rodrigues, K.F. & Anton, A. 2011. In Vitro Micropropagation of *Euchema* Seaweed. In S. Baby, & Y. and (Eds.), *2nd International Conference on Biotechnology and Food Science*. Bali, Indonesia. 58-60 pp.

30-DOMESTICATION_RED SEAWEED_ANDRE-TJAHJO

ORIGINALITY REPORT

12%

SIMILARITY INDEX

9%

INTERNET SOURCES

8%

PUBLICATIONS

4%

STUDENT PAPERS

PRIMARY SOURCES

| | | |
|---|---|----|
| 1 | www.semanticscholar.org Internet Source | 3% |
| 2 | Submitted to Universitas Diponegoro Student Paper | 3% |
| 3 | www.bioflux.com.ro Internet Source | 2% |
| 4 | Submitted to Defense University Student Paper | 1% |
| 5 | "Encyclopedia of Marine Biotechnology", Wiley, 2020 Publication | 1% |
| 6 | Annita Sari, Ambo Tuwo, Chair Rani, Amran Saru. "Identification and composition of fish types in the Youtefa bay tourism area", IOP Conference Series: Earth and Environmental Science, 2020 Publication | 1% |
| 7 | ojs.omniakuatika.net Internet Source | 1% |

8

Munawan, M Kasim, Ruslaini. "Growth rate of Eucheuma denticulatum cultivated in horizontal net and vertical net", IOP Conference Series: Earth and Environmental Science, 2021

Publication

1 %

9

bioresources.cnr.ncsu.edu

Internet Source

1 %

Exclude quotes On

Exclude matches < 1%

Exclude bibliography On