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Volume 16 Issue 1, March 2022

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Food Safety Control Measures to Address Emerging Omicron SARS-CoV-2 Variant of Concern

Muhammad Shahbaz¹*, Muhammad Bilal² and Hafiz M.N. Igbal³

https://doi.org/10.22207/JPAM.16.1.24

Case Report | Open Access

Case Study of Patients with HIV during the COVID-19 Pandemic

Maria Olalla Garcia* 🗓, Franklin Escobar Lumitaxi and Miryan Guambuguete Llumitaxi

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Mini Review | Open Access

Efflux-Mediated bile Resistance in Gram-Positive Pathogens

Amjed Alsultan¹* and Dhama Alsallami²

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Osteoarthritis, Corticosteroids and Role of CYP Genes in COVID-19 Patients: A Mini Review

Usha Adiga¹, Sachidananda Adiga², Varashree Bolar Suryakanth³* and Desy Thayyil Menambath¹

https://doi.org/10.22207/JPAM.16.1.28

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Dania Hassan¹, Roshni Ravindran¹ and Ashfaque Hossain^{1,2}*

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sistance and virulence of pathogenic microbes through screening and evaluation of Indian medicinal plant derived herbal products/extracts and phytocompounds especially targeting Quorum sensing, biofilm and virulence of pathogenic microorganisms. His research work on agriculturally important microorganisms on diazotrophs, biofilm forming PGPR and Impact of wastewater on soil health has been well documented. His recent interest on Interdisciplinary microbiological works and drug-macromolecule interactions are gaining importance in academic world.

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Kuldeep Dhama is currently working as Principal Scientist in ICAR-Indian Veterinary Research Institute (IVRI), Izatnagar, India. With 22 years of research and teaching experience in the areas of microbiology, immunology and virology, he has developed several diagnostics, vaccines, immunomodulatory modules and hypothesis to counter infectious diseases of animals, poultry and public health concerns. He also has been awarded NAAS Associateship (National Academy of Agricultural Science, India). To his credit he has handled 20 research projects and guided 17 M.V.Sc. and P.D. scholars.

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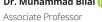
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RESEARCH ARTICLE

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The Production of Streptomyces W-5B Extract for Antibiofilm against Methicillin-resistant Staphylococcus aureus

Ari Asnani¹* , Azizah Purwanti¹, Windy Amalia Bakrudin¹ and Dwi Utami Anjarwati²

Abstract

The ability of Methicillin-resistant *Staphylococcus aureus* (MRSA) to form biofilms is one of the triggering factors for the emergence of MRSA resistance to antibiotics. Streptomyces W-5B has shown potency as an antibacterial producer against MRSA. However, the production of microbial bioactive compounds is strongly affected by the source of nutrients in the fermentation medium. Therefore, the objective of this study was to determine the optimal sources of carbon and nitrogen for the production of bioactive compounds with antibiofilm activities. The research method included cultivating Streptomyces W-5B, extract production, and variation of carbon (glucose, sucrose, starch) and nitrogen (casein, peptone, urea) sources for fermentation medium. Antibiofilm activities were measured based on inhibition of biofilm formation and biofilm degradation tests using the microtiter plate method with a crystal violet stain. The results showed that the highest inhibition of biofilm formation was $68.206 \pm 1.750\%$ after 12 days of incubation in a fermentation medium containing sucrose and urea. Meanwhile, the highest biofilm degradation was $73.023 \pm 1.972\%$ after nine days of incubation on a fermentation medium containing starch and urea. These findings indicated that Streptomyces W-5B has the potency to produce antibiofilm extract against MRSA.

Keywords: Antibiofilm, carbon source, MRSA, nitrogen source, Streptomyces

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INTRODUCTION

Methicillin-resistant Staphylococcus aureus (MRSA) is an S. aureus bacterium resistant to β-lactam antibiotics such as penicillin and its derivatives, namely methicillin, oxacillin, dicloxacillin, nafcillin, and cephalosporins.1 Penicillin-resistant strain of *S. aureus* produces a plasmid encoding a lactamase (penicillinase) capable of hydrolyzing the β-lactam ring of penicillin, causing antibiotic resistance. Currently, MRSA is one of the leading causes of infection in humans. Research conducted by Basanisi et al.² showed that of the 12.9% of samples infected with S. aureus, 8.3% were MRSA. The World Health Organization (WHO) already classifies MRSA as one of the priority pathogens that threaten human health due to antibiotic resistance.3 Resistance to MRSA increases due to the ability of MRSA to form biofilms.4

Biofilm is a matrix layer of extracellular polymeric substances (EPS) produced by bacteria. The EPS matrix generally consists of oligosaccharides, DNA, and proteins. 5 Bacteria with the ability to form biofilms can withstand stress and harsh host environments. Biofilms provide protection against antibiotics, thereby increasing resistance compared to bacteria in a planktonic state.4 The ability of MRSA to produce biofilms will inhibit the penetration of antibiotics, thus increasing resistance properties. Therefore, the exploration of bioactive compounds with antibiofilm properties is essential as an alternative therapy for MRSA infections. Antibiofilm compounds play a role in inhibiting the formation of biofilms and the degradation of biofilms that have been formed.

Bioactive compounds from actinobacteria, particularly the genus Streptomyces, have been reported as antibiofilm against MRSA. Streptorubin B extracted from Streptomyces sp. MC11024 belongs to a group of antibiotics capable of inhibiting and damaging the MRSA N315 biofilm.⁶ Ethyl acetate extract of Streptomyces sp. SBT343 is known to significantly inhibit the biofilm of several Staphylococcal species, including MRSA USA300.⁷ Bhakyashree & Kannabiran⁸ also reported that ethyl acetate extract of Streptomyces gancidius has antibacterial activity against MRSA strains ATCC 43300 and ATCC 700699.

Furthermore, Singh & Dubey9, reported

that *Streptomyces californicus* strain ADR1 had an antibiofilm activity that could inhibit and degrade biofilms of MRSA strains 562 and ATCC 43300. Asnani et al.¹⁰ succeeded in exploring *actinobacteria* from the mangrove area of Segara Anakan Cilacap, which has the potential as a source of antibacterial compounds, one of which is *Streptomyces* W-5B. Biosynthetic Gene Cluster (BGC) analysis of *Streptomyces* W-5B suggested eight BGCs that play a role in the biosynthesis of secondary metabolites.¹¹ The potency of *Streptomyces* W-5B encourages further research to optimize the production of antibiofilm compounds against MRSA.

Extracts produced by a microorganism will have optimum bioactivities if nutrient sources for microbial growth are selected accordingly. Carbon and nitrogen sources are the main components in a culture medium grown by microorganisms for cell growth processes and production of secondary metabolites, particularly in Streptomyces. 12 Carbon is the main component of all organic molecules in cells, whereas Nitrogen plays an essential role in preparing nucleic acids, amino acids, and enzymes.¹³ Hence, optimization of the fermentation process can be done by varying the composition of the media through simultaneous experiments to determine the interaction between variables. Therefore, this research aimed to optimize the production of antibiofilm extract from Streptomyces W-5B by varying carbon and nitrogen sources.

MATERIALS AND METHODS

The research was carried out from December 2020 to May 2021 at the University of Jenderal Soedirman, Purwokerto, Indonesia. *Streptomyces* W-5B was isolated from mangrove area in Segara Anakan, Cilacap, Indonesia. The species identification based on the 16S rRNA gene of *Streptomyces* W-5B showed 99,92% similarity with *Streptomyces cellulosae* strain NBRC 13027. The isolate MRSA 2983 was collected from a clinical specimen from the pus of a female patient in Prof. Dr. Margono Soekarjo Hospital, Banyumas regency, Indonesia. 14

Cultivation of Streptomyces W-5B

Cultivation of *Streptomyces* W-5B followed the procedure described by Asnani et al.¹⁰ *Streptomyces* W-5B was cultivated by

quadrant streak on Starch Casein Nitrate (SCN) agar medium consisting of starch, casein, KNO₃, K₂HPO₄, MgSO₄.7H₂O, NaCl, FeSO₄.7H₂O, and Bacto agar supplemented with nystatin. The culture was incubated for seven days at room temperature prior to being used.

A total of 10 plugs (6 mm in diameter) of *Streptomyces* W-5B culture were inoculated into 100 ml of liquid SCN medium [starch, casein, KNO₃, K₂HPO₄, MgSO₄.7H₂O, NaCl, and FeSO₄.7H₂O] supplemented with nystatin. The mixture was incubated using an orbital shaker at 90 rpm for eight days at room temperature. After incubation, the culture was used as an inoculum for the fermentation process.

Determination of the Optimal Incubation Time

The optimal incubation time was determined by varying the incubation time in the fermentation process. As much as 10% inoculum was inoculated in a liquid SCN medium. The culture was then incubated using an orbital shaker at 90 rpm with various incubation times 0, 3, 6, 9, 12, and 15 days. After each incubation, the cultures were centrifuged at 4000 rpm for 10 minutes at 4°C, then filtered to obtain Streptomyces W-5B extracts. The extracts obtained from various incubation times were tested against MRSA 2983 to evaluate extract potency to inhibit biofilm formation and degrade biofilm. The precipitate obtained was dried at 80°C for 1 hour for the dry weight. The production curve was made using the dry weight of the precipitate (g) and the percentage of inhibition of biofilm formation or biofilm degradation against the incubation time (days). The incubation time with the highest % inhibition and % degradation against MRSA 2983 was then used as the incubation time for the production of extracts with various carbon and nitrogen sources.

Determination of Carbon and Nitrogen Sources

The fermentation experiment used a complete randomized design with two factorials to evaluate the optimal carbon and nitrogen sources. The first factor was three different carbon sources (K1 = glucose, K2 = sucrose, K3 = starch), and the second factor was three different nitrogen sources (N1 = casein, N2 = peptone, N3 = urea). All treatments were replicated three times, so there were $3 \times 3 \times 3 = 27$ experimental units.

Up to 10% inoculum was inoculated in a

liquid SCN medium with a combination of carbon and nitrogen sources in a fermentation medium. Cultures were incubated using an orbital shaker at 90 rpm at room temperature at the optimal incubation time. Each extract obtained was tested for the activity to inhibit biofilm formation and to degrade biofilm.

Inhibition of the biofilm formation test

The ability of the extract to inhibit biofilm formation was analyzed using a semi-quantitative microtiter plate assay following the procedure described by Dinda et al. ¹⁴ The MRSA 2983 inoculated into Brain Heart Infusion (BHI) broth supplemented with 1% glucose (BHI-Glu). The culture was incubated at 37°C for 24 hours, adjusted to 0.5 McFarland standard, and diluted with BHI-Glu medium in a 1:100 ratio to become an MRSA subculture.

The inhibition of biofilm formation was carried out by adding 10 μ L of MRSA subculture and 100 μ L of extract into 100 μ L of BHI-Glu medium in each well in a 96-well microplate. The mixture was incubated at 37°C for 24 hours. After incubation, planktonic cells were carefully removed, and the microplate was rinsed with 300 μ L of PBS twice and stained with 50 μ L of 1% (w/v) crystal violet solution. The absorbance value was measured at 620 nm, and the percentage of inhibition of biofilm formation was calculated by the following formula:

% Inhibition=
$$\frac{\text{(OD control-OD test)}}{\text{(OD control)}} X 100\%$$

The minimum biofilm inhibition concentration of 50% (MBIC-50) was defined as extract concentration at which a 50% decrease in absorbance readings was detected in comparison to the absorbance of control biofilms formed in the absence of extract. MBIC-50 was determined by calculating the cut-off point at 50% inhibition by the following formula. 15:

MBIC-50=(OD control-OD Blank) X
$$\frac{50}{100}$$

Biofilm Degradation Test

The ability of the extract to degrade biofilm was also analysed with a semi-quantitative microtiter plate biofilm assay.14 Up to 10 μ L of MRSA subculture was added into 100 μ L of BHI-Glu medium in each well on a 96-well microplate, and the mixture was incubated at 37°C for 24 hours. After incubation, the planktonic cells were

carefully removed, and each well was added 100 μL of extract. The mixture was further incubated at 37°C for 24 h to expose the extract with the biofilm formed. After incubation, the microplate was washed similarly with the procedure mentioned above, and the absorbance value was measured at 620 nm. The percentage of biofilm degradation was calculated by the following formula:

% Degradation=
$$\frac{\text{(OD control-OD test)}}{\text{(OD control)}}$$
 X 100%

Likewise, the minimum biofilm reduction concentration of 50% (MBRC-50) was defined as extract concentration at which a 50% decrease in absorbance readings were detected in comparison to the absorbance of control biofilms formed in the absence of extract. MBRC-50 was determined by calculating the cut-off point at 50% reduction by the following formula.¹⁵:

MBRC-50=(OD control-OD Blank) X
$$\frac{50}{100}$$

Data Analysis

All data were analyzed using analysis of variance (ANOVA) at a confidence level of 95%. The results of the ANOVA were significantly different, followed by Duncan's test.

RESULTS

Streptomyces W-5B is a filamentous bacterium with brown pigmentation to medium, yellow substrate mycelium, and white aerial mycelium (Fig. 1). The colony is circular and has a powdery surface. Streptomyces W-5B has been reported to have antibacterial activity against MRSA 2983.¹¹

The Optimum Incubation Time

The production curve depicts the relationship between incubation time and Streptomyces W-5B extract production. The absorbance value represented the inhibitory activity, and biofilm degradation read at 620 nm. Then, the absorbance value was used to calculate the percentage of inhibition and degradation of the biofilm. The specific incubation time was determined to produce an extract with the highest activity, while the bacterial precipitate measured as dry weight was used to analyze the bacterial growth. The research results showed that the optimum incubation time to produce *Streptomyces* W-5B extract with the highest inhibition activity was 12 days, with the percentage of biofilm inhibition being 55,456 ± 3,306% (Fig. 2). This result followed the microbial growth, which showed optimum growth on 12-days based on the dry weight (0.0823 g). The percentage of biofilm inhibition at the incubation time of 12 days also had a value greater than the cut-off point of MBIC-50. These results indicated that an incubation time of 12 days would produce an extract that could inhibit biofilm formation by more than 50%.

The one-way ANOVA results showed a significant effect (Sig. <0.05) between the incubation time and the percentage of inhibition of biofilm formation. Further tests using Duncan's test showed that treatment with an incubation time of 12 days was significantly different from other treatments $(0,6,9,15\,\text{days})$ with the highest percentage of biofilm inhibition $(55,456\pm3,306\%)$.



Fig. 1. Streptomyces W-5B cultivated in starch casein nitrate agar, (a) Top view, (b) bottom view

The percentage of biofilm degradation by *Streptomyces* W-5B extract increased from day 0 to the optimal incubation time on day 9 with a percentage value of biofilm degradation of $56.146 \pm 8.784\%$ (Fig. 3). The biofilm degradation that crossed the MBRC-50 boundary line was the extract produced on the 9th day, which indicated that extract produced with an incubation time of 9 days could degrade 50% of the biofilm.

The one-way ANOVA results showed a significant effect (Sig. <0.05) between the incubation time and the percentage of biofilm degradation. Further tests using Duncan's test concluded that variations of incubation time show significantly different effects. However, the incubation time treatment on the 9th day showed no significant effect on the 3rd and 6th days. The optimal incubation time was on the ninth day,

with the highest percentage of biofilm degradation being $56.146 \pm 8.784\%$.

Based on these results, the fermentation process with various carbon and nitrogen sources used 12 days of incubation to produce an extract with inhibitory activity. In comparison, nine days of incubation was used to produce an extract with biofilm degradation activity.

The Optimum Carbon and Nitrogen Sources

Variations of carbon and nitrogen sources determined the best combination of carbon and nitrogen sources in fermentation medium to produce an extract with the highest inhibition biofilm formation and biofilm degradation activities. Based on the research results (Fig. 4), the highest percentage of inhibition of biofilm formation (68.206 \pm 1.750%) was produced from the K2N3 fermentation medium (sucrose-urea).

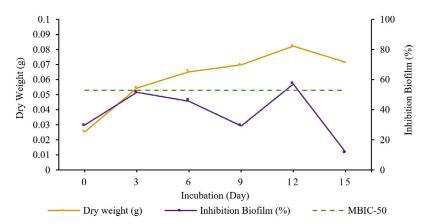


Fig. 2. Production of extract from Streptomyces W-5B with inhibition activity.

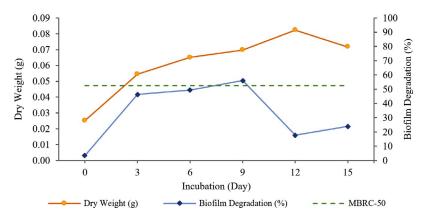


Fig. 3. Production of extract from Streptomyces W-5B with biofilm degradation activity.

The percentage of inhibition in the K2N3 medium was higher than the cut-off point of MBIC-50, which means that the extract produced in the K2N3 medium could inhibit biofilm formation by more than 50%. Other extracts with a higher value than the cut-off point of MBIC-50 were K1N2 (glucose-peptone), K1N3 (glucose-urea), K2N2 (sucrose-peptone), K3N1 (starch-casein), and K3N2 (starch-peptone).

The results of the two-way ANOVA showed a significant effect (Sig. <0.05) between the interaction of carbon and nitrogen sources on the percentage of inhibition of biofilm formation. Further tests using Duncan's test showed that the treatment with sucrose had a significantly different effect on other treatments.

The results from the biofilm degradation test indicated that the highest percentage of biofilm degradation (73.023 \pm 1.972%) was obtained from the extract produced by fermentation

medium K3N3 (starch-urea). The MBRC-50 value determines the minimum concentration capable of degrading 50% of the biofilm. ⁷ Interestingly, all extracts had a higher value than MBRC-50, which means that they can degrade more than 50% of the biofilm formed. Still, the best carbon-nitrogen combination was K3N3, with the highest biofilm degradation activity (Fig. 5).

The two-way ANOVA results showed a significant effect (Sig. <0.05) between the interaction of carbon and nitrogen sources on the percentage of biofilm degradation. Further tests using Duncan's test concluded that the percentage of biofilm degradation was significantly different with a combination of other carbon and nitrogen sources. The best combination for fermentation medium was K3N3 (starch-urea) which produced the highest (73.023 \pm 1.972%) biofilm degradation activity.

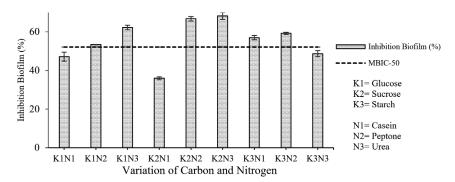


Fig. 4. Correlation of combined carbon and nitrogen sources in fermentation medium with the percentage of inhibition of biofilm formation.

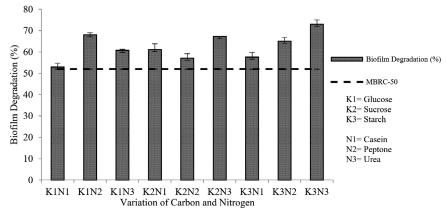


Fig. 5. Correlation of combined carbon and nitrogen sources in fermentation medium with the percentage of biofilm degradation

DISCUSSION

Streptomyces W-5B has been identified as Streptomyces cellulosae strain NBRC 13027. It has been reported to have antibacterial activity against MRSA 2983 with an inhibitory index of 0.47 mm and contained biosynthetic gene clusters (BGC) that have antibacterial potential. According to Memariani et al., 6 biofilm inhibition can occur at a lower concentration than the concentration to kill planktonic bacteria so that antibacterial compounds can also be used to inhibit biofilm formation. Thus, this research further analyzed the potency of Streptomyces W-5B to produce antibiofilm extract against MRSA.

Streptomyces W-5B was cultivated on SCN medium, which is a suitable medium for the growth of actinobacteria. The screening and isolation of 36 actinobacterial isolates using SCN agar were successfully carried out by Rajaram et al.¹⁷ Recently, SCN agar was also used to isolate antibiotic-producing bacteria (APB) from river sediments of Bangladesh. The 16S rDNA sequencing analysis revealed APB as Streptomyces genus.¹⁸ Actinobacteria grown on SCN agar medium showed fast growth and were able to produce secondary metabolites. Vellingiri et al.¹⁹ reported that SCN medium has been effective for Streptomyces hygroscopicus AVS7 to produce bioactive metabolites.

The high production of bioactive compounds was correlated with the microbial growth phase during incubation. Ryandini et al.20 reported that incubation time affects the production of bioactive compounds from actinobacteria. However, different bacteria might have different optimal incubation times to produce extracts with antibiofilm activity. Bakkiyaraj & Pandian,²¹ have reported that Streptomyces akiyoshiensis CAA-3 produced an antibiofilm extract against S. aureus ATCC 11632 and MRSA ATCC 33591 with an incubation time of six days. Lee et al.²² reported that Streptomyces parvulus produced an antibiofilm compound against MRSA ATCC 33591 with an incubation time of seven days. In general, the production of bioactive compounds occurs in the stationary phase or at the end of the exponential phase until the end of the microbial incubation period.23 Similarly, in this research, the extract with inhibition of biofilm formation activity occurred at the beginning of the stationary phase.

In contrast, the extract with biofilm degradation activity occurred exponentially. This result suggested biofilm degradation might correlate with the hydrolytic enzyme produced during the exponential phase. Indeed, Millenbaugh et al.²⁴ stated that biofilm degradation activity is related to the enzymatic activity produced by bacteria to reduce biofilms.

Antibiofilm compounds inhibit biofilm formation by preventing the adhesion of planktonic bacteria to the surface or host tissues to form biofilms. The mechanism of inhibition occurs at the beginning of the reversible attachment of planktonic bacteria to the surface before the biofilm matrix maturation occurs. Inhibition takes place because biofilm-producing bacteria are not yet protected by a mature biofilm matrix; hence they are still very susceptible to antibiotic compounds.²⁵

Carbon and nitrogen are macronutrients that affect the production of microbial bioactive compounds.¹³ In this research, the combination of sucrose and urea produces extract with the highest activity to inhibit biofilm formation. Sucrose is a better carbon source than other carbon sources for producing secondary metabolites that function as antibacterial.²⁶ The presence of sucrose in the fermentation medium can stimulate the activity of the phenylalanine ammonia lyase (PAL) enzyme involved in producing secondary metabolites. This PAL enzyme plays a vital role in microbial defense mechanisms against environmental stresses.27 The addition of sucrose can also suppress the salt content in the fermentation medium. The lack of salt will activate genes that produce secondary metabolites such as compounds from the polyketide group.²⁸ Urea acts as a simple nitrogen source that is easily used by actinobacteria so that actinobacteria can reach the stationary phase more quickly to produce secondary metabolite compounds that play a role in inhibiting the formation of MRSA biofilms.

Based on the research results, the combination of starch and urea produced the extract with the highest biofilm degradation activity. The mechanism of biofilm degradation is carried out by weakening hydrogen bonds, which are considered necessary for the stability of the EPS matrix.²⁹ Starch, a source of carbon in the form of polysaccharides, can help bacteria produce

organic molecules in cells, including anti-adhesion polysaccharides or antibiofilm polysaccharides. This antibiofilm polysaccharide can act as a biofilm degrader by damaging cell-to-cell interactions in the biofilm matrix.³⁰ Suzuki et al.⁶ reported that *Streptomyces* grown in media containing starch could produce compounds with degradation activity against *S. aureus* biofilms. Nandhagopal et al.³¹ have used starch as a carbon source to produce biofilm-degrading compounds against MRSA.

Urea as a nitrogen source will help bacteria synthesize essential components such as amino acids and proteins. These components play a significant role in producing bioactive metabolites such as enzymes and other metabolites that degrade biofilms. ³² Urea has been used in bacterial growth medium to produce an extracellular enzyme complex that can degrade *S. aureus* biofilms up to 80%. ³³ The enzymes effectively degrade biofilms because they lyse cells and tissues to damage the biofilm matrix.

CONCLUSION

Our results conclude that Streptomyces W-5B can produce MRSA antibiofilm compounds. The antibiofilm activity consists of antibacterial, inhibitor of biofilm formation, and biofilm degrading. Actinobacteria are known as antibiotic-producing bacteria. Many commercial antibiotics include important antimicrobials such as β-lactams, rifamycin, aminoglycosides, macrolides, tetracyclines, erythromycin, vancomycin, and streptomycin are derived from the secondary metabolism of actinobacteria. In the class Actinobacteria, the genus Streptomyces is the primary producer of various secondary metabolites and many antibiotics. One of the determining factors for producing primary and secondary metabolites is the fermentation media formulation. The fermentation medium can also act as an induction to obtain the desired metabolite optimally. In the future, it is necessary to identify and characterize secondary metabolites that specifically function as MRSA antibiofilm.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

AUTHORS' CONTRIBUTION

AA supervised the entire work. DUA led the antibiofilm work. AP and WAB performed all experiments. AA wrote and revised the manuscript. DUA reviewed the manuscript. AA and DUA approved the manuscript for publication.

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DATA AVAILABILITY

All datasets generated and analyzed during this research are included in the manuscript.

ETHICS STATEMENT

This article does not contain any studies with human participants or animals performed by any of the authors.

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JPAM – Article for Publication. Ref No.: JPAM/7441/2021

2 messages

Journal of Pure and Applied Microbiology <editor@microbiologyjournal.org> To: Ari Asnani <ari.asnani@unsoed.ac.id>

Sat, Nov 20, 2021 at 7:14 PM

Dear Dr. Ari.

We have received your article and have done a similarity check screening of your article for plagiarism, which is found to be 14%. We will let you know the review status in 8-10 weeks' time.

Kindly add the following statements in your article and provide us all author's Orcid ID's. As we have to incorporate these statements and Orcid ID's.

- Acknowledgments
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- Author's Contribution
- Funding(Clearly mention grant number, if any)
- Data Availability
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Kindly make all the necessary changes and submit your article again with your article reference number JPAM/7441/2021.

Please find a few examples of the following statements:

Acknowledgments

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We would like to express our heartfelt thanks to Professor XYZ for providing assistance on methods. Thanks to XYZ Co., Ltd. for providing assistance on bioinformatics analysis. The authors are grateful to Dr. XYZ for English editing of the manuscript. We would like to thank the Central Laboratory of Public Health (Ministry of Health- MSPyBS) and the National TB Program for the collaboration during the recruitment process and provided surveillance data.

Else if no acknowledgement is required write: None

Conflict of Interest (if no conflict of interest among authors)

The authors declare that there is no conflict of interest.

Authors' Contributions

(For Example)

All authors designed the experiments. Author Initials, Author Initials and Author Initials performed the experiments. Author Initials, Author Initials and Author Initials and Author Initials wrote the manuscript. All authors read and approved the manuscript.

OR

AB (Author Initials) drafted the manuscript, compiled information from the literature, and designed the figures and tables. MP (Author Initials) drafted the manuscript and gathered information from the literature. SE (Author Initials) supervised and reviewed the manuscript. CM (Author Initials) supervised and reviewed the manuscript and designed the figures and tables.

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.
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This study was carried out in accordance with the recommendations of NIH guidelines for animal use, Animal Care and
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This article does not contain any studies with human participants or animals performed by any of the authors.
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On Thu, 18 Nov 2021 at 15:41, Journal of Pure and Applied Microbiology <editor@microbiologyjournal.org> wrote:

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New Article

Name of Corresponding Author

First Name	Middle Name	Last Name	ORCID ID
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Name of Co-Author(s)

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Message

Dear Editor in Chief

Journal of Pure and Applied Microbiology

We would like to submit our article entitle The Production of Streptomyces W-5B Extract for Antibiofilm against Methicillin-resistant Staphylococcus aureus.

Thank you very much for your attention and kind consideration.

Your sincerely, Ari Asnani

• The authors confirm that the article including text, figures and tables, has neither been published before, not it is under consideration elsewhere and also all authors have made substantial contribution to this article and have given final approval for this version to be submitted.

Dear Dr. M.N. Khan The Editor Journal of Pure and Applied Microbiology (JPAM)

We have submitted our revised manuscript with reference number JPAM/7441/2021 in the system. We have already incorporated all authors' ORCID IDs and the required statements in the manuscript.

Thank you very much for your kind attention and consideration.

Best Regards, Ari Asnani, Ph.D.

[Quoted text hidden]

Ari Asnani, Ph.D. Jurusan Kimia - Fakultas MIPA Universitas Jenderal Soedirman Purwokerto, Jawa Tengah, Indonesia



JPAM/7441/2021 (Reviewer Comments)

3 messages

Journal of Pure and Applied Microbiology <editor@microbiologyjournal.org> To: Ari Asnani <ari.asnani@unsoed.ac.id>

Thu, Dec 16, 2021 at 2:16 PM

Dear Dr. Asnani,

In reference to your article reviewed, please find the reviewer's comments, make sure to do the corrections as suggested by the reviewer and revert back to us at your earliest convenience.

Note: kindly highlight the corrections in yellow colour or show the track changes. So that reviewer can follow the corrections.

Reviewer Comments:

Line number	Reviewer's Comments/Suggestions
77	How were the Streptomyces W-5B and clinical isolate MRSA 2983 identified?
128	Crystal violet concentration should be added
158	Information about Streptomyces W-5B should be in the material methods section not in the results
210	K3N1 (amylum-casein), and K3N2 (amylum-peptone) should be starch, not amylum
	The results section may be shorter, the comments here can be used in the discussion section.

Kindly acknowledge the email to confirm safe delivery.

Regards

Dr. M.N. Khan

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On Mon, 22 Nov 2021 at 09:23, Journal of Pure and Applied Microbiology <editor@microbiologyjournal.org> wrote:

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Message

Dear Dr. M.N. Khan

The Editor

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We would like to submit our revised manuscript.

We have already incorporated all authors' ORCID IDs and the required statements in the manuscript.

Thank you very much for your kind attention and consideration.

Best Regards, Ari Asnani, Ph.D.

> • The authors confirm that the article including text, figures and tables, has neither been published before, not it is under consideration elsewhere and also all authors have made substantial contribution to this article and have given final approval for this version to be submitted.



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Ari Asnani <ari.asnani@unsoed.ac.id>

To: Journal of Pure and Applied Microbiology <editor@microbiologyjournal.org>

Thu, Dec 16, 2021 at 2:55 PM

Dear Dr. M.N. Khan

The Editor

Journal of Pure and Applied Microbiology (JPAM)

We have received the email regarding reviewers comments.

We will do the corrections as suggested by the reviewer.

Thank you very much,

Yours sincerely,

Ari Asnani

[Quoted text hidden]

Ari Asnani, Ph.D.

Jurusan Kimia - Fakultas MIPA Universitas Jenderal Soedirman

Purwokerto, Jawa Tengah, Indonesia

Ari Asnani <ari.asnani@unsoed.ac.id>

To: Journal of Pure and Applied Microbiology <editor@microbiologyjournal.org>

Sat, Dec 25, 2021 at 4:09 AM

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On Thu, Dec 16, 2021 at 2:17 PM Journal of Pure and Applied Microbiology <editor@microbiologyjournal.org> wrote:

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77	How were the Streptomyces W-5B and clinical isolate MRSA 2983 identified?
128	Crystal violet concentration should be added
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210	K3N1 (amylum-casein), and K3N2 (amylum-peptone) should be starch, not amylum
	The results section may be shorter, the comments here can be used in the discussion section.

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Re: New submission from ari.asnani@unsoed.ac.id

2 messages

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Mon, Dec 27, 2021 at 2:16 PM

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We would like to submit our revised manuscript following the suggestions by the reviewer. Thank you very much for your kind attention and consideration.

Best Regards, Ari Asnani, Ph.D.

The authors confirm that the article including text, figures and tables, has neither been published before, not it is under
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Journal of Pure and Applied Microbiology <editor@microbiologyjournal.org>
To: Ari Asnani <ari.asnani@unsoed.ac.id>

Tue, Jan 4, 2022 at 1:43 PM

Dear Dr. Ari,

We are pleased to inform you that your article reference number: JPAM/7441/2021, entitled "The Production of Streptomyces W-5B Extract for Antibiofilm against Methicillin-resistant Staphylococcus aureus" is accepted for publication in the Journal of Pure and Applied Microbiology in our forthcoming issue. Soon, you will receive the acceptance letter from our publisher email address micro_drkhan@yahoo.com; once received, you are requested to kindly pay the APCs and share the transaction receipt for further processing of your article.

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