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1st International Conference on Material Science and Engineering for Sustainable Rural Development



Central Java, Indonesia

14–15 November 2018

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Preface: 1st International Conference on Material Science and Engineering for Sustainable Rural Development (ICMSE-SURE)

The *1st International Conference on Material Science and Engineering for Sustainable Rural Development (ICMSE-SURE)* was held in Java Heritage Hotel, Purwokerto, Indonesia. This two-day conference was held on 14-15 November 2018 with a theme of *Science and Engineering for Rural Innovation*. This conference in conjunction with **ICLAS-SURE** for life and applied science and **ICAH-SURE** for Arts and Humanities together in the topic for Rural Innovation.

The purposes of the conference are:

- to provide a forum for scientific discussion, professional networking, research collaboration, education, and dissemination of scientific research, innovation and industrial products.
- to increase the quality of research and development in the multidisciplinary approach for sustainable rural development.
- to encourage the local and regional young scientists to attend and present their works at the international level.

The success of the Conference would not have been attained without strong supports from contributing scientists and as well as Research and Society Service of Universitas Jenderal Soedirman Committee. I would like to thank all of them for helping to make a very successful conference.

We hope that you will enjoy a pleasant and valuable conference at Purwokerto, organized by the Research and Society Service Institute, Jenderal Soedirman University.

Thank you

Amin Fatoni, Ph.D.
ICMSE-SURE Chairman
Universitas Jenderal Soedirman
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The Protein Content and Protease Activity of Local Green Fly, *Chloroprocta* sp., Maggot Crude Extracts

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Abstract. Maggot extracts of local green flies, *Chloroprocta* sp., is a potential anti-biofilm candidate who can inhibit biofilm formation, break down the biofilm matrix, and reduce the viability of embedded *Staphylococcus epidermidis*. One component of *S. epidermidis* biomatrix is protein thus protease activity is presumably needed for biofilm detachment. Hence, this study aimed to analyze the protease activity and the protein content of maggot crude extracts from green flies, *Chloroprocta* sp. A quantitative enzymatic assay of protease activity with casein as a substrate was performed with some modifications. The protein content was measured with the Lowry method at a wavelength of 750 nm with tyrosine as a standard. The results revealed that the crude extracts from 0.425 grams maggots /mL PBS has protease activity of 4.875 U/mg and protein content of 29.252 ppm. These results indicated that the crude extract of maggot from local green flies, *Chloroprocta* sp. has protease as the active compound which could potentially be an anti-biofilm.

INTRODUCTION

Biofilm producing bacteria have a significant role as a cause of Healthcare-associated infections (HAIs). The mortality rate caused by HAIs in adult patients from 2003 to 2008 in Asia, Latin America, and Africa was 23.6%, 18.5%, and 29.3%, respectively [1]. The selection of antibiotics that are sensitive and have high penetration ability is very limited due to the increased incidence of antibiotic resistance to biofilm producing bacteria. One ingredient that has been widely investigated for its effect on biofilms and virulence factors is the maggot (larva) of green fly from the family *Calliphoridae* [2]. Maggot that has been studied in Europe comes from green flies *Lucilia sericata* [3], while in Singapore *Lucilia cuprina* is used [4]. Maggot requires a relatively short time to develop which are four days at 20°C and three days at 27°C. That condition causes obstacles in purchasing maggot from overseas countries' producers. Thus, the obtainability of indigenous green flies from Indonesia is important for organism alternative.

Recently, Anjarwati et al. [5] have reported that maggot from *Chloroprocta* sp. has anti-biofilm activity towards biofilm produced by *Staphylococcus epidermidis*. *Chloroprocta* sp. is a green fly species found predominantly in Semarang, the capital of Central Java Province, Indonesia. Maggot filtrate from *Chloroprocta* sp. was able to reduce the extracellular biofilm matrix and reduce the viability of biofilms produced by *S. epidermidis* [5,6]. However, the mechanism of biofilm degradation by *Chloroprocta* sp. is still unclear.

Generally, the composition of biofilm matrix is complex and varied among microorganism. The biomatrix composition is also varied even within the same species under different conditions. The essential component of biofilm matrix is exopolysaccharides such as cellulose and the poly-β-1,6-N-acetylglucosamine as the typical

components [7]. Surface proteins and proteinaceous components are also crucial in biofilm formations that provide biofilm structure in which the bacterial cells are initially attached [8]. Various approaches to treat biofilm have been developed [9]. The treatment of biofilm with broad specificity proteases leads to biofilm disintegration [10]. Another approach to biofilm treatment is using anti-biofilm peptide which inhibits developed biofilm and also destroys multiple species in biofilms [11]. Indeed, maggot extract from *L. sericata* was reported having an active component to overcome the causes of resistance in biofilms which were proteolysis enzymes or proteases, and peptides [12]. Hence, this research aims to identify protease activity and protein content of crude extract of *Chloroprocta sp.* The findings will give an insight on how biofilm disruption by indigenous *Chloroprocta sp.*

MATERIALS AND METHODS

Materials

The green flies *Chloroprocta sp.* was reared in the Laboratory of Nutrition in the Faculty of Agriculture and Animal Husbandry, Diponegoro University, Semarang. Phosphate buffered saline was from Oxoid (Thermo Fisher Scientific, USA). Casein, *Folin Ciocalteu* and L-tyrosine were from Sigma-Aldrich (Germany). Trichloroacetic acid and Na₂CO₃ were from Merck (Germany). Ethanol technical grade and deionized water were from local chemical supplier.

Rearing and Collecting Maggots

The maggots was reared following the methods described by Arora et al. [4] and Anjarwati et al. [6]. The green flies were trapped by placing raw fish in containers. Then, the eggs were collected from the containers and washed three times with sterile deionized water and ethanol to sterilize them. After that, the eggs were allowed to hatch into maggots and collected when they entered the late of second phase or the early third phase of the *Chloroprocta sp.* maggot growth. The maggots were washed three times by using ethanol and sterile deionized water in different containers.

Maggot Extraction

Maggot extraction was performed following the methods described by Honda et al. [13] and Anjarwati et al. [6]. The maggots were extracted by soaking the maggots in the sterile phosphate buffered saline (PBS) pH 7.3 for 1 hour at room temperature (25°C) in the dark. After that, the maggots were incubated in PBS for 48 hours at 37°C. The mixture was centrifuged at 25°C, 10,000 rpm for 15 minutes. The supernatant was collected and sterilized using the syringe filters (0.2 µm, Corning NY 14831). Finally, the maggots' extracts were stored in the freezer at -20°C before further examination.

Determination of Protease Activity

Protease activity was measured following protease activity assay by Sigma [14] in which the casein was used as substrate. 1 mL of maggot crude extract was added to 5 mL of 0.65% casein substrate. 1 mL of distilled water was added to 5 mL of 0.65% casein as blank. The mixture was incubated at 37°C for 10 minutes. Termination of the reaction was carried out through the addition of 5 mL of 110 mM Trichloroacetic acid reagent. After termination, 1 mL of maggot crude extract was added to the blank. Next, the mixture was re-incubated at 37°C for 30 minutes. 2 mL of filtrate was separated by centrifugation at 10,000 rpm for 10 minutes. 5 mL of Na₂CO₃ and 1 mL of *Folin Ciocalteu* reagent were added to the filtrate and the mixture was incubated at 37°C for 30 minutes. The absorbance of the mixture was measured at 660 nm.

Tyrosine standard curve was prepared by reacting 2 mL of standard tyrosine and distilled water (as blank) with 5 mL of Na₂CO₃ and 1 mL of *Folin Ciocalteu* reagent. The mixtures were incubated at 37°C for 30 minutes, then the absorbance was measured at 660 nm. One unit of protease activity is designated as the amount of protease needed to release 1 µmol tyrosine from the casein substrate per minute.

Determination of Protein Content

The protein content was measured following the Lowry method [15]. 0.5 mL of maggot crude extract was added 0.7 mL of Lowry solution and incubated for 20 minutes at room temperature in the dark. Then, 0.1 mL of diluted *Folin Ciocalteu* was added followed by incubation for 30 minutes at room temperature in the dark. After incubation, the samples were measured the absorbance at 750 nm.

RESULTS AND DISCUSSION

Recently, several countries have developed research on greenfly maggots as a therapy in the health sectors. Green fly maggot that has been studied in Indonesia was the genus *Chloroprocta* sp. which is found in the area of Central Java. The Green fly species, *Chloroprocta* sp., is a subfamily of *Chrysomyinae* from the family *Calliphoridae*, kingdom *Diptera*. The study of Anjarwati et al. [5] suggested that the maggot extract of *Chloroprocta* sp. has a protease component judged by a qualitative protease test using the gelatin hydrolysis method. Maggot extract with protease components in that study could damage extracellular biofilm matrix and reduce embedded *S. epidermidis* viability. This present study further analyzed the protease activity and the protein content of maggot crude extracts from greenflies, *Chloroprocta* sp.

The protein content of maggot crude extract was measured using Lowry method [15]. The Lowry method is a development of the Biuret method. In this method, two reactions are involved. Initially, the Cu (II)-protein complex will be formed as the biuret method, which in the alkaline atmosphere Cu (II) will be reduced to Cu (I). The Cu⁺ ion will then reduce the *Folin-Ciocalteu reagent*, the phosphomolybdate-phosphotungstate complex, resulting in the blue heteropoly-molybdenum due to Cu catalyst aromatic oxidation (amino acid side chain) reaction, which gives intense blue color and can be detected colorimetrically. The strength of blue depends mainly on the content of tryptophan and tyrosine residues. The advantage of the Lowry method is more sensitive (100 times) than the Biuret method so that it requires fewer protein samples. The detection limit ranges from a concentration of 0.01 mg/mL. However, Lowry's method had more interference due to its sensitivity.

The protease activity of maggot crude extract was determined following Sigma's method [14]. The casein was used as a substrate. The proteases hydrolyze casein and release tyrosine as well as other amino acids. *Folin Ciocalteu* reacts mainly with free tyrosine and produce blue colored compounds, which can be quantified by spectrophotometric as absorbance. The more tyrosine released from casein, the more is absorbance value which indicates the higher protease activity. The absorbance values generated by the samples are compared to a tyrosine standard curve to determine the activity of protease samples which is the equivalent amount of released tyrosine in micromoles from casein per minute.

The results of this research showed that 0.425 grams/mL maggot crude extract in PBS has protease activity of 4.875 U/mg and protein content of 29.252 ppm (Table 1).

TABLE 1. Analysis of protein content and protease activity of maggot crude extract only.

| | Vol. Enzyme (μL) | Total Vol. (mL) | A 750 nm | A Tyrosine (As - Ab)* | Tyrosine (ppm) | Protease Activity (U/mg Enzyme) |
|---------------|---------------------|--------------------|----------|--------------------------|----------------|------------------------------------|
| Crude extract | 0.1 | 238 | 0.888 | 0.839 | 29.252 | 4.875 |

*Absorbance for Blank (Ab) at 750 nm was 0.049.

The proteases are class of enzymes that hydrolyze peptide bonds. They can act as anti-biofilm due to their ability to disrupt surface proteins in biofilm matrix. Commercial protease enzymes have been reported as a good anti-biofilm agent which target the protein produced by *S. epidermidis* and *Staphylococcus aureus* during early biofilm formation [10]. The anti-biofilm activity was proportional to enzyme quantity. The commercial Flavourzyme which is a combination of exoprotease and endoprotease has anti-biofilm activity specific against *S. epidermidis* at a concentration of 3 U/mL and over [10]. Extracellular protease from actinomycetes strains AN090250 and AN091562 have been found to inhibit *S. aureus* biofilm development and degrade pre-existing *S. aureus* biofilm [16]. The extracellular proteases are known to have a broad substrate specificity hence the enzymes can degrade any protein in biofilm matrix that maintains the biofilm integrity.

In this research, maggot extract of *Chloroprocta* sp. affirmatively contains protein and protease activity. These findings may explain why the extract can reduce biofilms produced by *S. epidermidis* [6, 7]. Proteases in maggot

extract can accelerate the natural release of biofilms by endogenous enzymes. In addition, the biofilm structural damage reduces the ability of biofilms to provide nutrients for embedded bacteria [17]. Further research that can be done includes identification of types of protease in *Chloroprocta* sp., protease effects on biofilm regulator genes, increased protease production in maggot, in vivo tests and clinical trials of maggot extract as anti-biofilm on biofilms produced by monospecies and multispecies bacteria.

CONCLUSION

The maggots extract from indigenous green flies *Chloroprocta* sp. contained protein and protease activity. The ability of protease enzyme to disintegrate peptide bond makes the indigenous green flies *Chloroprocta* sp. as anti-biofilm producing organism to reduce surface protein in biofilm matrix from *S. epidermidis*.

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Ari Asnani <asnani@gmail.com>

ICMA-LPPM Review Complete

2 messages

ICMASure Unsoed <icmasure.unsoed@gmail.com>
To: ari.asnani@unsoed.ac.id, asnani@gmail.com

Mon, Jan 14, 2019 at 6:57 AM

We are glad to inform you that we have received the external review results of your manuscript from well-known experts. Please check the reviewed manuscript and note the comments/suggestions/answers to questions of the reviewers.

As the proceedings will have to be published soon, we request you to

incorporate the reviewers' comments/suggestions/answers to questions in the manuscript as soon as possible, not later than 31 January 2019.

We look forward to hearing from you. Thank you very much for your kind attention and cooperation.

Regards,


The Committee of ICMA-SURE LPPM Unsoed 2018

 **review report of abs-113_1782192727.docx**
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Ari Asnani <asnani@gmail.com>
To: Dwi Utami Anjarwati <dzikrarana@gmail.com>

Sun, May 19, 2019 at 10:06 AM

[Quoted text hidden]

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Ari Asnani <asnani@gmail.com>

2nd Revision & Filled AIP Agreement RE: Revised article by Dwi Utami Anjarwati et al.

2 messages

Ari Asnani <ari.asnani@unsoed.ac.id>
To: ICMASure Unsoed <icmeasure.unsoed@gmail.com>
Cc: "asnani@gmail.com" <asnani@gmail.com>

Tue, Feb 12, 2019 at 7:33 AM

Dear Editors,

The second revision of article from [review report of **abs-113_1782192727**], entitle "The Protein Content and Protease Activity of Local Green Fly, *Chloroprocta* sp., Maggot Crude Extracts".

Authors: Dwi Utami Anjarwati, Rizka Hidayati, Dian Kristiantoro, IDSAP Peramiarti, Ari Asnani

Corresponding author Ari Asnani (ari.asnani@unsoed.ac.id)

Hereby, we would like to submit our second revised article which already follow the AIP format.

We also attache the filled AIP agreement.

Thank you very much for your kind attention.

Best Wishes,
Ari Asnani

-----Original message-----

From: ICMASure Unsoed <icmeasure.unsoed@gmail.com>
Sent: Monday 11th February 2019 7:04
To: Ari Asnani <ari.asnani@unsoed.ac.id>
Subject: Re: Revised article by Dwi Utami Anjarwati et al.

Mohon artikel disesuaikan dengan template terlampir dan mengisi copyright transfer.

The Committee of ICMA-SURE LPPM Unsoed 2018

2 attachments



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534K

Ari Asnani <asnani@gmail.com>
To: Dwi Utami Anjarwati <dzikrarana@gmail.com>

Sun, May 19, 2019 at 10:09 AM

[Quoted text hidden]

2 attachments



Revised Dwi Utami Anjarwati et al.doc

5/18/22, 10:04 PM

Gmail - 2nd Revision & Filled AIP Agreement RE: Revised article by Dwi Utami Anjarwati et al.

127K



Agreement Dwi Utami Anjarwati et al.pdf

534K

Dear the editor,

I would like to report my review on the paper entitled " The Protein Content and Protease Activity of Local Green Fly, *Chloroprocta* sp., Maggot Crude Extracts".

This paper studied the protease activity and protein content of the crude extract of *Chloroprocta* sp., with the proportion of anti-biofilm being important. By using the Lowry method, the authors were able to determine the protein content of the crude extract of *Chloroprocta* sp. The protease activity can also be measured using assay-casein as a substrate with some modification. They concluded that the crude extract *Chloroprocta* sp. was able to reduce the extracellular biofilm from the measurement results. The results are of great interest due to the applicability of this research to a biofilm candidate.

Here are some general comments and suggestions for improvement:

First and most importantly, the contribution of this research to a unified history should be linked to convey its scientific significance. The authors seem reluctant to draw conclusions about their interpretation.

In particular, the authors should address the following:

- Explain how to relate the results to biofilm properties (whether they inhibit damage or biofilm formation).
- What is the significant number of protein content (29,252 ppm) with protease activity? Does it belong as an anti-biofilm to a high / low protein content?
- Sections 3 and 4 in the Results and Discussion section only seem to explain how the protein content can be obtained using the Lowry method.
- The conclusion reads: "...thus greenfly *Chloroprocta* sp. might be a potential candidate for anti-biofilm producing organism." Perhaps it would be better if the authors also explain why, just because *Chloroprocta* sp. contained protein with protease activity can then reduce the extracellular biofilm of *S. epidermidis*. How did this process come about?
- Written in the research method " The method for obtaining maggots was carried out in the Laboratory of Nutritional of Agriculture and Animal Husbandry, Faculty of Diponegoro University?"

Overall, this paper contains very good research ideas. Then I recommend that this paper be published in the journal after some minor revisions.

Thank you,
Referee



Ari Asnani <asnani@gmail.com>

Revised article by Dwi Utami Anjarwati et al.

2 messages

Ari Asnani <ari.asnani@unsoed.ac.id>

Mon, Feb 11, 2019 at 9:56 AM

To: ICMASure Unsoed <icmasure.unsoed@gmail.com>, "asnani@gmail.com" <asnani@gmail.com>

Revised article from [review report of **abs-113_1782192727**]

Dear ICMASure Unsoed Organizer

We would like to submit our revised article entitle:

"The Protein Content and Protease Activity of Local Green Fly, *Chloroprocta* sp., Maggot Crude Extracts"

Authors: Dwi Utami Anjarwati, Rizka Hidayati, Dian Kristiantoro, IDSAP Peramiarti, Ari Asnani

with corresponding author Ari Asnani (ari.asnani@unsoed.ac.id)

We apologise for the late submitting our revised article.

Thank you very much for your kind assistance.

Wassalam,
Ari Asnani

-----Original message-----

From: ICMASure Unsoed <icmasure.unsoed@gmail.com>

Sent: Sunday 13th January 2019 23:57

To: Ari Asnani <ari.asnani@unsoed.ac.id>; asnani@gmail.com

Subject: ICMA-LPPM Review Complete

We are glad to inform you that we have received the external review results of your manuscript from well-known experts. Please check the reviewed manuscript and note the comments/suggestions/answers to questions of the reviewers.

As the proceedings will have to be published soon, we request you to

incorporate the reviewers' comments/suggestions/answers to questions in the manuscript as soon as possible, not later than 31 January 2019.

We look forward to hearing from you. Thank you very much for your kind attention and cooperation.

Regards,

The Committee of ICMA-SURE LPPM Unsoed 2018

**Revisi Dwi Utami Anjarwati et al.doc**

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Ari Asnani <asnani@gmail.com>

Sun, May 19, 2019 at 10:10 AM

To: Dwi Utami Anjarwati <dzikrarana@gmail.com>

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