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by Endang Ariyani Setyowati

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Prevalence and diversity of ectoparasites in scavenging chickens (Gallus domesticus) and their association to body weight

EDY RIWIDIHARSO[•], DARSONO, ENDANG ARIYANI SETYOWATI, HERY PRATIKNYO, EMING SUDIANA, SLAMET SANTOSO, EDY YANI, IMAM WIDHIONO

Faculty of Biology, Universitas Jenderal Soedirman. Jl. Dr. Soeparno No. 63, Karangwangkal, Purwokerto Utara, Banyumas 53122, Central Java, Indonesia. Tel./fax.: +62-281-638794, *email: edyriwi@yahoo.co.id

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Abstract. *Rividiharso E, Darsono, Setyowati EA, Pratiknyo H, Sudiana E, Santoso S, Yani E, Widhiono I. 2020. Prevalence and diversity of ectoparasites in scavenging chickens* (Gallus domesticus) *and their association to body weight. Biodiversitas 21: 3163-3169.* Domestic chickens (*Gallus domesticus*) which are traditionally fed by scavenging on farms predispose them to ectoparasites infestation. In this study, a preliminary survey was conducted on the common ectoparasites on *G. domesticus* in the rural areas of Banyumas District, Central Java, Indonesia. The purpose of this study was to determine the diversity of ectoparasites, their prevalence, and their relationship to the chickens' body weight. This research was conducted by the survey method from December 2019 to April 2020 in five villages around the city of Purwokerto, Banyumas District viz., Kedungwuluh, Kedungwringin, Kutasari, Karangsalam, and Karanggintung. Analysis of ectoparasites diversity was conducted by using the Shannon-Wiener index **1***H*') and Evenness index (E). Analysis of variance was used to calculate the difference in prevalence among sample locations. Regression correlation analysis was used to determine the relationship between prevalence and chicken body weight. The results showed that there were six ectoparasite species i.e., *Dermanysus gallinae, Megninia ginglymura, Menacanthus cornutus, Menopon gallinae, Lipeurus caponis*, and *Haemaphysalis* sp. parasitizing *G. domesticus* in five villages. The number of ectoparasite individuals among locations was significantly different (F 559 < 32.45; $p_{.005}$). The highest number of ectoparasite prevalence was associated with chicken weight loss (r = 0.98). The prevalence of ectoparasite of domestic chickens farm in the Banyumas Regency is classified as low but has the potential to influence body weight.

Keywords: Ayam kampong, body weight, ectoparasite, prevalence, traditional farm

INTRODUCTION

Rural chicken farms are generally managed very simply. Consequently, they are very vulnerable to both endo and ectoparasites attack (Suhaila et al. 2015). Domestic chicken production in rural areas faces many obstacles such as insufficient feed, poor management, and poor biosecurity (Nik-Hasan et al. 2015). The habit of releasing native chickens throughout the day to get food on their own makes them very susceptible to ectoparasite attacks. This is because these chickens accidentally contract infective eggs, larvae, and intermediate hosts from ectoparasites. According to Rahman and Haziqoh (2015), ectoparasite attack is one of the main problems in the farming of indigenous chickens in the countryside.

The common ectoparasites in native chickens are fleas and mites. The fleas are generally from the order Phthiraptera, sub order Mallophaga (Suaila et al. 2015). Ilyes et al. (2013) found that ectoparasite attacks can cause health problems in chickens by damaging tissue and causing blood loss and toxicosis dermatitis. In addition to these symptoms, Aleya and Sabrina (2011) also found allergies. Several types of ectoparasites can also cause or act as a vector of various pathogens, which, in severe attacks, will weaken the hens and cause death (Swai et al. 2010; Nik-Hasan et al. 2015). According to Gunya et al. (2020), the most common external parasite in the scavenging chicken were mites, followed by lice. Lice and mites can cause chick mortality, attributed to starvation and immune suppression from the infestation (Nnadi and George 2010).

Ectoparasites are very common in native chickens in traditional chicken farms in various countries such as USA (California state) (Murillo and Mullens 2016), India (Sreedevi et al. 2016), Bulgaria (Prelezov and Koinarski 2006), Egypt (El-Aw et al. 2008), and Benin (Love et al. 2017). Nik-Hasan et al. (2015) found ectoparasites and their prevalence in Sandakan, Malaysian Borneo to be: *Dermanyssus gallinae* (78%), *Lipeurus caponis* (80%) and *Menopon gallinae* (100%). Love et al. (2017), in their research on chewing lice in selected poultry in Benin, recorded the highest prevalence of ectoparasite was *Menacanthus stramineus* (56%), followed by *Menopon gallinae* (34%) and *Lipeurus caponis* (10%).

Ectoparasites in chickens, aside from the insect class, are also found from the mite class Arachnida and subclass Acari and ticks (Ixodidae). Two species of mites commonly found in chickens around the world are *D. gallinae* and *Megninia ginglymura*. *D. gallinae* is a blood-sucking mite that causes decreased egg-laying ability and anemia in chickens and can be a vector of disease in humans and other animals (Sparagano et al. 2009). The presence of these mites will affect the hens' ability to lay eggs and can cause disease from viruses and bacteria (Marangi et al.



2009). *M. ginglymura* is alm⁽³⁾ found in chickens and associates with feathers. Its saliva can cause lesions, allergic reactions with pruritus, serous scabs, stress, and itching, leading to the development of petechial hemorrhage, crust formation, secondary bacterial infections, and pyoderma (Faleireo et al. 2015). Huge mortality, slow growth rate, and malnutrition au⁶ common problems in scavenging chicken (FAO 2014). Parasitic infection is one of the non-infectious health hazards in backyard chicken, whose severity and diversity are primarily influenced by geography and climatic interaction. The parasitic problem has been identified as one of the causes of low productivity in free-range chicken (Olivirea et al. 2017).

In Banyumas District, chicken farms are maintained traditionally and only at the household scale, with a relatively small chicken population reared in a small scale system. However, this activity is very important for the rural economy and is used as a medium-term saving. Although ectoparasites in native chickens are an important factor in poultry, research on the diversity and prevalence of ectoparasites and their effects on native chicken body weight is still very rare in Indonesia. Therefore, the purpose of this study was to determine the diversity and prevalence of ectoparasites, their attack rates, and their relationship to the bodyweight of the native chicken. The long-term goal of this research is to establish a basis for controlling ectoparasite attacks in left-over native chickens.

MATERIALS AND METHODS

Study area

The study was conducted on 100 scavenging chickens randomly obtained from five villages around Purwokerto, Banyumas District, Central Java, Indonesia, viz., Kedungwuluh (sub-district West Purwokerto), Kedungwringin (sub-district Patikraja), Kutasari (sub-district Baturaden), Karangsalam (sub-district Kedungbanteng) and Karanggintung (sub-district Sumbang) (Figure 1). The chickens had been left scavenging around the house.

Procedures

Chicken sampling procedures

Chicken samples were randomly obtained directly from the owner in the selected village. They were completely left day and night. The average age of the chicken is eight months based on information from the owner. From each village, twenty chickens were taken, consisting of ten hens and ten cocks. Each chicken was weighed. At the time of chicken sampling, environmental conditions of each location were observed, which included the cleanliness of the chicken's living area, chicken population per family, maintenance methods, feeding, and the size of the scavenging area (Table 1).



Figure 1. Sampling location at Banyumas District, Central Java, Indonesia

Table 1. Environmental parameters of five domestic chicken sampling sites

Parameter	Kedungwuluh	Kedungwringin	Kutasari	Karangaslam	Karanggintung
Temperature (°C)	30-32	30-32	30-32	30-32	30-32
Humidity (%)	90-95	90-95	90-95	90-95	90-95
Cleanliness	Clean	Clean	Clean	Clean	Dirty
Populations (individual)/ family	10-15	5-10	10-15	10-15	15-20
Living area (m)	<50	<50	<50	<50	>50
Maintenance methods	free	cage	cage	cage	cage

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Ectoparasite sampling procedure

Isolation of ectoparasites in the chickens was carried out by examining the following body parts; head-neck, back, wings, chest-legs, and tail. Further, the collected ectoparasites were put into a ample bottle containing 70% ethanol for observation in the Entomology and Parasitology Laboratory, Faculty of Biology, Jenderal Soedirman University. Ectoparasites were separated based on groups namely insects, mites, and ticks under a binocular microscope Olympus CH 20. For identification purposes, the specimens were prepared on an object glass and placed in Hoyer's solution for clearing in one week, and then they were identified according to Murillo and Mullens (2016).

Data analysis

Analysis of ectoparasites diversity data was done using the Shannon-Wiener index (H') and Evenness index (E). All data were analyzed using BD Pro software (McAleece et al. 1997). Shannon-Wiener diversity and Evenness indexes were calculated with the formula (Magurran 2000):

$$\mathbf{H}' = \sum_{i=1}^{s} p i \ln p i$$

Where; H': Shannon-Wiener diversity index, pi: total individual(s) found within species i.

$$E = \frac{H'}{\ln S}$$

Where; E: Evenness index, H': Shannon-Wiener diversity index, S: total species of ectoparasite.

Simpson's dominance index was used to measure dominance following this formula:

$$D = \sum (pi)^2 = \sum (\frac{ni}{N})^2$$

Where; D: Simpson's dominance index, ni: total individuals within a species, N: total individuals within all species.

Table 2. Abundance of ectoparasite in kampong chicken

Analysis of variance (ANOVA) was used to calculate the difference in prevalence between sample locations, and between males and females. Regression correlation analysis was used to determine the relationship between prevalence and chicken body weight. All analysis was run with the help of SPSS 21 software.

RESULTS AND DISCUSSION

Ectoparasite diversion

A total of 1047 ectoparasites consisting of three species of lice, two species of mite, and one species of ticks were found in this study. The species of lice encountered were *Menopon gallinae* (Phthiraptera, Menoponidae), *Menacanthus stramineus* (Phthiraptera, Menoponidae), and *Lipeurus caponis* (Phthiraptera, Philopteridae). The mites consisted of *Megninia ginglymura* (Astigmata, Analgidae) and *Dermanyssus gallinae* (Astigmata, Dermanyssidae) and the ticks were *Haemaphysalis* sp. (Ixodida: Ixodidae) (Table 2, Figure 1).

The number of species found is relatively small compared with other studies, probably due to the number of sampling size. High ectoparasite attacks are caused by the level of yard hygiene, air humidity, ambient temperature, and rainfall (Rahman and Haziqoh 2015). This result is in a conduct with Nik-Hasan et al. (2015) in their study of parasitic burden and its relation with the bodyweight of free-range chicken in oil palm dominated Sandakan District of Malaysian Borneo where also only four species of ectoparasite were found.

Rahman and Haziqoh (2015) found ten species of ectoparasite in their study on native chicken in urban areas. Furthermore, a high diversity of ectoparasites was found in native chickens in the African region with recorded species including *Menopon gatoae*, *Menacanthus stramineus*, *Menacanthus cornutus*, *Menacanthus pallidulus*, *Lipeurus caponis*, *Goniocotes gallinae*, *Goniodes dissimilis*, *Goniodes gigas* (El-Aw et al. 2013). Zeryehun and Yohannes (2015) recorded five species of mites in the chickens reared under traditional backyard production system in southern Ethiopia.

	Location										
Species	I		II		III		IV		v		Σ
-	Cock	Hen	Cock	Hen	Cock	Hen	Cock	Hen	Cock	Hen	
Lice											
Menopon gallinae	31	57	28	20	24	21	34	46	24	25	310
Lipeurus caponis	16	27	18	13	14	16	21	25	11	23	184
Menacanthus cornutus	24	37	34	22	19	28	32	37	21	36	290
Tick											
Haemaphysallis sp.	16	2	17	9	7	3	14	16	18	9	111
Mite											
Dermanyssus gallinae	10	15	12	7	5	1	10	13	7	11	91
Megninia ginglymura	5	5	11	11	5	6	8	16	9	12	88
Total	102	143	120	82	74	75	119	153	90	116	1074

Note: I: Kedungwuluh, II: Kedungwringin, III: Kutasari, IV: Karangsalam, V: Karanggintung

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Among all the sampling sites in this study, the most abundant ectoparasite was *Men. gallinae* (310 individuals, 29.60%), followed by *Me. cornuthus* (290 individuals, 27%), while the least was *M. ginglymura* with 88 individuals (8%) (Table 1). This result showed that there were no significant differences among the sampling sites (F_{4.24}; p > 0.05). The dominance of shaft louse (*Men. gallinae*) is very common because this ectoparasite is widespread. *M. gallinae* attacks the base of the feat **9** rs and cause loss of chicken feathers. The adults of *Men. gallinae* measure about 2 mm in length, and females deposit eggs **9** lgly at the base of the shaft on thigh and breast feathers. Poultry lice typically transfer to new chicken by direct host contact. The abundance number of *Men. gallinae* tabein it disperses easily among chicken.

Shannon diversity index (H') among the sampling sites was almost the same, ranging from H': 1.551 (site III) to H': 1.712 (site II). This means that all chicken sampled were invested by the same number species of ectoparasite (six ectoparasites) (Table 2). The density of the cage greatly influences the number of ectoparasites in chickens due to the high level of contact between individual chickens, as most transmissions of ectoparasites between individuals are by direct contact (Suahila et al. 2015). Besides insects, this study also found two species of mites, i.e., D. gallinae (91 individuals, 8.4%) and M. ginglymura (88 individuals, 8.1%). The presence of these mites is common, because Bey are widespread in chickens throughout the world. D. gallinae is hematophagous and 3 esponsible for egg downgrading and anemia in birds (Sparagano et al. 2009). This ectoparasite is the most important affecting laying hens worldwide. The presence of this mite has been associated with a viral zoonosis and several bacterial infections (Marangi et al. 2009). One species of ticks

(Acari, Ixodidae) found in this study was *Haemaphysallis* sp., which is a bloodsucking tick and is widely distributed in Southeast Asia. This is a very important parasite in chicken farms because it can cause infection and become a vector of other diseases (Ernieenor et al. 2017). *Haemaphysallis* sp. was also found in the study of Rahman and Haziqoh (2015) in Malaysia with a low population.

The results showed that ectoparasite was found in the four parts observed, namely head-neck, back, wings, chest-legs, and tail. However, ectoparasite species have preferences for different chicken body parts. *L. caponis* was found in all of the observed chicken body parts, *M. cornutus* was found in four parts of the chicken body except wings, *Men. gallinae* was found only in the head-neck, back, and chest-legs. *M. ginglymura* was found only on the wings and tail (Table 4).

Among the body parts of chickens, it was observed that the back and chest-legs were the most common abodes for ectoparasites with five species present, followed by headneck section with four species, and the wing had with three species. The lice identified were *Men. gallinae*, *Me. cornutus*, *L. caponis*, the mites were *Goniocotes gallinae*, *M. ginglymura*, and the scrab was *Haemaphysallis* sp.

Prevalence

At the five sampling locations, the highest prevalence was of *Men. gallinae* (45%), followed by *L. caponis* (40%), *Me. cornutus* (38%), and *M. ginglymura* mites (25%). This is caused by the nature of *Men. gallinae* (chicken body louse) as a voracious feeder so it can spread very fast. In addition, the development of *Men. gallinae* population is greatly influenced by changes in environmental factors, especially the air temperature (Ilyes et al. 2013).

Table 3. Diversity parameters of five different sampling site

Diversity Parameters	Kedungwuluh	Kedungwringin	Kutasari	Karangaslam	Karanggintung
Taxa_S	6	6	6	6	6
Individuals	245	202	149	272	206
Dominance_D	0.2393	0.1941	0.2428	0.2066	0.1956
Shannon_H	1.575	1.712	1.551	1.675	1.707
Simpson_1-D	0.7607	0.8059	0.7572	0.7934	0.8044
Evenness_e^H/S	0.8048	0.9238	0.7863	0.8895	0.9183
Equitability_J	0.8788	0.9558	0.8658	0.9347	0.9524

Table 4. Distribution of ectoparasite in the body regions of the domestic chicken

	Ectoparasite											
Body Regions	M.gallinae		M. cornutus		L. caponis		D. gallinae		M.ginglymura		Haemaphysallis sp.	
-	Σ	%	Σ	%	Σ	Σ	Σ	%	Σ	%	Σ	%
Head-neck	142	37.9	130	34.7	72	19.2	31	8.3	-	-	-	
Back	137	56.4	67	27.6	16	6.6	10	4.1	-	-	2	0.8
Wing	-	-	-	-	26	39.4	-	-	11	4.5	40	60.6
Breast-foot	31	10.5	65	22.03	54	18.3	50	16.9	-	-	36	12.2
Tail	-	-	28	29.5	16	16.8	-	-	59	20.0	33	34.7

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Figure 2. Ectoparasite species found in kampong chickens at five sampling sites. A. Menopon gallinae; B. Menacanthus cornutus; C. Lipeurus caponis; D. Haemaphysallis sp.; E. Dermanyssus gallinae; F. Megninia ginglymura

The results of this study are in line with the majority of studies that found Men. gallinae with the highest prevalence. Suhaila et al. (2015) in North Malaysia found four species of ectoparasites, consisting of three lice and a tick, the highest prevalence being of Men. gallinae (93.8%), followed by Me. pallidulus (81.3%), Haemaphysalis sp. (10,5%), and L. caponis (18.8%). Furthermore, Men. gallinae was also the most common ectoparasite with 76.7% occurrence, followed by L. caponis (63.3%) in Penang Malaysia (Rahman and Haziqoh 2015). The same results were found by Ilyes et al. (2013) in their research in Algeria, with the highest prevalence was Men. gallinae (97%), followed by L. caponis (41%), and Me.cornutus only 20%. The dominance of Men. gallinae was also recorded by Deephali et al. (2005) in India, Prelezov and Koinarski (2006) in Bulgaria and Marin-Gomez and Benavides-Montano (2007) in Columbia. In Benin, three

species of lice (Men. gallinae, Me. stramineus, and L. caponis) were detected and identified on the local poultry. The most abundant species in the study area is Men. gallinae, with a prevalence percentage of about 69.84% (Love et al. 2017), while Mansur et al. (2019) in Egypt found Men. gallinae to be the most common ectoparasite with 76.7% occurrence, followed by L. caponis (63.3%) and Me. pallidulus (41.7%). The difference in this discovery may be due to the difference in environmental factors, especially the daily temperature and humidity (Kaboudi et al. 2019). Overall, the results of this study indicate that the prevalence of ectoparasite in native chicken in Banyumas District is low, compared to the results of research elsewhere. The highest prevalence of Me. cornutus only found in the research of Zeryehun and Yohannes (2013) in Ethiopia and Kumar and Kumar (2014) in India.

Table 5. Prevalence of ectoparasite in domestic chicken in five different sampling sites

Species		Prevalence (%)				
species	Kedungwuluh Kedungwringin Kutasari Karangaslam			Karanggintung	Means ± SD	
M. gallinae	40	45	40	45	55	45 ± 5.5
L .caponis	45	40	35	35	45	40 ± 4.5
M. cornutus	35	40	35	35	45	38 ± 4.0
Haemaphysalis sp.	15	20	25	35	35	26 ± 8.0
D. gallinae	25	10	30	25	35	25 ± 8.4
M. synglineura	15	15	30	30	35	25 ± 8.4

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Body weight

In this study, there was a real relationship (r: 0.98) between ectoparasite attacks and weight loss; the higher the ectoparasites attack, the lower the chicken's body weight. This is probably due to irritation for the chickens, thereby reducing sleep and appetite. Ectoparasites of poultry live on the skin or penetrate into the skin or even in the air sacs, while some live under the feathers. The ectoparasites cause irritation, interfere with feed consumption and are thus associated with emaciation, anemia, and erentual loss of production (Mishra et al. 2017). Although poultry lice are not known to transmit any avian pathogens, the presence of lice frequently accompanies poor health attributable to other causes, and is especially harmful to young birds, where largenumbers of lice may cause sleep disruption. Amblycera lice may cause irritation to the skin, restlessness, overall weakening, and cessation of feeding. These cause body weight loss, inferior laying capacity, and skin lesions that may become the site for secondary infections (Mullen and Durden 2002). Katoch et al. (2012) reported a 24% loss in the bodyweight of scavenging chicken in India due to parasitism.

Based on the results of the study, the diversity and prevalence of ectoparasite in native chickens in Banyumas District were classified as low. There we only found six species with the highest prevalence of *Men. gallinae* at 45%. Ectoparasite infestations in native chickens can cause chicken weight loss. Therefore, even though the prevalence of ectoparasite is low, this is a key area of consideration by the village chicken farmers to prevent ectoparasite attacks on their chicken, hence boosting productivity.

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