

# Biotechnologically potential genes in a polysaccharide-degrading epibiont of the Indonesian brown algae *Hydroclathrus* sp. *by Oedjijono Oedjijono*

---

**Submission date:** 17-Feb-2023 04:29PM (UTC+0700)

**Submission ID:** 2016425996

**File name:** Biotechnology\_potential\_genes.....pdf (1.93M)

**Word count:** 9638

**Character count:** 55402

RESEARCH

Open Access

25

# Biotechnologically potential genes in a polysaccharide-degrading epibiont of the Indonesian brown algae *Hydroclathrus* sp.



Stalis Norma Ethica<sup>1†</sup>, Dewi Seswita Zilda<sup>2†</sup>, Oedjijono Oedjijono<sup>3</sup>, Muhtadi Muhtadi<sup>4</sup>, Gintung Patantis<sup>5</sup>, Sri Darmawati<sup>1</sup>, Sri Sinto Dewi<sup>6</sup>, Agus Sabdon<sup>7</sup> and Agustinus Robert Uria<sup>8,9\*</sup>

## Abstract

**Background** Marine bacteria have recently attracted increasing attention to be harnessed for the production of valuable enzymes, vitamins, and bioactive compounds. Bacteria associated with the surfaces of marine macroalgae, called epibionts, are particularly interesting from ecological and biotechnological points of view, as they often exhibit antimicrobial activities to compete with pathogenic bacteria for nutrients and spaces. In search for biotechnologically potential genes from marine bacteria, we sequenced and analysed the genome of the epibiont HI03-3b, a polysaccharide-degrading bacterium associated with the surface of the Indonesian brown algae *Hydroclathrus* sp.

**Results** The algal epibiont HI03-3b has a genome of approximately 4,860,704 bp in size with 42.02 mol% G + C content, consisting of 5655 open reading frames (ORFs), 4409 genes coding for proteins (CDSs), 94 genes for tRNAs, and 32 genes for rRNAs. The genome sequence of HI03-3b was most closely related to that of *Cytobacillus firmus* NCTC10335 with the average amino acid identity (AAI) of 95.0%, average nucleotide identity (ANI) of 94.1%, and a recommended DNA-DNA hybridization (DDH) of 57.60%. These scores are lower than the most frequently used standard for species demarcation (95% ANI cutoff) and the new species threshold (DDH > 70.0% for the same bacterial species). Some differences in genome features and gene composition were observed between HI03-3b and NCTC10335, such as genes encoding carbohydrate active enzymes. These suggest that HI03-3b is unique and likely a novel species within *Cytobacillus* genus and we therefore proposed its name as *Cytobacillus wakatobiense* HI03-3b. Genome sequence analyses indicated the presence of genes involved not only in polysaccharide and protein degradation but also in vitamin and secondary metabolite biosynthesis. Some of them encode enzymes and compounds with biotechnological interest, such as protease, chitinase, subtilisin, pullulanase, and bacillolysin, which are often associated with antimicrobial or antibiofilm activities. This antimicrobial potential is supported by our finding that the extracellular protein fraction of this epibiont inhibited the growth of the bacterial pathogen *Staphylococcus aureus*.

**Conclusion** The epibiont *Cytobacillus* HI03-3b harbours genes for polysaccharide and protein degradation as well as for natural product biosynthesis, suggesting its potential ecological roles in outcompeting other bacteria during biofilm formation as well as in protecting its algal host from predation. Due to the presence of genes for vitamin biosynthesis, it might also provide the algal host with vitamins for growth and development. Some of these metabolic genes are biotechnologically important, as they could become a platform for bioengineering to generate various seaweed-derived substances sustainably, such as antibiofilm agents and vitamins, which are beneficial for human health.

<sup>†</sup>Stalis Norma Ethica and Dewi Seswita Zilda contributed equally to this work.

\*Correspondence:

Agustinus Robert Uria

[a.uria@pharm.hokudai.ac.jp](mailto:a.uria@pharm.hokudai.ac.jp); [agustinus.uria@gmail.com](mailto:agustinus.uria@gmail.com)

Full list of author information is available at the end of the article

**Keywords** Marine macroalgae, Bacterial epibiont, Genome sequencing, Biotechnologically potential genes, Ecological role, Antimicrobial activity

## Background

Marine macroalgae, also called seaweeds, are valuable sources of health-promoting secondary metabolites, enzymes, and vitamins for being developed as nutraceuticals, pharmaceuticals, and cosmeceuticals [24, 36, 42]. Particularly, enzymes from seaweeds have attracted increasing attention for biotechnological applications due to their unique features [55]. However, the natural purification of algae-derived enzymes is extremely difficult due to their high content of polysaccharides, polyphenols, and stable cell walls [45]. Furthermore, the limited supply of seaweed-derived bioactive compounds and enzymes represents a big challenge in their development into high-value marketable products, because most existing cultivation techniques developed for producing commoditized biomass may not necessarily be optimized for seaweed bioactive production [24].

Microorganisms associated with seaweeds have recently been recognized as the producers of novel bioactive compounds and enzymes [40]. Their ability to produce bioactives is considered as an ecological strategy to compete for nutrients and space on the surfaces of marine macroalgae [18]. This strategy also helps their algal hosts to chemically defend against the secondary colonization by other microscopic and macroscopic epibionts [17]. A notable example of epibionts that play this crucial ecological role is *Pseudoalteromonas* species inhabiting seaweed surfaces, as they produce toxic compounds, bacteriolytic substances, and extracellular enzymes for outcompeting other bacteria during biofilm formation [26, 27]. Continuous attempts to isolate potential seaweed-associated bacteria for identifying biotechnologically relevant genes are urgently needed to produce bioactive compounds and enzymes with biotechnological interest in sustainable ways.

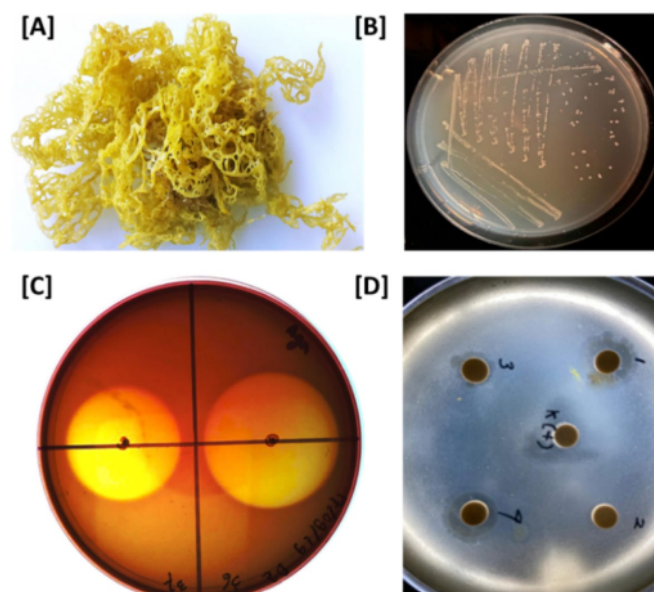
In search for biotechnologically potential genes from seaweed epibionts, we initially isolated a novel polysaccharide-degrading bacterial species associated with the Indonesian brown algae *Hydroclathrus* sp. [70]. We found that the cell-free culture of this epibiont was able to inhibit the bacterial pathogen *S. aureus*, indicating its potential ability to produce antimicrobial substance extracellularly. This preliminary result encouraged us to sequence the whole genome of this bacterium in order to better understand its ecological role and biotechnological potential. Analysing the HI03-3b genome sequence

has enabled us to identify metabolic genes, including those involved in polysaccharide and protein degradation as well as in natural product biosynthesis. Since polysaccharides and proteins represent key components of the extracellular polymeric substances (EPS) of pathogenic microbial biofilms [57], this finding could become a basis for further exploring antimicrobial enzymes and compounds to treat persistent pathogenic biofilms. Subsequent heterologous expression of these genes is necessary to produce useful enzymes or compounds in sustainable ways for biotechnological applications.

## Methods

### Isolation and bioassay of a potential algae-associated epibiont

A polysaccharide-degrading bacterium, designated as HI03-3b, was isolated from the brown algae *Hydroclathrus* sp. inhabiting sandy shallow water around Hoga Island, Wakatobi, South Sulawesi, Indonesia, at the site coordinate of 5.28317 S and 123.45377 E. Briefly, the algae sample was collected at the depth of 0.5–1.0 m (Fig. 1A). It was placed in a 50-ml conical tube and stored in a cool box. A total of 5 g seaweed sample was put into a bottle containing sterile seawater and vortexed for approximately 10 min to release the bacterial cells attached on the surface of *Hydroclathrus* sp. Aliquot of the seaweed mixture was inoculated into solid minimal seawater (MS) media by dilution ( $10^{-5}$ – $10^{-8}$ ) and then incubated for 3–7 days at 30 °C. The MS medium was made up of 0.5% tryptone, 0.1% yeast extract, 0.1% sodium alginate, and 2% agar dissolved in seawater (30 ppt). The colonies growing on solid MS were isolated and purified. The ability of the bacterial isolates to degrade polysaccharides was assessed by a 24-h incubation and subsequent staining with Lugol's iodine solution, followed by staining with  $\text{CaCl}_2$  solution (10%) with 1-h incubation to observe the capability of degrading alginate [70]. The most potential bacterial isolate was subsequently tested for the antimicrobial ability of its extracellular fraction against *Staphylococcus aureus* ATCC®25923™, an opportunistic bacterial pathogen that can cause various infections and food poisoning [14, 63]. Briefly, the cell-free culture was initially prepared and concentrated through a 10-kDa membrane filtration (Whatman). The concentrated extracellular fraction (50 µl) was applied on a paper disc placed on the solid medium streaked with *S. aureus*.



**Fig. 1** Isolation of the potential algae-associated epibiont HI03-3b. The Indonesian brown algae *Hydroclathrus* sp. (photo taken by S. N. Ethica) where HI03-3b was isolated (A) and colony morphology of HI03-3b (B). Bioassays of HI03-3b showing a clearing zone surrounding colonies (duplo) on an alginate-containing MS plate after Lugol staining (C) and inhibition zone by the extracellular culture against the lawn bacterial pathogen *S. aureus* (D)

### Whole-genome sequencing

The genome of *Cytobacillus* sp. HI03-3b was sequenced using Oxford Nanopore Technology (ONT) carried out at the PT Genetika Science Indonesia, a certified service provider of ONT in Indonesia. Briefly, the HI03-3b genomic DNA was initially extracted using QIAGEN Genomic-tip 500/G followed by size selection with Agencourt AMPure XP beads and Circulomics Short Read Eliminator Kit. The libraries of HI03-3b genomic DNA for sequencing were subsequently constructed using Ligation Sequencing Kit (SQK-LSK109) according to the instructions of the manufacturer Oxford Nanopore [41], UK. In principle, the size-selected DNA fragments were end prepped using Ultra II End-prep enzyme mix and ligated with barcode adapter. With adapter-ligated DNA as the template, a barcoding PCR reaction was set up in LongAmp Taq 2× master mix. The resulting pooled barcoded libraries were then end/nick repaired and dA tailed using the NEBNext End Repair/dA-tailing module. The DNA concentration in each library preparation step was measured using Qubit fluorometer based on Qubit dsDNA HS (High Sensitivity) assay (Invitrogen™). The product of each reaction step was washed with 70% EtOH with the help of magnetic separator. The prepared library was loaded into the MinION™ Flow Cells on a ONT MinION sequencing device (Oxford Nanopore

Ltd., UK). V14 kit chemistry in combination with the new R10.4.1 nanopore was used to provide Q20+ ( $\geq 99\%$ ) raw read accuracy with high sequencing yield. FastQC version 0.11.9 (written by Simon Andrews of Babraham Bioinformatics) was subsequently run to do control checks per base sequence quality data.

### Genome sequence assembly

Genome sequence assembly was carried out according to the main steps summarized in Fig. S1A. Bacterial genome sequence datasets were initially assembled using Flye Assembler Version 2.9, a de novo assembler for single-molecule sequencing reads [31, 56]. The Flye assemblies were subsequently polished with one-round Medaka (<https://nanoporetech.com>) for error correction to prep high-quality genome sequences (available online: <https://github.com/nanoporetech/medaka>). Quality parameters of the assembled HI03-3b genome sequence using QUAST [22] (Galaxy Version 5.2.0+galaxy1). The parameters were set up with the minimum IDY% considered as proper alignment of 95.0 and the lower threshold for a contig length (in bp) of 500. The GC% content and read count of HI03-3b genomic sequence were determined using RSeQC (v 2.6.4) [67]. Taxonomic distribution analysis was conducted using MyTaxa Scan result from MiGA to determine the



degree of affiliation or novelty of sequences based on the genome-aggregate average amino acid identity [38, 52]. The order and direction of contigs generated after genome sequence assembly were determined based on blastn pairwise alignment [2] and subsequently verified by average nucleotide identity (ANI) analysis [29] on PROKSEE with CGView Server [58, 59] using the complete genome sequences of closely related taxa as the references.

#### Taxa novelty analysis

HI03-3b genomic sequence was subjected to MiGA (Microbial Genomes Atlas) [52] (version v1.0.0 — prima 14 April 2021) against all taxonomically classified taxa with available genome sequence data for determining its taxonomic classification (<http://microbial-genomes.org/>). This was based on average nucleotide identity and amino acid identity (ANI/AAI) concepts. ANI is a whole-genome similarity metric, which can facilitate high-resolution taxonomic analysis. In taxonomic studies, the standard for species demarcation is the 95% ANI cutoff [29]. HI03-3b genome sequence was compared to the genome sequences of closely related taxa using Mauve [13] (version 43 snapshot 2015 February 25 build 0 (c) 2003–2015). Genome-to-Genome Distance Calculator (GGDC) 3.0 using a generalized linear model (GLM) [41] was run to confirm similarity level between the genome sequences of HI03-3b and the most closely related species. This analysis outcome was based on DNA-DNA hybridization (DDH) values to determine relatedness between bacterial species [21]. Phylogenetic analysis was conducted in MEGA X [32] based on the unweighted pair-group method with arithmetic mean (UPGMA) as the distance analysis method for constructing a tree [43].

#### Genome sequence annotation

The assembled HI03-3b genome was visualized using PROKSEE on the CGView Server [58, 59] and subsequently annotated with Prokka version 1.1.0 [54], allowing the prediction of the numbers of CDSs (coding sequences) as well as genes for tRNAs and rRNAs (5S, 16S and 23S). These were verified using tRNAscan-SE 2.0 [9, 37] and NCBI record (Ref. Seq.: NZ\_JAK-DDU000000000.1). To predict genomic islands, the entire HI03-3b genome sequence was aligned against the complete genome sequence of the closely related taxon *Cytobacillus oceanisediminis* YPW-V2 [Accession Number: CP015506.1] using IslandViewer 4 [5] with the default parameters described in this link: [www.pathogenomics.sfu.ca/islandviewer/about/](http://www.pathogenomics.sfu.ca/islandviewer/about/). To predict HI03-3b primary metabolisms, all CDSs resulted from GeneMarkS [14] analysis [6] were analysed using KofamKOALA [3] against Kofam, a customized HMM database of KEGG

Orthologs (KOs) [3] combined with BLASTx analysis [2]. Carbohydrate-active enzyme (CAZy) database [8] was used as the reference to identify genes encoding carbohydrate active enzymes on HI03-3b genome sequence. By referring to gene position on contigs based on GeneMarkS analysis [6], biotechnologically potential genes were annotated on the circular HI03-3b genome map using PROKSEE on the CGView Server [58, 59]. Further analysis using AntiSMASH version 6.0 [7] was performed to identify natural product biosynthetic gene clusters (BGCs).

## Results

### Polysaccharide-degrading epibiont with antimicrobial activity

In search for biotechnologically potential genes from marine sources, we screened bacterial isolates from the surface of the Indonesian brown algae *Hydroclathrus* sp. (Fig. 1A). This led to the isolation of a polysaccharide-degrading bacterium, designated as HI03-3b, as indicated by the presence of a clearing zone around its colonies after staining with Lugol's iodine solution and  $\text{CaCl}_2$  solution (10%) (Fig. 1B and C). Interestingly, the cell-free culture of this algae-associated epibiont exhibited inhibition against *S. aureus* (Fig. 1D), indicating its potential ability to produce antimicrobial protein or exopolysaccharide. This subsequently encouraged us to sequence the genome of HI03-3b to identify useful genes that might encode proteins or enzymes with potential antimicrobial properties.

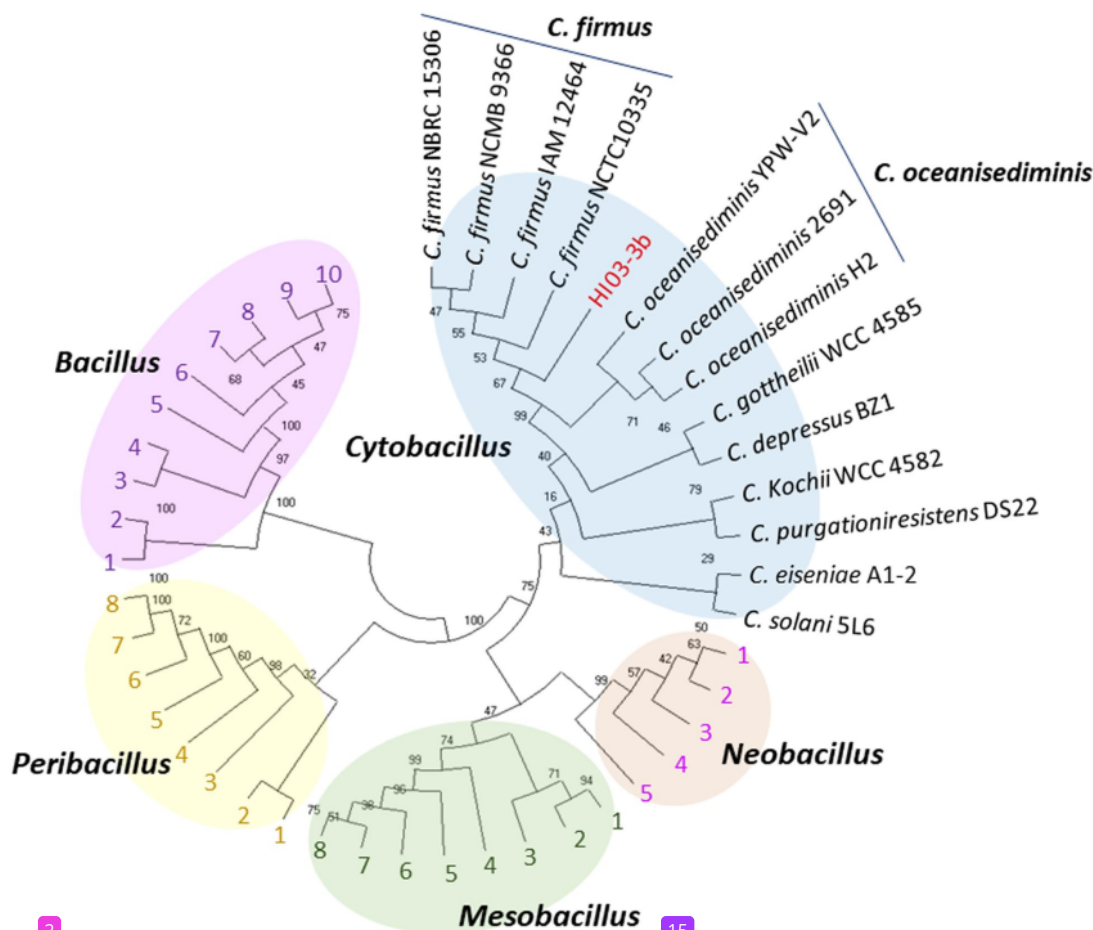
### Genome sequence and taxa novelty analyses

The FastQC analysis (written by Simon Andrews of Babraham Bioinformatics) indicated that the resulting HI03-3b sequencing data contained 1,097,444 sequence reads with the sequence length range of 139 to 14972 bp. All of these sequence reads were assembled into a circular genome of 4,860,704 bp with 42.02 mol% G + C content (Fig. S1B and C). The assembled HI03-3b genome sequence consisted of 11 contigs with the largest size of 2,127,197 bp. The assembly quality was checked using QUAST [22], showing good quality indicated by low values of L50 and L75. Taxonomic distribution analysis based on MyTaxa scan result from MiGA [52] showed the majority of light blue colour (Fig. S2), indicating the high quality of HI03-3b genome sequence with minor contamination.

Blastn search of the entire 16S rRNA gene sequence of HI03-3b showed homology with those from the family Bacillaceae. Further phylogenetic analysis of the HI03-3b's 16S rDNA sequence with those from some representative genera within Bacillaceae (*Bacillus*, *Cytobacillus*, *Mesobacillus*, *Neobacillus*, and *Peribacillus*)

showed that HI03-3b was most closely related to *Cytobacillus* especially within the *C. firmus* clade (Fig. 2). Based on this outcome, we performed genome sequence multiple alignment between HI03-3b and three closely related species (*C. firmus* NCTC10335, *C. oceanisedirminis* YPW-V2, and *C. oceanisedirminis* 2691) using Mauve [13]. It was found some differences in genomic composition among them, as visualized in Fig. S3.

Furthermore, based on MiGA [52], HI03-3b genome sequence showed the most similar ANI (average nucleotide identity) score of 94.1% and AAI (average amino acid identity) score of 95% to those of *C. firmus* NCTC10335 (GenBank assembly accession: GCA900445365). The next top hits were *Sporosarcina globispora* DSM 4 [GenBank assembly accession: GCA001274725.1] and *C. oceanisedirminis* CGMCC 1.10115 [GCA007830235] with



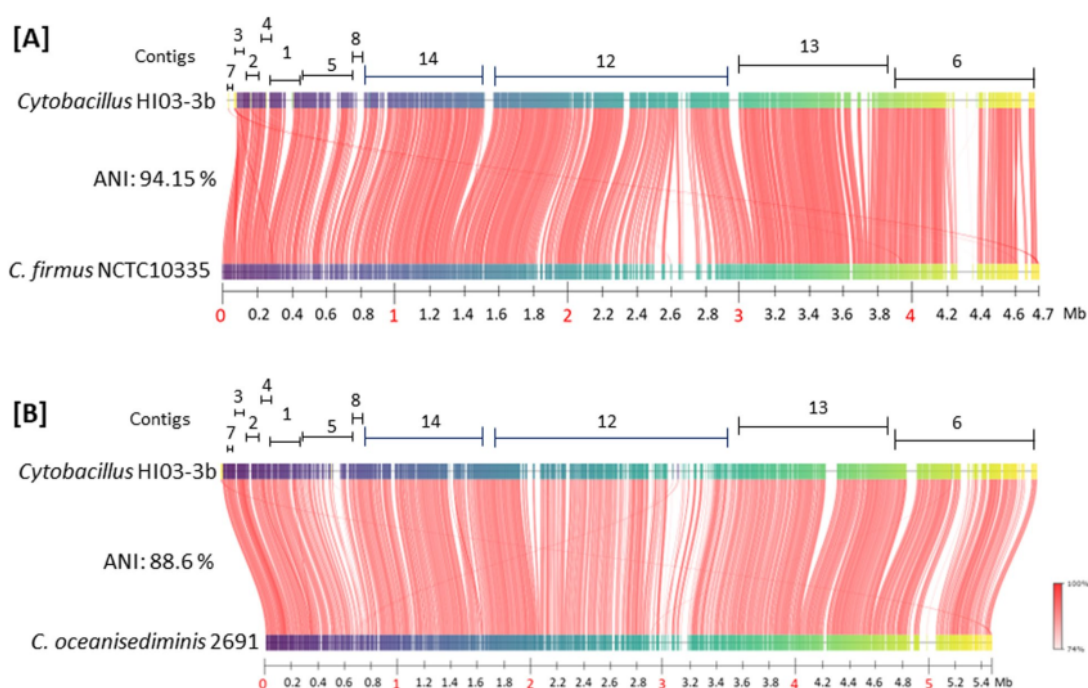
**Fig. 2** The phylogenetic analysis of HI03-3b 16S-rRNA gene sequence using the UPGMA method [43]. The bootstrap consensus tree inferred from 1000 replicates represents the evolutionary history of the taxa analyzed [19]. The evolutionary distances were calculated according to the maximum composite likelihood method [62]. Evolutionary analyses were performed in MEGA X [32], which involved 45 16S-rRNA gene sequences from representative members of some genera within Bacillaceae family. There were a total of 1593 positions in the final dataset. The phylogenetic tree covers representative members of some genera within Bacillaceae family (*Bacillus*, *Cytobacillus*, *Mesobacillus*, *Neobacillus*, and *Peribacillus*). *Neobacillus* clade is as follows: 1, *N. bataviensis* NBRC 102449; 2, *N. drementensis* IDA1967; 3, *N. novalis* NBRC 102450; 4, *N. niacini* NBRC 15566; and 5, *N. pocheonensis* Gsoil 420. *Mesobacillus* clade is as follows: 1, *M. foraminis* CV53; 2, *M. zeae* JJ-247; 3, *M. campisalis* SA2-6; 4, *M. stamsii* BoGlc83; 5, *M. thioparans* BMP-1; 6, *M. boroniphilus* T-152; 7, *M. subterraneus* COOI3B; and 8, *M. jeotgali* YKJ-10. *Peribacillus* clade is as follows: 1, *P. kribbensis* BT080; 2, *P. cavernae* LS; 3, *P. asahii* A001; 4, *P. psychrosaccharolyticus* 23296; 5, *P. muralis* LMG 20238; 6, *P. frigoris* DSM 8801; 7, *P. simplex* NBRC 15720; and 8, *P. simplex* LMG 11160. *Bacillus* clade is as follows: 1, *B. aerophilus* 28K; 2, *B. stratosphericus* 1KF2a; 3, *B. licheniformis* DSM 13; 4, *B. licheniformis* ATCC 14580; 5, *B. velezensis* FZB42; 6, *B. vallismortis* DSM 11031; 7, *B. subtilis* IAM 12118; 8, *B. inaquosorum* BGSC 3A28; 9, *B. halotolerans* DSM 8802; and 10, *B. niacini* IFO15566

the AAI/ANI scores of 88.1%/86.7 and 82.81%/81.46%, respectively. To determine the order and direction of the 11 contigs of HI03-3b genome sequence, we run Blastn [2] for pairwise DNA-DNA sequence comparison using the genome sequences of *C. oceanisedirimens* 2691 and *C. firmus* NCTC10335 as the subject sequences (Table S1). To validate the order and direction of HI03-3b contigs, we then carried out ANI analysis [29] on PROKSEE with CGView Server [58, 59] using *C. oceanisedirimens* 2691 and *C. firmus* NCTC10335 as the references, respectively (Fig. 3).

Finally, we run Genome-to-Genome Distance Calculator (GGDC) 3.0 [41] to confirm the similarity level of the genome sequences between HI03-3b and *C. firmus* NCTC10335 as the closest taxa. The GGDC analysis revealed a recommended a DNA-DNA hybridization (DDH) of 57.60% to *C. firmus* NCTC10335. This value is lower than the new species threshold (DDH > 70.0% for the same species and DDH > 79.0% for the same subspecies). The difference in % G+C between the two genome sequences was 0.30. The genome comparative and phylogenetic studies suggest that HI03-3b is a novel species within *Cytobacillus*, and therefore, we propose the name as *Cytobacillus wakotobiense* HI03-3b (HI refers to Hoga Island, the place where it was derived from).

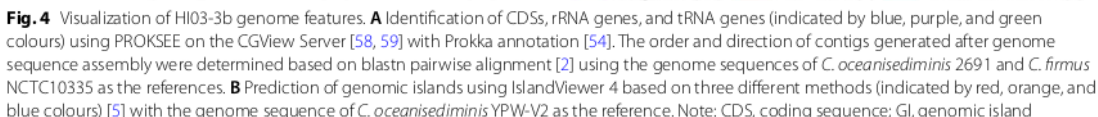
### Genome properties and primary metabolisms

By referring to Blastn comparison and ANI analysis in Fig. 3, the 11 contigs of the assembled HI03-3b genome sequence were visualized in the right order and direction using PROKSEE on the CGView Server [58, 59] with Prokka annotation [54]. The results showed HI03-3b genomic features, such as open reading frames (ORFs), CDSs (coding sequences), 94 genes for tRNAs, and 32 genes for rRNAs (5S, 16S, and 23S) (Fig. 4A). The sequenced HI03-3b genome harboured 5655 ORFs based on GeneMark.hmm PROKARYOTIC Version 3.26 analysis [6] and 4409 CDSs based on the NCBI record (Table S2). Furthermore, tRNAscan-SE 2.0 [9, 37], showed 24 types of tRNAs encoded on HI03-3b genome, which were dominated by tRNAs for Gly, Glu, and Leu (Table S3). Based on KofamKOALA [3], 12 of 32 rRNA genes belong to 16S rRNA genes (Table S4). By aligning with the reference *C. oceanisedirimens* YPW-V2 on IslandViewer 4 [5], we found 13 genomic islands in HI03-3b [21], including polysaccharide biosynthesis gene cluster, based on the integrated methods of SIGI-HMM [66] and IslandPath-DIMOB [28] (Fig. S4B). However, no pathogenic islands/genes were detected within the HI03-3b genome [4].



**Fig. 3** Pairwise genome sequence comparison to determine the order and direction of HI03-3b contigs. **A** HI03-3b contigs were mapped onto with the genome sequence of *C. firmus* NCTC10335 (Acc. Nu. NZ\_UFTC01000001.1) with the ANI score of 94.15 %. **B** HI03-3b contigs were compared with the complete genome sequence of *C. oceanisedirimens* 2691 (NCBI Acc. Nu. GCA\_000294775.2) with the ANI score of 88.6 %





Based on the carbohydrate-active enzyme (CAZy) database [8], it was found that the HI03-3B genome harbours some genes encoding carbohydrate active enzymes (Table S13), including glycoside hydrolase (GHs) and glycosyltransferases (GTs). Some of these genes encoding enzymes that might be related to the degradation of polysaccharides, such as putative type 1 pullulanase (GH13 family), chitinase (GH18 family), and  $\alpha$ -glucosidase

We also found putative genes involved in secondary metabolite biosynthesis (Fig. 5), such as those coding for enzymes responsible for terpenoid backbone biosynthesis predicted using KEGG Orthologs [52, 3]. Further analysis using AntiSMASH version 6.0 [7] allowed us to identify the presence of a biosynthetic gene cluster (BGC) of ribosomally synthesized and post-translationally modified peptides (RiPP)-like terpene in the contig 12 of HI03-3b genome (Fig. 5, Fig. S5 for the detail), which was verified by Bagel4 analysis [15]. Other BGCs identified in this sequenced genome code for polyketide ketosynthase (PKS) type 3 in the contig 13 (Fig. 5, Fig. S6 for the detail),



**Table 1** Some biotechnologically promising genes in HI03-3b, which encode putative enzymes with potential anti-biofilm activities

Gene ID	AA	KEGG Orthology search	BLASTx search		Proposed function
			Protein & acc. no.	I/S (%)	
HI03-309	214	Cyanophycinase [EC:3.4.15.6]	Cyanophycinase [SUW02533.1]	89/95	Cyanophycinase
HI03-932	411	Zinc protease [EC:3.4.24.-]	Insulinase family protein [WP_227887042.1]	99/100	Zinc-dep. metalloproteinase
HI03-1104	1395	Minor extracellular serine protease Vpr [EC:3.4.21.-]	S8 family serine peptidase [WP_197246214.1]	100/100	Serine peptidase
HI03-1355	387	Chitinase [EC:3.2.1.14]	Glycosyl hydrolase family 18 [WP_227887042.1]	98/99	Chitinase [WP_248347211.1]
HI03-1694	593	Bacillolysin [EC:3.4.24.28]	M4 metallopeptidase [WP_222500569.1]	99/99	Bacillolysin [WP_248347527.1]
HI03-2166	449	Serine protease [EC:3.4.21.-]	S8 family peptidase [WP_226617693.1]	99/99	Serine protease [WP_248347905.1]
HI03-2670	521	Pullulanase [EC:3.2.1.41]	Type 1 pullulanase [WP_222500182.1]	99/99	Pullulanase
HI03-3302	306	L-Asparaginase [EC:3.5.1.1]	Asparaginase [WP_222497870.1]	100/100	L-Asparaginase [WP_248348545.1]
HI03-3438	958	Pullulanase [EC:3.2.1.41]	Type 1 pullulanase [WP_222500182.1]	99/99	Pullulanase [WP_248348709.1]
HI03-3561	391	Leader peptide-processing serine protease [EC:3.4.21.-]	S8 family serine peptidase [WP_048011423.1]	99/99	Subtilisin [WP_248348651.1]

Abbreviations: AA amino acid size, I identity, S similarity

siderophore/petrobactin BGC (Fig. 5, Fig. S7), and a lasopeptide BGC in the contig 8 (Fig. S8). Further research is required to isolate secondary metabolites and enzymes from this HI03-3b strain in order to test them against biofilm-forming pathogens.

#### Genome sequence data and strain availability

We deposited our HI03-3b genome sequence in the GenBank database, which was publicly available under the accession number JAKDDU000000000.1 (BioProject ID: PRJNA785558, and BioSample: SAMN23566444). Some of the biotechnologically potential genes in HI03-3b have been annotated by GenBank with the following accession numbers: WP\_248347211.1 (chitinase/glycosyl hydrolase family 18 protein), WP\_248347527.1 (bacillolysin/M4 family metallopeptidase), WP\_248347905.1 (serine protease/S8 family peptidase), WP\_248348545.1 (L-asparaginase), WP\_248348709.1 (pullulanase), WP\_248348651.1 (subtilisin/S8 family serine peptidase), WP\_248349705.1 (L-rhamnose isomerase), and WP\_248349198.1 (nucleoside hydrolase). This HI03-3b strain was registered in National Center for Biotechnology Information (NCBI) with the taxonomy ID 2862822. Authors maintain this HI03-3b strain at the Microbiology Laboratory of Universitas Muhammadiyah Semarang and at National Research and Innovation Agency (BRIN), Indonesia.

#### Discussion

We had isolated a biotechnologically potential bacterial epibiont, designated as HI03-3b, from the surface of the Indonesian brown algae *Hydroclathrus* sp. The ability of this epibiont to degrade complex polysaccharide and to

inhibit a pathogenic gram-positive bacterium had motivated us to sequence the whole genome. HI03-3b genome sequence showed the highest AAI and ANI scores to *C. firmus* NCTC10335, which was supported by 16S rRNA gene phylogenetic analysis. Further genome comparison between HI03-3b and three closely related *Cytobacillus* species, including NCTC10335 strain, suggested some differences in genomic composition among them. Further GGDC analysis indicated that a recommended DNA-DNA hybridization (DDH) between the two strains is lower than the new species threshold (DDH > 70.0% for the same species). The genome features, such as the numbers of protein-coding genes (CDSs), tRNAs, and rRNAs, were different between HI03-3b and *C. firmus* NCTC10335 (Table S2). Taken all together, we propose HI03-3b as a novel species within *Cytobacillus* genus, and therefore, we named it *Cytobacillus wakatobiense* HI03-3b.

The genome sequence analysis provided insights into metabolic pathways, which helped us to understand the ecological role and biotechnological importance of HI03-3b. For example, some genes identified in this HI03-3b genome are involved in the biosynthesis of vitamins, such as thiamine (B1), riboflavin (B2), pantothenate (B5), pyridoxine (B6), biotin (B7), lipoic acid, α-aminobutyrate (B12), and menaquinone (K2) (see Table S11). This indicates that HI03-3b may play a crucial ecological role in providing the algal host with vitamins for growth and development [16]. From biotechnological perspective, the presence of these vitamin biosynthetic pathways suggests that HI03-3b could be harnessed as a natural alternative for industrial vitamin production.

We also found the presence of genes involved in polyamine biosynthesis in HI03-3b, which leads to spermidine form [38] (see Fig. S4 for spermidine biosynthetic pathway). The human gut bacteria *Bacteroides thetaio-**taomicron* and *Fusobacterium varium* represent other bacterial species reported to synthesize spermidine both in vivo and in vitro [49]. From medical perspective, spermidine is known as a bioactive metabolite that can extend life span in model organisms, suggesting its potential application in delaying ageing and promoting longevity in human [39]. Spermidine has particularly been shown to increase epithelial renewal and anti-inflammatory macrophage development in the colon, highlighting its importance in the maintenance of intestinal homeostasis and immunity [46]. From ecological point of view, spermidine was found essential for robust biofilm formation in *Bacillus subtilis* [25]. In addition to spermidine, TatD DNase can contribute to biofilm formation, as has recently shown in the biofilm formation of the bacterium *Trueperella pyogenes* [69]. Interestingly, TatD DNase-encoding gene was found in HI03-3b genome. Taken together, we propose that HI03-b may rely on molecules such as spermidine and TatD DNase to promote its colonization and biofilm formation on the macroalgal surface.

Among 4409 protein-coding sequences identified in this strain, some genes encode enzymes for polysaccharide and protein degradation known to exhibit anti-biofilm properties against pathogenic bacteria [57]. Notable examples are protease [50], chitinase [12], a mixture of bacillolysin and subtilisin (Protamex®) [20, 48], 1,51 paraginase [64, 65], and pullulanase [51] (Table 1). From the ecological point of view, the presence of these genes suggests that HI03-3b may produce antimicrobial or anti-biofilm enzymes in competing for nutrient and space against other epibionts as well as in protecting algal host from predation [17, 18]. This was supported by our finding that the extracellular cell-free supernatant of HI03-3b exhibited antimicrobial activity against the bacterial pathogen *S. aureus*.

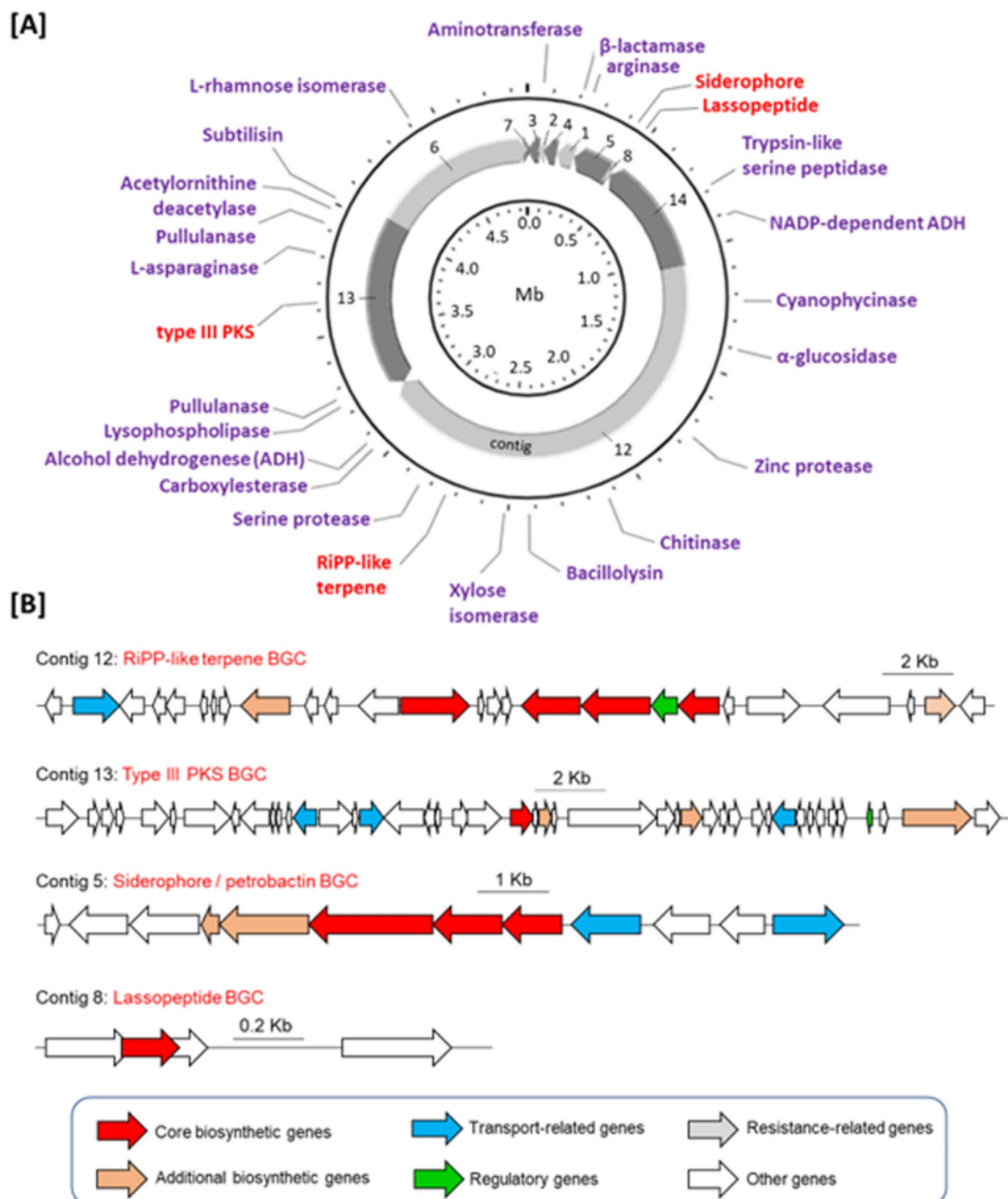
Other biotechnologically potential genes identified in this HI03-3b include those encoding arginase,

xylose isomerase, carboxylesterases, phospholipases, and arginine decarboxylase (see Table S14). Arginase for example has been used for the environmentally friendly preparation of L-ornithine as food supplement and nutrition product [34]. Xylose isomerase has widely been used in the production of high-fructose corn syrup (HFCS) and bioethanol [47]. It is potentially explored to produce some value-added chemicals in the food, cosmetics, and pharmaceutical industries [44]. Carboxylesterases (CEs) have been applied in xenobiotic and endobiotic degradations, biocatalysis, drug metabolism [68]. Phospholipases have been used in scientific and medical research, such as inhibitors for generating anti-inflammatory agents and as diagnostic markers for microbial infections [30, 61]. Arginine decarboxylase catalyses conversion of L-arginine into 29 natine, a valuable pharmaceutical intermediate with various potential therapeutic functions in neurotransmitter systems, nitric oxide synthesis, and polyamine metabolism [60].

HI03-3b genome harbors a RiPP-like terpene BGC predicted to encode the biosynthesis of thiazole-containing heterocyclic bacteriocin as a new subfamily of ribosomally synthesized peptides antimicrobial peptides (AMPs) ([23, 93]). This RiPP-like terpene BGC is characterized by the presence of genes encoding proteins involved in heterocycloanthracin biosynthesis, such as toxin precursor, SagB-type dehydrogenase domain (nitroreductase family), serine protease, YcaO cyclodehydratase, thiazole-containing bacteriocin maturation protein, and a transport protein [23] (Fig. S5). Bacteriocins that belong to heterocycloanthracin family are known as antimicrobial and antibiofilm agents [53], best exemplified by sonorensin that exhibited antibiofilm activity against *S. aureus* and food bio-preservative potential [10, 11]. It was reported that bacteriocins of heterocycloanthracin family are initially synthesized as biologically inactive peptides (protoxins or precursors) with an N-terminal leader peptide. This protoxin subsequently undergoes enzymatic modifications that involve the cleavage of the leader peptide by protease and the formation of a

(See figure on next page.)

**Fig. 5** The position of biotechnologically potential genes on the HI03-3b genome map. **A** Certain loci encoding biotechnologically potential enzymes and secondary metabolite biosynthesis were determined based on PROKSEE analysis on the CGView Server [58, 59]. **B** Natural product BGC [19] are identified in the HI03-3b genome sequence based on antiSMASH analysis [7] supported by BLASTx [2], showing the presence of core and additional biosynthetic genes, transport-related genes, regulatory genes, and other genes. The core biosynthetic genes of RiPP-like terpene BGC (from left to right) are predicted to code for squalene-hopene cyclase, leader peptide (SagB-type dehydrogenase domain), YcaO cyclodehydratase, and thiazole-containing bacteriocin maturation protein. PKS type III BGC contains a chalcone synthase gene (indicated by red colour) as the core biosynthetic gene. The siderophore BGC [17] joins the core biosynthetic genes encoding lucA/lucC family proteins. Based on RiPPMiner prediction [1] the core lassopeptide BGC encodes a precursor peptide that consists of a leader (VKAPGSTGEGHWKLGNSAEKSGIPRVAVKCVEMWRNTSGE) and core peptide sequence [34] (GDSLVCN). A crosslink may occur between serine (S) and cysteine (C) residues in the core peptide [1]. Note: PKS, polyketide synthase; BGC, biosynthetic gene cluster; RiPP, ribosomally synthesized and post-translationally modified peptides



**Fig. 5** (See legend on previous page.)



thiazole or oxazole ring by cyclodehydratase and dehydrogenase. The resulting mature toxin is then exported by a transport protein [23, 33]. Validation of the function of this RiPP-like terpene BGC in HI03-3b through gene mutagenesis or heterologous expression and subsequent identification of the bacteriocin produced are necessary to explore the biotechnological potential as an anti-biofilm and food preservative agent.

We compared the presence of some biotechnologically potential genes between *Cytobacillus* HI03-3b and the most closely related strain *C. firmus* NCTC10335 (Table S15). It was found that several genes encoding carbohydrate active enzyme [24], HI03-3b, such as  $\beta$ -glucosylceramidase [EC:3.2.1.45], xylose isomerase [EC:5.3.1.5], L-rhamnose isomerase [EC:5.3.1.14], chitinase [EC:3.2.1.14], and arginase [EC:3.5.3.1], were absent in NCTC10335. In contrast,  $\alpha$ -amylase [EC:3.2.1.1] identified in NCTC10335 was not found in HI03-3b. Another difference is that HI03-3b contained lassopeptide BGC, while NCTC10335 harboured lanthipeptide class 2. Some potential genes present in both strains occur in different copy numbers (Table S15), such as those encoding oligo-1,6-glucosidase [EC:3.2.1.10],  $\alpha$ -glucosidase [EC:3.2.1.20], zinc protease [EC:3.4.24.-], serine protease [EC:3.4.21.-], bacillolysin [EC:3.4.24.28], beta-lactamase class A [EC:3.5.2.6], and acetylornithine deacetylase [EC:3.5.1.16]. The difference in genome features and gene composition between *Cytobacillus* HI03-3b and its closely related strain strongly suggests the uniqueness of HI03-3b that would enable it to adapt to the algal surface environment. This was supported by the finding of 13 genomic islands in HI03-3b that possibly occur through horizontal gene transfer, which may provide this epibiont with adaptive traits to live and survive on the surface of brown algae [4].

## Conclusion

The whole-genome sequence analysis of the brown algae-associated epibiont *Cytobacillus* sp. HI03-3b showed the presence of biotechnologically potential genes, including those encoding commercially useful enzymes, such as chitinase, pullulanase, protease, bacillolysin, subtilisin, and L-asparaginase, as well as biosynthetic genes for secondary metabolites and vitamins. These might ecologically be important as strategies of the epibiont to outcompete with other bacteria during biofilm formation, protect its algal host from pathogenic infection, and provide the host with necessary vitamins. Some of such metabolic genes could potentially be exploited as a basis for the development of anti-infective agents against pathogenic biofilms. Further functional studies by gene heterologous expression, and mutagenesis or enzyme activity assays, are necessary to validate the function of these genes.

7

## Abbreviations

ORF	Open reading frame
CDS	Coding sequence
8	Minimal seawater
ANI	Average nucleotide identity
AAI	Average amino acid identity
CAZy	Carbohydrate-active enzyme
DDH	DNA-DNA hybridization
BGC	Biosynthetic gene cluster
NP	Natural products

13

## Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s43141-023-00461-5>.

**Additional file 1: Figure S1.** Visualization and quality assessment of whole HI03-3b genome assembly. [A] Workflow of HI03-3b genome sequence assembly. [B] Quality parameters of the assembled HI03-3b genome sequence using Quast [22]. [C] GC content of the assembled HI03-3b genome sequence against density of reads using RSeQC [67]. **Figure S2.** Taxonomic distribution analysis based on MyTaxa Scan result from MiGA ([38, 52]) on the assembled HI03-3b genome sequence, showing that the high quality genome (the major light blue) with minor contamination. **Figure S3.** Genome comparison of HI03-3b with three closely related *Cytobacillus* strains using Mauve [13]. Each colored block represents a region of the HI03-3b genome sequence, which align with parts of other genomes. Blocks above the central line show forward orientation relative to the first genome sequence, while blocks below the central line indicate regions aligning in the reverse complement orientation. Inside each block, the height of the similarity profile correlates with the average level of conservation region of the genome. **Figure S4.** Genes involved in spermidine biosynthesis identified in the HI03-3b genome, which were predicted based on KofamKOALA [3] and BLASTx [2]. Notes: spermidine has been shown to increase epithelial renewal and anti-inflammatory macrophage development in the colon, highlighting its importance in the maintenance of intestinal homeostasis and immunity [46]. From medical perspective, spermidine has been known to extend life span in model organisms, indicating its potential application in delaying aging and promoting longevity in human [39]. **Figure S5.** RiPP-like terpene BGC identified in the HI03-3b genome, which were predicted based on antiSMASH [7] and BLASTx [2]. **Figure S6.** PKS type III BGC identified in the HI03-3b genome, which were predicted based on antiSMASH [7] and BLASTx [2]. **Table S1.** The order and direction of HI03-3b contigs were validated by comparing with the genome sequences of *C. oceanisedirimens* 2691 and *C. firmus* NCTC10335. **Table S2.** Comparison of genomic features between *Cytobacillus* HI03-3b (in this work) and *C. firmus* NCTC10335 (GenBank assembly accession number GCA\_900445365.1). **Table S3.** Estimated numbers of tRNAs in HI03-3b genome sequence analyzed using tRNAscan-SE 2.0. ([9] [37]). **Table S4.** Numbers of 16S-rRNAs in HI03-3b genome sequence based on KofamKOALA [3]. **Table S5.** Predicted carbohydrate metabolisms in HI03-3b based on KofamKOALA [3]. **Table S6.** Energy metabolisms in HI03-3b based on KofamKOALA [3] analysis. **Table S7.** Lipid metabolisms in HI03-3b based on KofamKOALA [3] analysis. **Table S8.** Nucleotide metabolisms in HI03-3b based on KofamKOALA [3] analysis. **Table S9.** Predicted amino acid metabolisms in HI03-3b based on KofamKOALA [3]. **Table S10.** Predicted glycan metabolism in HI03-3b based on KofamKOALA [3]. **Table S11.** Carbon and vitamins metabolisms in HI03-3b based on KofamKOALA [3]. **Table S12.** Predicted terpenoid biosynthesis in HI03-3b based on KofamKOALA [3]. **Table S13.** Some genes encoding carbohydrate active enzymes. **Table S14.** Genes on HI03-3b genome encoding biotechnologically potential enzymes. **Table S15.** Comparison of biosynthetically potential genes between *Cytobacillus* HI03-3b and *C. firmus* NCTC10335.

## Acknowledgements

We acknowledge the Indonesian Ministry of Education, Cultural, Research and Technology (Kemendikbud Ristek) for funding this research project. This work was also supported by National Research and Innovation Agency (BRIN), Republic of Indonesia.

## Authors' contributions

DSZ and SNE conceived and planned the experiments. SD and AS provided project supervision and discussion. DSZ and GP contributed to seaweed sample collection, isolation of bacterial strains and culture preparation for whole-genome sequencing. SNE and OO contributed to the initial version of the manuscript draft. DSZ performed the bioactivity tests. MM and SSD contributed to the interpretation of the results. ARU and SNE analysed the genomic sequence data. ARU improved and finalized the manuscript. The authors read and approved the final manuscript.

## Funding

This work was supported by the Indonesian Ministry of Education, Cultural, Research and Technology (Kemendikbud Ristek) through Basic Science Research Program (PD, Penelitian Dasar) 2021 (Grant ID: 166/E4.I/AK.04.PT/2021) and 2022 (Grant ID: 027/06.10.26/PG/SP2H/TD/2022).

## Availability of data and materials

We deposited our H103-3b genome sequence in the GenBank database, which was publicly available under the accession number JAKDDU000000000.1 (BioProject ID: PRJNA785558 and BioSample: SAMN2356444). Some of the biotechnologically potential genes in H103-3b have been annotated by GenBank with the following accession numbers: WP\_248347211.1 (chitinase/glycosyl hydrolase family 18 protein), WP\_248347527.1 (bacillolysins/M4 family metalloproteinase), WP\_248347905.1 (serine protease/S8 family peptidase), WP\_248348545.1 (L-asparaginase), WP\_248348709.1 (pullulanase), and WP\_248348511.1 (subtilisin/S8 family serine peptidase). This H103-3b strain was registered in National Center for Biotechnology Information (NCBI) with the taxonomy ID 2862822. Authors maintain this H103-3b strain at the Microbiology Laboratory of Universitas Muhammadiyah Semarang and at National Research and Innovation Agency (BRIN), Indonesia.

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

## Author details

<sup>1</sup>Magister Program of Clinical Laboratory Science, Universitas Muhammadiyah Semarang (UNIMUS), Jalan Kedondong Raya, Semarang 50273, Indonesia.

<sup>2</sup>Research Center for Deep Sea, Earth Sciences and Marine Biotechnology, National Research and Innovation Agency (BRIN), Jl. Pasir Putih Raya, Pademangan, North Jakarta City, Jakarta 14430, Indonesia.

<sup>3</sup>Faculty of Biology, Universitas Jenderal Soedirman, Purwokerto 53122, Indonesia.

<sup>4</sup>Faculty of Pharmacy, Universitas Muhammadiyah Surakarta (UMS), Surakarta 57162, Indonesia.

<sup>5</sup>Research Center for Marine and Land Bioindustry, Earth Sciences and Maritime Research Organization, National Research and Innovation Agency (BRIN), Kodek Bay, North Lombok, West Nusa Tenggara 83352, Indonesia.

<sup>6</sup>Diploma Study Program of Medical Laboratory Technology, Faculty of Nursing and Health Sciences, Universitas Muhammadiyah Semarang, Semarang 50273, Indonesia.

<sup>7</sup>Department of Marine Science, Faculty of Fisheries and Marine Science, Diponegoro University, Semarang 50272, Indonesia.

<sup>8</sup>Faculty of Pharmaceutical Sciences, Hokkaido University, Kita 12 Nishi 6 Kita-ku, Sapporo 060-0812, Japan.

<sup>9</sup>Global Station for Biosurfaces and Drug Discovery, Global Institution for Collaborative Research and Education (GI-CoRE), Hokkaido University, Kita 12 Nishi 6, Sapporo 060-0812, Japan.

## References

1. Agrawal P, Khater S, Gupta M, Sain N, Mohanty D (2017) RIPPMiner: a bioinformatics resource for deciphering chemical structures of RPPs based on prediction of cleavage and cross-links. *Nucleic Acids Res* 45(W1):W80–W88. <https://doi.org/10.1093/nar/gkx408>
2. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ (1990) Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2)
3. Aramaki T, Blanc-Mathieu R, Endo H, Ohkubo K, Kanehisa M, Goto S, Ogata H (2020) KofamKOALA: KEGG Ortholog assignment based on profile HMM and adaptive score threshold. *Bioinformatics* 36:2251–2252. <https://doi.org/10.1093/bioinformatics/btz859>
4. Beatriz, Fernández-Gómez Antonio, Fernández-Guerra Emilio O, Casamayor José M, González Carlos, Pedrós-Alíó Silvia G, Acinas (2012) Patterns and architecture of genomic islands in marine bacteria. *BMC Genomics* 13(1):347. <https://doi.org/10.1186/1471-2164-13-347>
5. Bertelli C, Laird MR, Williams KP, Lau BY, Hoad G, Winsor GL, Brinkman FS (2017) IslandViewer 4: expanded prediction of genomic islands for larger-scale datasets. *Nucleic Acids Res* 45:W30–W35. <https://doi.org/10.1093/nar/gkx343>
6. Besemer J, Lomsadze A, Borodovsky M (2001) GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. *Nucleic Acids Res* 29:2607–2618. <https://doi.org/10.1093/nar/29.12.2607>
7. Blin K, Shaw S, Kloosterman AM, Charlop-Powers Z, van Wezel GP, Medema MH, Weber T (2021) antiSMASH 6.0: improving cluster detection and comparison capabilities. *Nucleic Acids Res* 49:W29–W35. <https://doi.org/10.1093/nar/gkab335>
8. Cantarel BL, Coutinho PM, Rancurel C, Bernard T, Lombard V, Henrissat B (2009) The Carbohydrate-Active Enzymes database (CAZY): an expert resource for Glycogenomics. *Nucleic Acids Res* 37:D233–D238. <https://doi.org/10.1093/nar/gkn663>
9. Chan PP, Lin BY, Mak AJ, Lowe TM (2021) tRNAscan-SE 2.0: improved detection and functional classification of transfer RNA genes. *Nucleic Acids Res* 49:9077–9096. <https://doi.org/10.1093/nar/gkab688>
10. Chopra L, Singh G, Choudhary V, Sahoo DK (2014) Sonorensin: an antimicrobial peptide, belonging to the heterocycloanthracin subfamily of bacteriocins, from a new marine isolate, *Bacillus sonorensis* MT93. *Appl Environ Microbiol* 80:2981–2990. <https://doi.org/10.1128/AEM.04259-13>
11. Chopra L, Singh G, Kumar Jena K, Sahoo DK (2015) Sonorensin: a new bacteriocin with potential of an anti-biofilm agent and a food bio-preservative. *Sci Rep* 5:13412. <https://doi.org/10.1038/srep13412>
12. Chung M-C, Dean S, Marakasova ES, Nwabueze AO, van Hoek ML (2014) Chitinases are negative regulators of *Francisella novicida* biofilms. *PLoS One* 9:e93119. <https://doi.org/10.1371/journal.pone.0093119>
13. Darling ACE, Mau B, Blattner FR, Perna NT (2004) Mauve: multiple alignment of conserved genomic sequence with rearrangements. *Genome Res* 14:1394–1403. <https://doi.org/10.1101/gr.2289704>
14. David MZ, Daum RS (2010) Community-associated methicillin-resistant *Staphylococcus aureus*: epidemiology and clinical consequences of an emerging epidemic. *Clin Microbiol Rev* 23:616–687. <https://doi.org/10.1128/CMR.00081-09>
15. de Jong A, van Hijum SAFT, Bijlsma JJE, Kok J, Kuipers OP (2006) BAGEL: a web-based bacteriocin genome mining tool. *Nucleic Acids Res* 34:W273–W279. <https://doi.org/10.1093/nar/gkl237>
16. Droop MR (2007) Vitamins, phytoplankton and bacteria: symbiosis or scavenging? *J Plankton Res* 29:107–113. <https://doi.org/10.1093/plankt/fbm009>
17. Egan S, Harder T, Burke C, Steinberg P, Kjelleberg S, Thomas T (2013) The seaweed holobiont: understanding seaweed–bacteria interactions. *FEMS Microbiol Rev* 37:462–476. <https://doi.org/10.1111/1574-6976.12011>
18. Egan S, Thomas T, Kjelleberg S (2008) Unlocking the diversity and biotechnological potential of marine surface associated microbial communities. *Curr Opin Microbiol Ecol Ind* 11:219–225. <https://doi.org/10.1016/j.mib.2008.04.001>
19. Felsenstein J (1985) Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39:783–791. <https://doi.org/10.1111/j.1558-5646.1985.tb00420.x>
20. Fernandes P (2016) Enzymes in Fish and Seafood Processing. *Front Bioeng Biotechnol* 4:59. <https://doi.org/10.3389/fbioe.2016.00059>

Received: 19 August 2022 Accepted: 6 January 2023  
Published online: 14 February 2023

21. Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, Tiedje JM (2007) DNA-DNA hybridization values and their relationship to whole-genome sequence similarities. *Int J Syst Evol Microbiol* 57:81–91. <https://doi.org/10.1099/ijs.0.64483-0>
22. Gurevich A, Saveliev V, Vyahhi N, Tesler G (2013) QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>
23. Haft DH (2009) A strain-variable bacteriocin in *Bacillus anthracis* and *Bacillus cereus* with repeated Cys-Xaa-Xaa motifs. *Biol Direct* 4:15. <https://doi.org/10.1186/1745-6150-4-15>
24. Hafting JT, Craigie JS, Stengel DB, Loureiro RR, Buschmann AH, Yarish C, Edwards MD, Critchley AT (2015) Prospects and challenges for industrial production of seaweed bioactives. *J Phycol* 51:821–837. <https://doi.org/10.1111/jpy.12326>
25. Hobbey L, Li B, Wood JL, Kim SH, Naidoo J, Ferreira AS, Khomutov M, Khomutov A, Stanley-Wall NR, Michael AJ (2017) Spermidine promotes *Bacillus subtilis* biofilm formation by activating expression of the matrix regulator *slrR*. *J Biol Chem* 292:12041–12053. <https://doi.org/10.1074/jbc.M117.789644>
26. Holmström C, Egan S, Franks A, McCloy S, Kjelleberg S (2002) Antifouling activities expressed by marine surface associated *Pseudoalteromonas* species. *FEMS Microbiol Ecol* 41:47–58. <https://doi.org/10.1111/j.1574-6941.2002.tb00965.x>
27. Holmström C, Kjelleberg S (1999) Marine *Pseudoalteromonas* species are associated with higher organisms and produce biologically active extracellular agents. *FEMS Microbiol Ecol* 30:285–293. <https://doi.org/10.1111/j.1574-6941.1999.tb00656.x>
28. Hsiao W, Wan I, Jones SJ, Brinkman FSL (2003) IslandPath: aiding detection of genomic islands in prokaryotes. *Bioinformatics* 19:418–420. <https://doi.org/10.1093/bioinformatics/btg004>
29. Jain C, Rodríguez-R LM, Phillippy AM, Konstantinidis KT, Aluru S (2018) High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nat Commun* 9:5114. <https://doi.org/10.1038/s41467-018-07641-9>
30. Karabina S-A, Gora S, Atout R, Ninio E (2010) Extracellular phospholipases in atherosclerosis. *Biochimie Phospholipases A2 Lipid Mediators* 92:594–600. <https://doi.org/10.1016/j.biochi.2010.02.002>
31. Kolmogorov M, Yuan J, Lin Y, Pevzner PA (2019) Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>
32. Kumar S, Stecher G, Li M, Niyaz C, Tamura K (2018) MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms. *Mol Biol Evol* 35:1547–1549. <https://doi.org/10.1093/molbev/msy096>
33. Lee SW, Mitchell DA, Markley AL, Hensler ME, Gonzalez D, Wohlrab A, Dorrestein PC, Nizet V, Dixon JE (2008) Discovery of a widely distributed toxin biosynthetic gene cluster. *Proc Natl Acad Sci U S A* 105:5879–5884. <https://doi.org/10.1073/pnas.0801338105>
34. Li M, Qin J, Xiong K, Jiang B, Zhang T (2021) Review of arginase as a promising biocatalyst: characteristics, preparation, applications and future challenges. *Crit Rev Biotechnol* 0:1–17. <https://doi.org/10.1080/07388551.2021.1947962>
35. Liu X, Kokare C (2017) Chapter 11 - Microbial enzymes of use in industry. In: Brahmachari G (ed) *Biotechnology of Microbial Enzymes*. Academic Press (Elsevier), London, UK, pp 267–298. <https://doi.org/10.1016/B978-0-12-803725-6.00011-X>
36. López-Hortas L, Flórez-Fernández N, Torres MD, Ferreira-Anta T, Casas MP, Balboa EM, Falqué E, Domínguez H (2021) Applying seaweed compounds in cosmetics, cosmeceuticals and nutraceuticals. *Mar Drugs* 19:552. <https://doi.org/10.3390/md19100552>
37. Lowe TM, Chan PP (2016) tRNAscan-SE On-line: integrating search and context for analysis of transfer RNA genes. *Nucleic Acids Res* 44:W54–W57. <https://doi.org/10.1093/nar/gkw413>
38. Luo C, Rodríguez-R LM, Konstantinidis KT (2014) MyTaxa: an advanced taxonomic classifier for genomic and metagenomic sequences. *Nucleic Acids Res* 42:e73. <https://doi.org/10.1093/nar/gku169>
39. Madeo F, Carmona-Gutierrez D, Kepp O, Kroemer G (2018) Spermidine delays aging in humans. *Aging (Albany NY)* 10:2209–2211. <https://doi.org/10.18632/aging.101517>
40. Martin M, Portetelle D, Michel G, Vandenbol M (2014) Microorganisms living on macroalgae: diversity, interactions, and biotechnological applications. *Appl Microbiol Biotechnol* 98:2917–2935. <https://doi.org/10.1007/s00253-014-5557-2>
41. Meier-Kolthoff JP, Carlsbe JS, Peinado-Olarte RL, Göker M (2022) TYGS and LPSN: a database tandem for fast and reliable genome-based classification and nomenclature of prokaryotes. *Nucleic Acids Res* 50:D801–D807. <https://doi.org/10.1093/nar/gkab902>
42. Menaa F, Wijesinghe PAUJ, Thirupuranathar G, Uzair B, Iqbal H, Khan BA, Menaa B (2020) Ecological and industrial implications of dynamic seaweed-associated microbiota interactions. *Mar Drugs* 18:641. <https://doi.org/10.3390/md18120641>
43. Michener CD, Sokal RR (1957) A quantitative approach to a problem in classification. *Evolution* 11:130–162. <https://doi.org/10.1111/j.1558-5646.1957.tb02884.x>
44. Miyamoto RY, de Melo RR, de Mesquita Sampaio IL, de Sousa AS, Moraes ER, Sargo CR, Zanphorlin LM (2021) Paradigm shift in xylose isomerase usage: a novel scenario with distinct applications. *Crit Rev Biotechnol* 1–20. <https://doi.org/10.1080/07388551.2021.1962241>
45. Mücke M, Taube R, Veltel S (2022) Purification of native enzymes from macroalgae. *Wiley Analytical Science Magazine: Separation LC Electrophoresis*. Wiley, Weinheim, Germany. <https://analyticalscience.wiley.com/doi/10.1002/was.0004000219>
46. Nakamura A, Kurihara S, Takahashi D, Ohashi W, Nakamura Y, Kimura S, Onuki M, Kume A, Sasazawa Y, Furusawa Y, Obata Y, Fukuda S, Saiki S, Matsumoto M, Hase K (2021) Symbiotic polyamine metabolism regulates epithelial proliferation and macrophage differentiation in the colon. *Nat Commun* 12:2105. <https://doi.org/10.1038/s41467-021-22212-1>
47. Nam KH (2022) Glucose isomerase: functions, structures, and applications. *Appl Sci* 12:428. <https://doi.org/10.3390/app12010428>
48. Nguyen HTM, Sylla KSB, Randriamahatody Z, Donnay-Moreno C, Moreau J, Tran LT et al (2011) Enzymatic hydrolysis of yellowfin tuna (*Thunnus albacares*) by-products using protamex protease. *Food Technol Biotechnol* 49:48–55
49. Noack J, Dongowski G, Hartmann L, Blaut M (2000) The human gut bacteria *Bacteroides thetaiotaomicron* and *Fusobacterium varium* produce putrescine and spermidine in cecum of pectin-fed gnotobiotic rats. *J Nutr* 130:1225–1231. <https://doi.org/10.1093/jn/130.5.1225>
50. Park J-H, Lee J-H, Cho MH, Herzberg M, Lee J (2012) Acceleration of protease effect on *Staphylococcus aureus* biofilm dispersal. *FEMS Microbiol Lett* 335:31–38. <https://doi.org/10.1111/j.1574-6968.2012.02635.x>
51. Petrucci B, Briggs RE, Swords WE, De Castro C, Molinaro A, Inzana TJ (2017) Capsular polysaccharide interferes with biofilm formation by *Pasteurella multocida* serogroup A. *mBio* 8:e01843–e01817. <https://doi.org/10.1128/mBio.01843-17>
52. Rodríguez-R LM, Gunturu S, Harvey WT, Rosselló-Mora R, Tiedje JM, Cole JR, Konstantinidis KT (2018) The Microbial Genomes Atlas (MiGA) web-server: taxonomic and gene diversity analysis of Archaea and Bacteria at the whole genome level. *Nucleic Acids Res* 46:W282–W288. <https://doi.org/10.1093/nar/gky467>
53. Santos VL, Nardi Drummond RM, Dias-Souza MV (2017) 16 - Bacteriocins as antimicrobial and antibiofilm agents. In: Thomaz-Soccol V, Pandey A, Resende RR (eds) *Current Developments in Biotechnology and Bioengineering*. Elsevier, Amsterdam, pp 403–436. <https://doi.org/10.1016/B978-0-444-63660-7.00016-4>
54. Seemann T (2014) Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>
55. Sekiguchi J, Matsumiya M, Mochizuki A (1995) Distribution of chitinolytic enzymes in seaweeds. *Fish Sci* 61:876–881. <https://doi.org/10.2331/fishsci.61.876>
56. Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM (2015) BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* 31:3210–3212. <https://doi.org/10.1093/bioinformatics/btv351>
57. Stiefel P, Mauerhofer S, Schneider J, Maniura-Weber K, Rosenberg U, Ren Q (2016) Enzymes enhance biofilm removal efficiency of cleaners. *Antimicrob Agents Chemother*. <https://doi.org/10.1128/AAC.00400-16>
58. Stothard P, Grant JR, Van Domselaar G (2017) Visualizing and comparing circular genomes using the CGView family of tools. *Brief Bioinform* 20:1576–1582. <https://doi.org/10.1093/bib/bbx081>



59. Stothard P, Wishart DS (2005) Circular genome visualization and exploration using CGView. *Bioinformatics* 21:537–539. <https://doi.org/10.1093/bioinformatics/bti054>
60. Sun A, Song W, Qiao W, Chen X, Liu J, Luo Q, Liu L (2017) Efficient agmatine production using an arginine decarboxylase with substrate-specific activity. *J Chem Technol Biotechnol* 92:2383–2391. <https://doi.org/10.1002/jctb.5245>
61. Sutto-Ortiz P, Camacho-Ruiz M d l A, Kirchmayr MR, Camacho-Ruiz RM, Mateos-Díaz JC, Noiriél A, Carrière F, Abousalham A, Rodríguez JA (2017) Screening of phospholipase A activity and its production by new actinomycete strains cultivated by solid-state fermentation. *PeerJ* 5:e3524. <https://doi.org/10.7717/peerj.3524>
62. Tamura K, Nei M, Kumar S (2004) Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proc Natl Acad Sci* 101:11030–11035. <https://doi.org/10.1073/pnas.0404206101>
63. Treangen TJ, Maybank RA, Enke S, Friss MB, Diviak LF, Karaolis DKR, Koren S, Ondov B, Phillippy AM, Bergman NH, Rosovitz MJ (2014) Complete genome sequence of the quality control strain *Staphylococcus aureus* subsp. *aureus* ATCC 25923. *Genome Announc* 2:e011110–e011114. <https://doi.org/10.1128/genomeA.01110-14>
64. Vimal A, Kumar A (2021) Antimicrobial potency evaluation of free and immobilized L-asparaginase using chitosan nanoparticles. *J Drug Deliv Sci Technol* 61:102231. <https://doi.org/10.1016/j.jddst.2020.102231>
65. Vimal A, Kumar A (2022) L-asparaginase: need for an expedition from an enzymatic molecule to antimicrobial drug. *Int J Pept Res Ther* 28:9. <https://doi.org/10.1007/s10989-021-10312-x>
66. Waack S, Keller O, Asper R, Brodag T, Damm C, Fricke WF, Surovcik K, Meinicke P, Merkl R (2006) Score-based prediction of genomic islands in prokaryotic genomes using hidden Markov models. *BMC Bioinformatics* 7:142. <https://doi.org/10.1186/1471-2105-7-142>
67. Wang L, Wang S, Li W (2012) RSeQC: quality control of RNA-seq experiments. *Bioinformatics* 28:2184–2185. <https://doi.org/10.1093/bioinformatics/bts356>
68. Wheelock CE, Shan G, Ottea J (2005) Overview of carboxylesterases and their role in the metabolism of insecticides. *J Pestic Sci* 30:75–83. <https://doi.org/10.1584/jpestics.30.75>
69. Zhang Z, Liang Y, Yu L, Chen M, Guo Y, Kang Z, Qu C, Tian C, Zhang D, Liu M (2021) TatD DNases contribute to biofilm formation and virulence in *Trueperella pyogenes*. *Front Microbiol* 12:758465. <https://doi.org/10.3389/fmicb.2021.758465>
70. Zilda DS, Yulianti Y, Sholihah RF, Subaryono S, Fawzya YN, Irianto HE (2019) A novel *Bacillus* sp. isolated from rotten seaweed: identification and characterization alginate lyase its produced. *Biodivers J Biol Divers* 20:1166–1172. <https://doi.org/10.13057/biodiv/d200432>

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Submit your manuscript to a SpringerOpen<sup>®</sup> journal and benefit from:**

- Convenient online submission
- Rigorous peer review
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

---

Submit your next manuscript at ► [springeropen.com](https://www.springeropen.com)

# Biotechnologically potential genes in a polysaccharide-degrading epibiont of the Indonesian brown algae *Hydroclathrus* sp.

## ORIGINALITY REPORT

15%

SIMILARITY INDEX

11%

INTERNET SOURCES

13%

PUBLICATIONS

%

STUDENT PAPERS

## PRIMARY SOURCES

- |   |   |     |
|---|---|-----|
| 1 | Amr M. Mowafy, Sherouk Khalifa, Ashraf Elsayed. "Brevibacillus DesertYSK and Rhizobium MAP7 stimulate the growth and pigmentation of <i>Lactuca sativa</i> L.", <i>Journal of Genetic Engineering and Biotechnology</i> , 2023<br>Publication   | 2%  |
| 2 | <a href="http://www.esp.org">www.esp.org</a><br>Internet Source   | 1%  |
| 3 | <a href="http://www.mdpi.com">www.mdpi.com</a><br>Internet Source   | 1%  |
| 4 | Shintaro Kusaka, Kazuki Yamamoto, Motoko Shinohara, Yusuke Minato, Satoshi Ichikawa. "Design, synthesis and conformation-activity relationship analysis of LNA/BNA-type 5'-O-aminoribosyluridine as MraY inhibitors", <i>Bioorganic &amp; Medicinal Chemistry</i> , 2022<br>Publication | <1% |
| 5 | Geneviève Girard, Joost Willemse, Hua Zhu, Dennis Claessen, Kanungnid Bukarasam,  | <1% |

Michael Goodfellow, Gilles P. van Wezel.  
"Analysis of novel kitasatosporae reveals  
significant evolutionary changes in conserved  
developmental genes between Kitasatospora  
and Streptomyces", Antonie van  
Leeuwenhoek, 2014

Publication

---

6	<a href="http://asp-eurasipjournals.springeropen.com">asp-eurasipjournals.springeropen.com</a> Internet Source	<1 %
7	<a href="http://www.biorxiv.org">www.biorxiv.org</a> Internet Source	<1 %
8	<a href="http://www.frontiersin.org">www.frontiersin.org</a> Internet Source	<1 %
9	<a href="http://link.springer.com">link.springer.com</a> Internet Source	<1 %
10	<a href="http://f1000research.com">f1000research.com</a> Internet Source	<1 %
11	Jeff T. Hafting, James S. Craigie, Dagmar B. Stengel, Rafael R. Loureiro et al. "Prospects and challenges for industrial production of seaweed bioactives", Journal of Phycology, 2015 Publication	<1 %
12	Vivek Manyapu, Ayush Lepcha, Sanjeev Kumar Sharma, Rakshak Kumar. "Role of psychrotrophic bacteria and cold-active	<1 %



13

Bing Sun, Yuwei Huang, L. Filipe C. Castro, Su Yang et al. "The chromosome-level genome and key genes associated with mud-dwelling behavior and adaptations of hypoxia and noxious environments in loach (*Misgurnus anguillicaudatus*)", BMC Biology, 2023

Publication

<1 %

14

Lateef B. Salam, Oluwafemi S. Obayori, Mathew O. Ilori, Olukayode O. Amund. "Effects of cadmium perturbation on the microbial community structure and heavy metal resistome of a tropical agricultural soil", Bioresources and Bioprocessing, 2020

Publication

<1 %

15

Lisa L. Drake, Dmitri Y. Boudko, Osvaldo Marinotti, Victoria K. Carpenter, Angus L. Dawe, Immo A. Hansen. "The Aquaporin Gene Family of the Yellow Fever Mosquito, *Aedes aegypti*", PLoS ONE, 2010

Publication

<1 %

16

Nasruddin, Indri Kartika Putri, Sodiq Kamal, Heni Setyowati Esti Rahayu et al. "Evaluation the effectiveness of combinative treatment of cold plasma jet, Indonesian honey, and micro-

<1 %

well dressing to accelerate wound healing",  
Clinical Plasma Medicine, 2017

Publication

17

[hdl.handle.net](https://hdl.handle.net)

Internet Source

<1 %

18

[www.researchgate.net](https://www.researchgate.net)

Internet Source

<1 %

19

Dini Hu, Simon Ming-Yuen Lee, Kai Li, Kai Meng Mok. "Exploration of Secondary Metabolite Production Potential in Actinobacteria Isolated From Kandelia candel Mangrove Plant", Frontiers in Marine Science, 2022

Publication

<1 %

20

Wei Zhou, Songyu Zeng, Jinfeng Yu, Jun Xiang, Fumei Zhang, Mohd S. Takriff, Gongtao Ding, Zhongren Ma, Xueyan Zhou. " Complete genome sequence of NWMCC0046, a candidate for the laundry industry ", Journal of Basic Microbiology, 2022

Publication

<1 %

21

[journals.plos.org](https://journals.plos.org)

Internet Source

<1 %

22

Amany A. El-Shahir, Noha A. El-Tayeh, Omar M. Ali, Arafat Abdel Hamed Abdel Latef, Naglaa Loutfy. "The Effect of Endophytic Talaromyces pinophilus on Growth,

<1 %

# Absorption and Accumulation of Heavy Metals of *Triticum aestivum* Grown on Sandy Soil Amended by Sewage Sludge", Plants, 2021

Publication

23

[aops.springeropen.com](https://aops.springeropen.com)

Internet Source

<1 %

24

[patents.google.com](https://patents.google.com)

Internet Source

<1 %

25

[www.seaweedafrica.org](http://www.seaweedafrica.org)

Internet Source

<1 %

26

[www.researchsquare.com](https://www.researchsquare.com)

Internet Source

<1 %

27

Rachel Rath, Vincent Peta, Heike Bücking. "Duganella callida sp. nov., a novel addition to the Duganella genus, isolated from the soil of a cultivated maize field", International Journal of Systematic and Evolutionary Microbiology, 2021

Publication

<1 %

28

[jjbs.hu.edu.jo](http://jjbs.hu.edu.jo)

Internet Source

<1 %

29

Anran Sun, Wei Song, Weihua Qiao, Xiulai Chen, Jia Liu, Qiuling Luo, Liming Liu. "Efficient agmatine production using an arginine decarboxylase with substrate-specific

<1 %



# activity", Journal of Chemical Technology & Biotechnology, 2017

Publication

30

Jamie K. Lemon, Pavel P. Khil, Karen M. Frank, John P. Dekker. "Rapid Nanopore Sequencing of Plasmids and Resistance Gene Detection in Clinical Isolates", Journal of Clinical Microbiology, 2017

Publication

<1 %

31

refubium.fu-berlin.de

Internet Source

<1 %

32

doaj.org

Internet Source

<1 %

33

download.atlantis-press.com

Internet Source

<1 %

34

edepot.wur.nl

Internet Source

<1 %

35

www.pas-uplbca.edu.ph

Internet Source

<1 %

36

Chopra, L., G. Singh, V. Choudhary, and D. K. Sahoo. "Sonorensin: an Antimicrobial Peptide, Belonging to the Heterocycloanthracin Subfamily of Bacteriocins, from a New Marine Isolate, Bacillus sonorensis MT93", Applied and Environmental Microbiology, 2014.

Publication

<1 %

37 Erin Oi-Yan Wong, Emma Joan Elizabeth Brownlie, Katharine Michelle Ng, Sornnujah Kathirgamanathan et al. "The CIAMIB: a Large and Metabolically Diverse Collection of Inflammation-Associated Bacteria from the Murine Gut", mBio, 2022

Publication

<1 %

38 Pooneh Angoorani, Hanieh-Sadat Ejtahed, Seyed Davar Siadat, Farshad Sharifi, Bagher Larijani. "Is There Any Link between Cognitive Impairment and Gut Microbiota? A Systematic Review", Gerontology, 2022

Publication

<1 %

39 [hal.univ-lorraine.fr](https://hal.univ-lorraine.fr)

Internet Source

<1 %

40 [icfp2022.dryfta.com](https://icfp2022.dryfta.com)

Internet Source

<1 %

41 [www.medrxiv.org](https://www.medrxiv.org)

Internet Source

<1 %

42 Mega Novita, Irna Farikhah, Dian Marlina, Chong-Geng Ma, Benjamin Walker, Kazuyoshi Ogasawara. "First-Principles calculations of the interconfigurational transition energies of 4f - 4f-15d of Ln<sup>3+</sup> ions in LiYF<sub>4</sub> and CaF<sub>2</sub>", Optical Materials, 2021

Publication

<1 %

43	Yujin Choi, Haelim Son, Kyoung Lee. "Complete Genome Sequence of <i>Micrococcus yunnanensis</i> TT9, Isolated from a Healthy Volunteer", Microbiology Resource Announcements, 2022 Publication	<1 %
44	bly.covenantuniversity.edu.ng Internet Source	<1 %
45	doctiktak.com Internet Source	<1 %
46	dokumen.pub Internet Source	<1 %
47	git-r3lab.uni.lu Internet Source	<1 %
48	github.com Internet Source	<1 %
49	nottingham-repository.worktribe.com Internet Source	<1 %
50	peerj.com Internet Source	<1 %
51	A. Lomsadze. "Gene identification in novel eukaryotic genomes by self-training algorithm", Nucleic Acids Research, 11/27/2005 Publication	<1 %

52

Fadeev, Eduard, Fabio De Pascale, Alessandro Vezzi, Sarel Hübner, Dikla Aharonovich, and Daniel Sher. "Why Close a Bacterial Genome? The Plasmid of *Alteromonas Macleodii* HOT1A3 is a Vector for Inter-Specific Transfer of a Flexible Genomic Island", *Frontiers in Microbiology*, 2016.

Publication

<1 %

53

Jaka Widada, Ema Damayanti, Mustofa. "Complete Genome Sequence of the Marine-Derived Bacterium sp. Strain GMY02 ", *Microbiology Resource Announcements*, 2021

Publication

<1 %

54

Kenney, Grace E, and Amy C Rosenzweig. "Genome mining for methanobactins", *BMC Biology*, 2013.

Publication

<1 %

55

Nur Hidayati, Hayatun Fuad, Hendra Munandar, Dewi Seswita Zilda et al. "Proteolytic and Clot Lysis Activity Screening of Crude Proteases Extracted from Tissues and Bacterial Isolates of *Holothuria Scabra*", *IOP Conference Series: Earth and Environmental Science*, 2021

Publication

<1 %

56

Sara C. Gomes, Mirela R. Ferreira, Andreia F. Tavares, Inês N. Silva, Jörg D. Becker, Leonilde M. Moreira. "A Histone-Like Nucleoid

<1 %



# Structuring Protein Regulates Several Virulence Traits in Burkholderia multivorans", Applied and Environmental Microbiology, 2021

Publication

57

[academic.oup.com](https://academic.oup.com)

Internet Source

<1 %

58

[bmcgenomics.biomedcentral.com](https://bmcgenomics.biomedcentral.com)

Internet Source

<1 %

59

[dr.ntu.edu.sg](https://dr.ntu.edu.sg)

Internet Source

<1 %

60

[dspace.cuni.cz](https://dspace.cuni.cz)

Internet Source

<1 %

61

[dspace.vutbr.cz](https://dspace.vutbr.cz)

Internet Source

<1 %

62

[ediss.uni-goettingen.de](https://ediss.uni-goettingen.de)

Internet Source

<1 %

63

[escholarship.org](https://escholarship.org)

Internet Source

<1 %

64

[www.biomedcentral.com](https://www.biomedcentral.com)

Internet Source

<1 %

65

[www.drperlmutter.com](https://www.drperlmutter.com)

Internet Source

<1 %

66

[www.genome.jp](https://www.genome.jp)

Internet Source

<1 %

67

[www.microbiologyresearch.org](http://www.microbiologyresearch.org)

Internet Source

&lt;1 %

68

Hani Pira, Chandra Risdian, Mathias Müsken, Peter J. Schupp, Joachim Wink.

"Winogradskyella luteolasp.nov., Erythrobacter ani sp. nov., and Erythrobacter crassostrea sp.nov., isolated from the hemolymph of the Pacific Oyster Crassostrea gigas", Archives of Microbiology, 2022

Publication

&lt;1 %

69

Perumal Karthick, Raju Mohanraju.

"Antimicrobial Potential of Epiphytic Bacteria Associated With Seaweeds of Little Andaman, India", Frontiers in Microbiology, 2018

Publication

&lt;1 %

70

Priscila Sutto-Ortiz, María de los Angeles

Camacho-Ruiz, Manuel R. Kirchmayr, Rosa

María Camacho-Ruiz et al. "Screening of phospholipase A activity and its production by new actinomycete strains cultivated by solid-state fermentation", PeerJ, 2017

Publication

&lt;1 %

71

The Marine Microbiome, 2016.

Publication

&lt;1 %

72

Xiaoting Yan, Huazhe Si, Yuhang Zhu, Songze

Li, Yu Han, Hanlu Liu, Rui Du, Phillip B. Pope,

Qiang Qiu, Zhipeng Li. "Integrated multi-omics

&lt;1 %

of the gastrointestinal microbiome and  
ruminant host reveals metabolic adaptation  
underlying early life development",  
Microbiome, 2022

Publication

73

[biblio.ugent.be](https://biblio.ugent.be)

Internet Source

<1 %

74

[www.nature.com](https://www.nature.com)

Internet Source

<1 %

Exclude quotes      On

Exclude matches      Off

Exclude bibliography      On